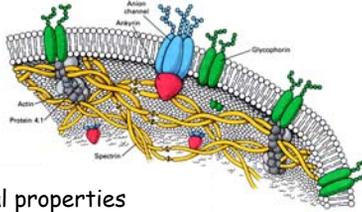
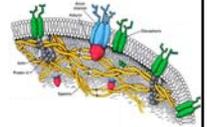


Biological Membranes



- Structure
- Function
- Composition
- Physicochemical properties
- Self-assembly
- Molecular models

Lipid Membranes



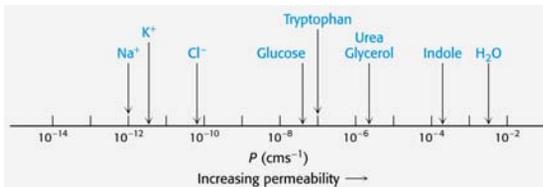
- Receptors, detecting the signals from outside:
 - Light
 - Odorant
 - Taste
 - Chemicals
 - Hormones
 - Neurotransmitters
 - Drugs
- Channels, gates and pumps
- Electric/chemical potential
 - Neurophysiology
 - Energy
- Energy transduction:
 - Photosynthesis
 - Oxidative phosphorylation

A highly selective permeability barrier

Internal membranes for organelles

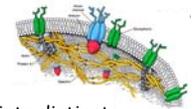
Bilayer Permeability

- Low permeability to charged and polar substances
- **Water** is an exception: small size, lack of charge, and its high concentration
- **Shedding solvation shells for ions is very unlikely**



Common Features of Biological Membranes

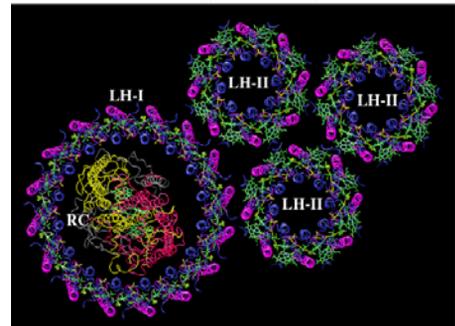
- Sheet-like structure
- TWO-molecule thick (60-100Å)
- **Lipids, Proteins, and carbohydrates**
- Lipids form the barrier. Proteins mediate distinct functions.
- Non-covalent assemblies (self-assembly, protein-lipid interaction)
- **Asymmetric (always)**
- Fluid structures: 2-dimensional solution of oriented lipids and proteins
- Electrically polarized (**inside negative** ~-60mV)
- Spontaneously forming in water
- Protein/lipid ratio = 1/4 - 4/1
- Carbohydrate moieties are always outside the cell



Protein/Lipid ratio

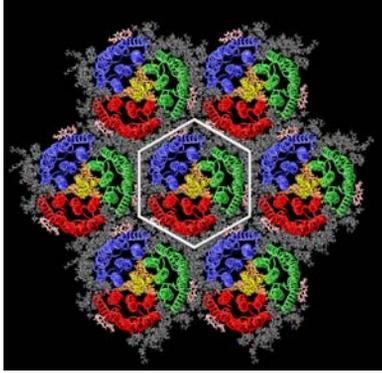
- Pure lipid: insulation (neuronal cells)
- Other membranes: on average 50%
- Energy transduction membranes (75%)
 - Internal membranes of mitochondria and chloroplast
 - Purple membrane of halobacteria
- Different functions = different protein composition

Protein / Lipid Composition



Light harvesting complex of purple bacteria

Protein / Lipid Composition



The purple membrane of halobacteria

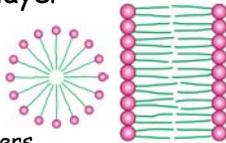
General features of Lipids

- Small molecules
- Amphipathic (amphiphilic)
Hydrophobic/hydrophilic moieties
- Spontaneously form vesicles, micelles, and bilayers in aqueous solution



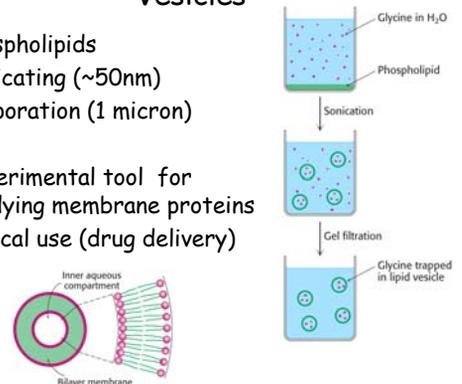
Micelle / Bilayer

- Fatty acids (one tail)
- Phospholipids (two tails)
- Micelle max 20 nm
- Bilayer up to millimeters
- Self-assembly process
- Hydrophobic interaction is the driving force (also in protein folding and in DNA stacking)
- Extensive; tendency to close on themselves; self-sealing (a hole is unfavorable)



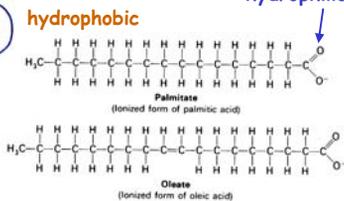
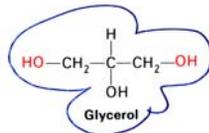
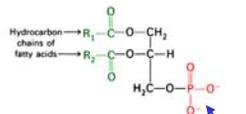
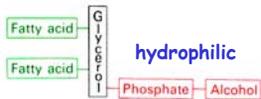
Vesicles

- Phospholipids
- Sonicating (~50nm)
- Evaporation (1 micron)
- Experimental tool for studying membrane proteins
- Clinical use (drug delivery)

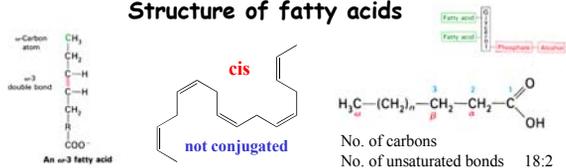


Phospholipids

hydrophobic



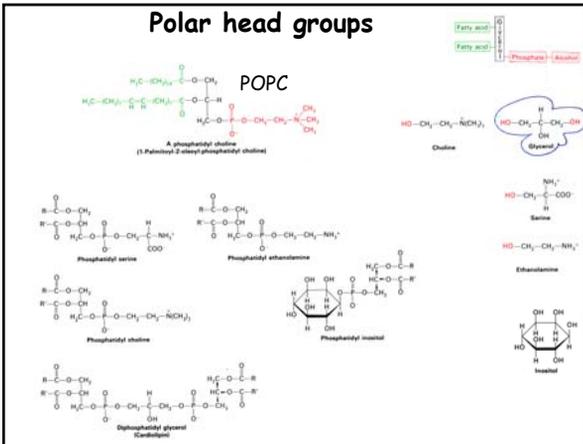
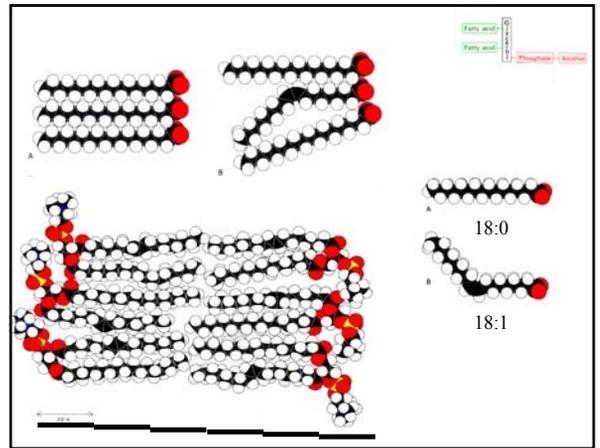
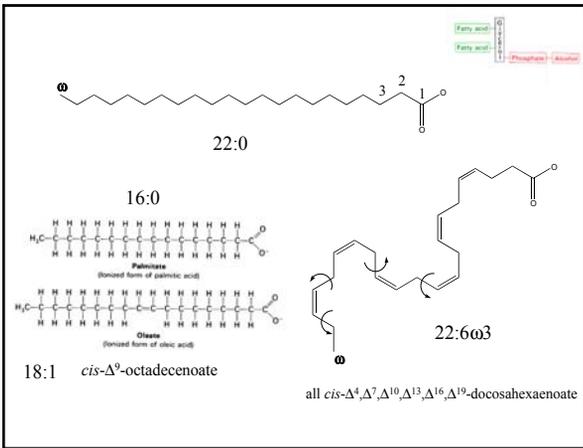
Structure of fatty acids



$H_2C-(CH_2)_n-CH_2-CH_2-C(=O)OH$
No. of carbons
No. of unsaturated bonds 18:2

TABLE 12.1 Some naturally occurring fatty acids in animals

Number of carbons	Number of double bonds	Common name	Systematic name	Formula
12	0	Laurate	n-Dodecanoate	$CH_3(CH_2)_{10}COO^-$
14	0	Myristate	n-Tetradecanoate	$CH_3(CH_2)_{12}COO^-$
16	0	Palmitate	n-Hexadecanoate	$CH_3(CH_2)_{14}COO^-$
18	0	Stearate	n-Octadecanoate	$CH_3(CH_2)_{16}COO^-$
20	0	Arachidate	n-Eicosanoate	$CH_3(CH_2)_{18}COO^-$
22	0	Behenate	n-Docosanoate	$CH_3(CH_2)_{20}COO^-$
24	0	Lignocerate	n-Tetracosanoate	$CH_3(CH_2)_{22}COO^-$
16	1	Palmitoleate	cis- Δ^7 -Hexadecenoate	$CH_3(CH_2)_5CH=CH(CH_2)_9COO^-$
18	1	Oleate	cis- Δ^9 -Octadecenoate	$CH_3(CH_2)_3CH=CH(CH_2)_9COO^-$
18	2	Linolate	cis,cis- Δ^9,Δ^{11} -Octadecadienoate	$CH_3(CH_2)_3CH=CHCH_2CH=CH(CH_2)_4COO^-$
18	3	Linolenate	all-cis- $\Delta^9,\Delta^{11},\Delta^{13}$ -Octadecatrienoate	$CH_3CH_2CH=CHCH_2CH=CHCH_2COO^-$
20	4	Arachidonate	all-cis- $\Delta^5,\Delta^8,\Delta^{11},\Delta^{14}$ -Eicosatetraenoate	$CH_3CH_2CH=CHCH_2CH=CHCH_2COO^-$



Diffusion in Membrane

Einstein relation for diffusion:

$$6Dt = \langle |\mathbf{r}_i(t) - \mathbf{r}_i(0)|^2 \rangle$$

$$|\mathbf{r}_i(t) - \mathbf{r}_i(0)| = S$$

$6Dt = \langle S^2 \rangle$ in a 3 dimensional space
 $4Dt = \langle S^2 \rangle$ in 2 dimensions (in-plane diffusion in a membrane)
 $2Dt = \langle S^2 \rangle$ in 1 dimension

Lipid Diffusion in Membrane

Rapid

Lateral diffusion

$$D = 1 \mu\text{m}^2 \cdot \text{s}^{-1}$$

50 Å in $\sim 2.5 \times 10^{-5}$ s

~9 orders of magnitude difference

Very slow

Transverse diffusion (flip-flop)

Once in several hours!

$$(10^4 \text{ s})$$

$D_{\text{lip}} = 10^{-8} \text{ cm}^2 \cdot \text{s}^{-1}$
 $D_{\text{wat}} = 2.5 \times 10^{-5} \text{ cm}^2 \cdot \text{s}^{-1}$

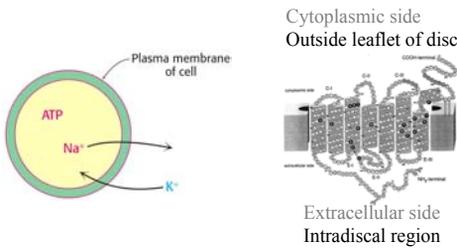
Fluid Mosaic Model of Membrane

Lateral Diffusion Allowed

Flip-flap Forbidden

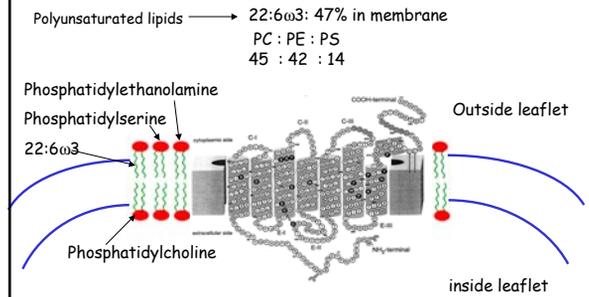
Ensuring the conservation of membrane asymmetric structure

Importance of Asymmetry

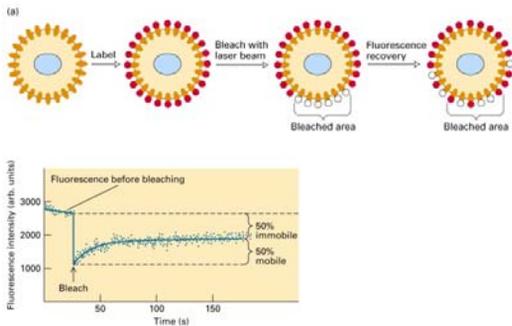


Apart from some passive transport mechanisms, all membrane proteins function in a directed way, and their correct insertion in the cell membrane is essential for their biological function.

Highly asymmetric and inhomogeneous lipid composition of membrane

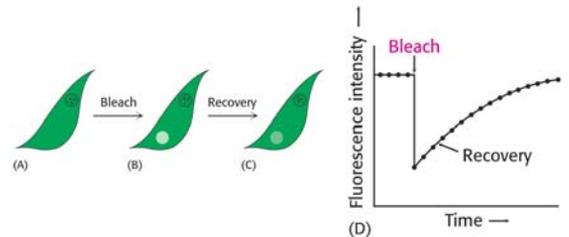


Fluorescence recovery after photobleaching (FRAP)

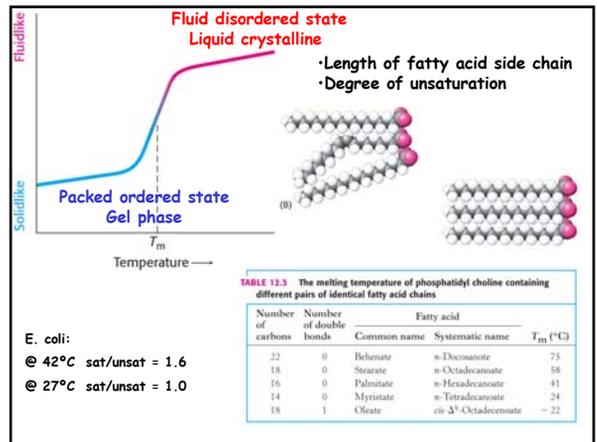


Lipid Diffusion in Membrane

- FRAP - fluorescent recovery after photobleaching (Albert's movie)



"FRAP experiment movie" The Molecular Biology of the Cell



A Brief Introduction to Molecular Dynamics Simulations

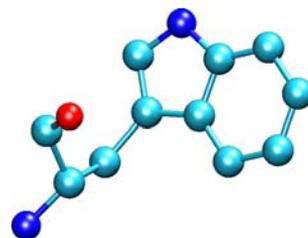
Macroscopic properties are often determined by atomic-level behavior.

Quantitative and/or qualitative information about macroscopic behavior of macromolecules can be obtained from simulation of a system at atomistic level.

Molecular dynamics simulations calculate the motion of the atoms in a molecular assembly using Newtonian dynamics to determine the net force and acceleration experienced by each atom. Each atom i at position r_i , is treated as a point with a mass m_i and a fixed charge q_i .

What is the Force Field?

In molecular dynamics a molecule is described as a series of charged points (atoms) linked by springs (bonds).



To describe the time evolution of bond lengths, bond angles and torsions, also the non-bonding van der Waals and electrostatic interactions between atoms, one uses a **force field**.

The **force field** is a collection of equations and associated constants designed to reproduce molecular geometry and selected properties of tested structures.

Energy Terms Described in the CHARMM Force Field

Bond



Angle



Dihedral



Improper



Energy Functions

$$U(\vec{R}) = \underbrace{\sum_{bonds} k_i^{bond} (r_i - r_0)^2}_{U_{bond}} + \underbrace{\sum_{angles} k_i^{angle} (\theta_i - \theta_0)^2}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihedral} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{dihedral}} + \underbrace{\sum_i \sum_{j \neq i} 4 \epsilon_{ij} \left[\left(\frac{\sigma_{ij}}{r_{ij}} \right)^{12} - \left(\frac{\sigma_{ij}}{r_{ij}} \right)^6 \right]}_{U_{nonbond}} + \sum_i \sum_{j \neq i} \frac{q_i q_j}{\epsilon r_{ij}}$$

U_{bond} = oscillations about the equilibrium bond length

U_{angle} = oscillations of 3 atoms about an equilibrium bond angle

$U_{dihedral}$ = torsional rotation of 4 atoms about a central bond

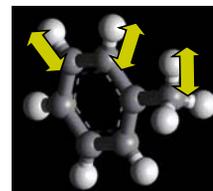
$U_{nonbond}$ = non-bonded energy terms (electrostatics and Lenard-Jones)

Time Scale of Biological Events

Motion	Time Scale (sec)
Bond stretching	10^{-14} to 10^{-13}
Elastic vibrations	10^{-12} to 10^{-11}
Rotations of surface sidechains	10^{-11} to 10^{-10}
Hinge bending	10^{-11} to 10^{-7}
Rotation of buried side chains	10^{-4} to 1 sec
Allosteric transitions	10^{-5} to 1 sec
Local denaturations	10^{-5} to 10 sec

The 1 fs Time Step Limit

- Dynamics simulations are limited by the highest frequency vibration.
- Ideally the timestep should be 1/10 of the period of the highest frequency vibration.
- X-H bond stretching ($\sim 10^{-14}$ s) is the fastest mode.



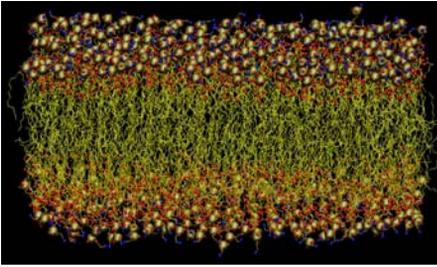
SPEED LIMIT

1 fs

Technical difficulties in Simulations of Biological Membranes

- **Time scale**
- Inhomogeneity of biological membranes

60 x 60 Å
Pure POPE
5 ns
~100,000
atoms



Coarse grain modeling of lipids

by: J. Siewert-Jan Marrink and
Prof. Dr. Alan E. Mark, University of
Groningen, The Netherlands

150 particles



9 particles!

