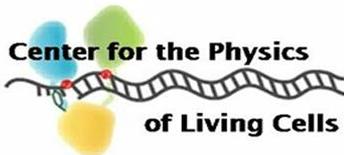


# Structure of the HIV-1 Capsid Assembly by a hybrid approach

Juan R. Perilla

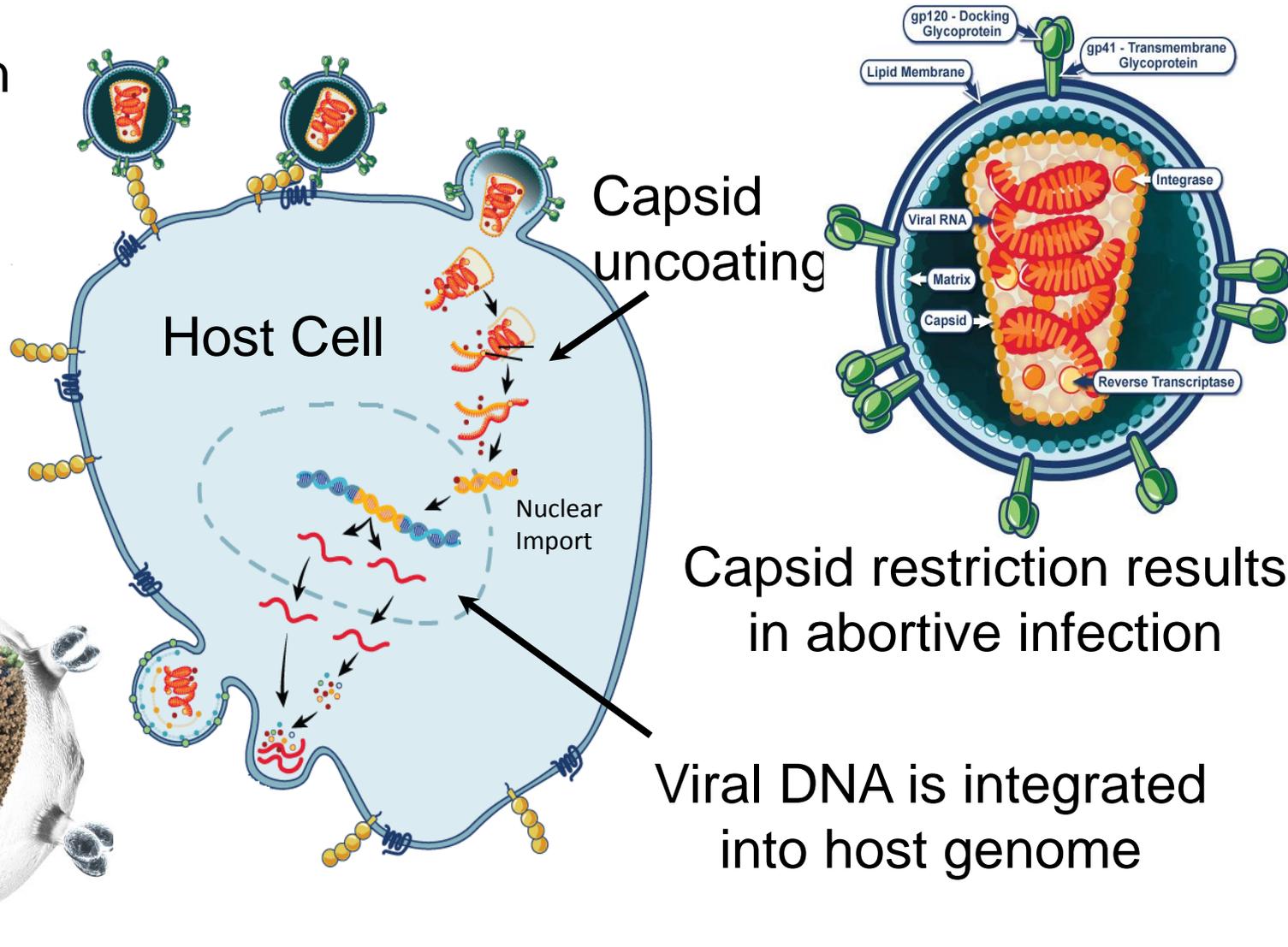
[juan@ks.uiuc.edu](mailto:juan@ks.uiuc.edu)

Theoretical and Computational Biophysics Group  
University of Illinois at Urbana-Champaign



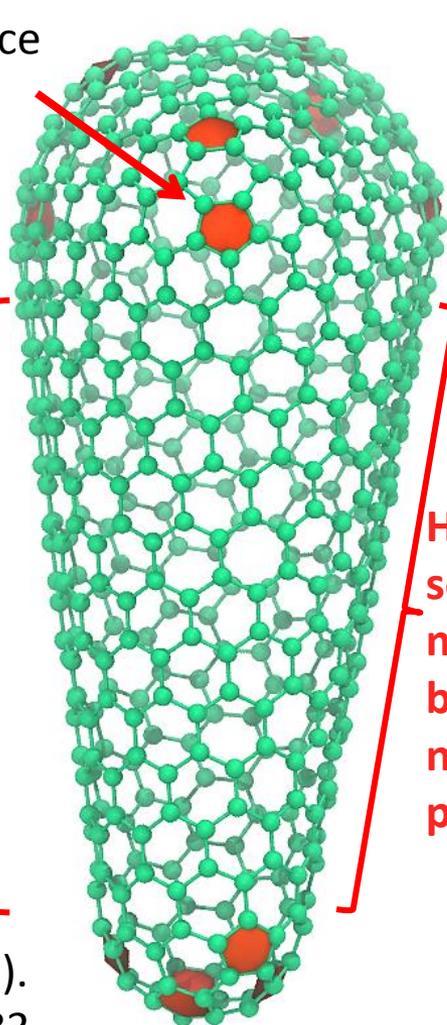
# HIV infective cycle

Virion



# Structure of the HIV capsid

Pentamers introduce sharp declinations

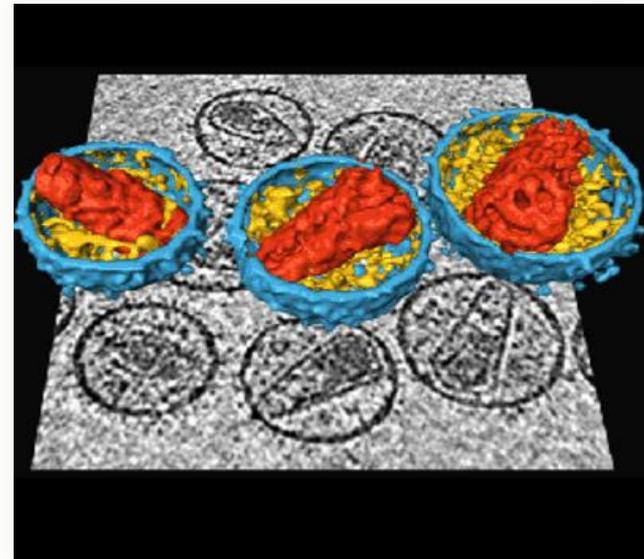
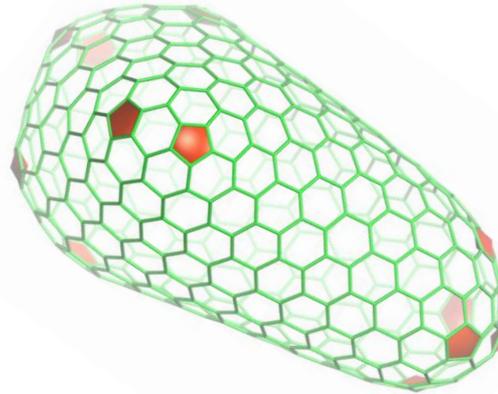


Continuously changing curvature in the hexagonal lattice

Highly schematic model; beads are not proteins!

Ganser, B. K. (1999). *Science*, 283, 80–83

Capped fullerene cone



Briggs, J. et al. (2006). *Structure*, 14, 15–20

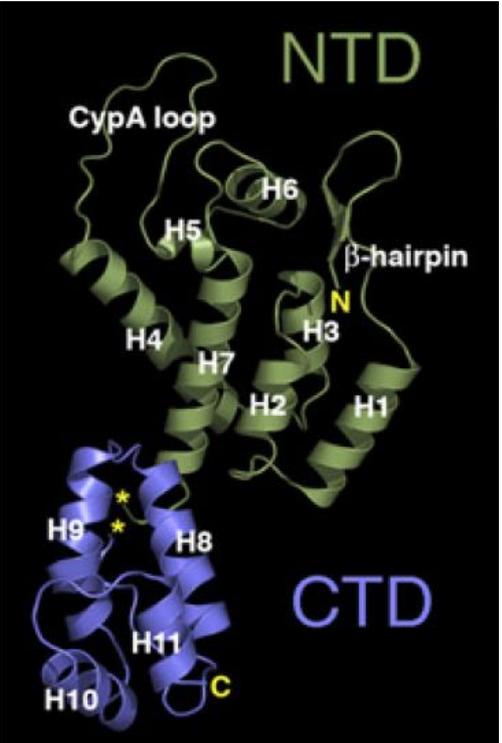
# Structure of Full Length CA

## NTDs and CTDs play different roles

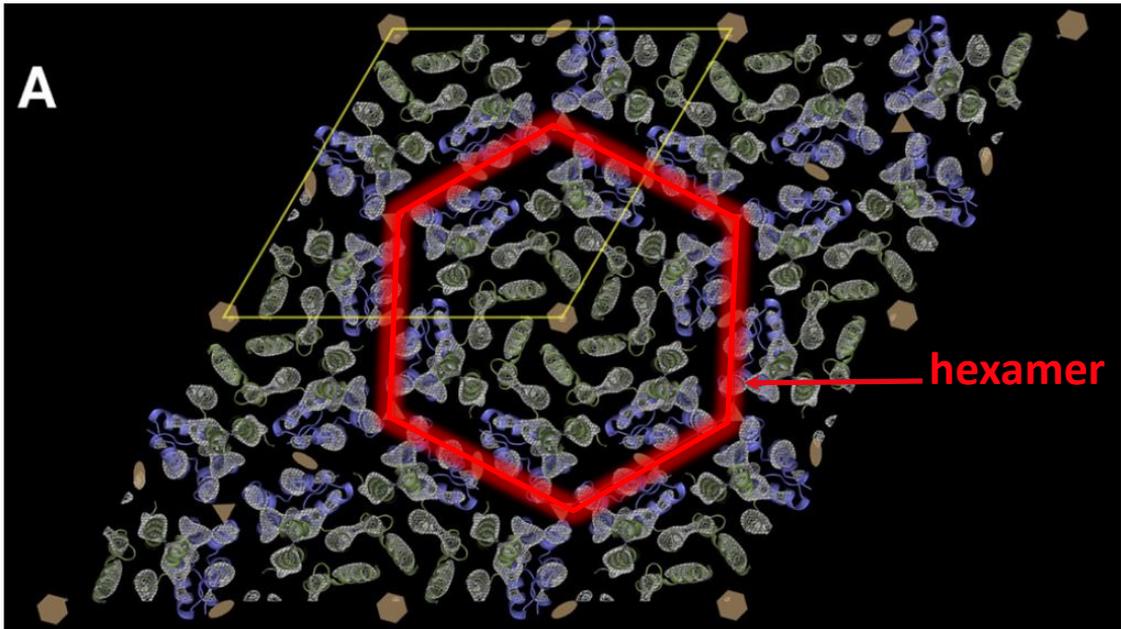
hexameric center      trimeric center

NTD = N-terminal domain

There are also dimeric symmetry centers!



CTD = C-terminal domain



CA-hexameric planar lattice in crystal

NTD fills inside space of hexamers, CTD fills outside.

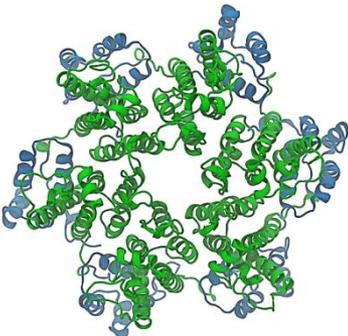
Ganser-Pornillos, B. K. et al. (2007). *Cell*, 131(1), 70–9.

# Structure of Full Length CA

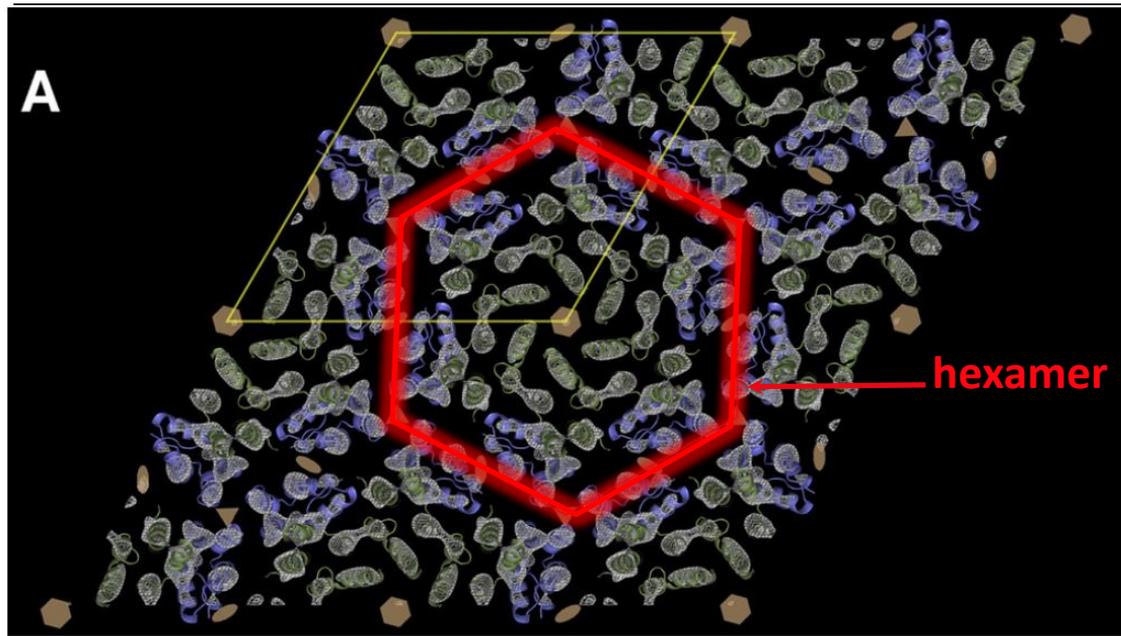
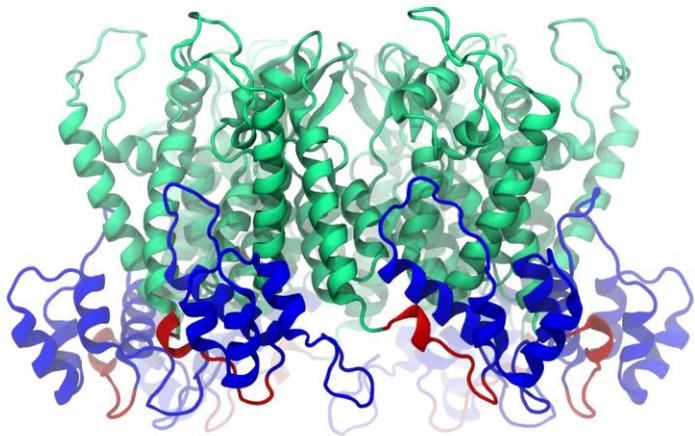
## NTDs and CTDs play different roles

hexameric center      trimeric center  
There are also dimeric symmetry centers!

C-terminal domain



N-terminal domain

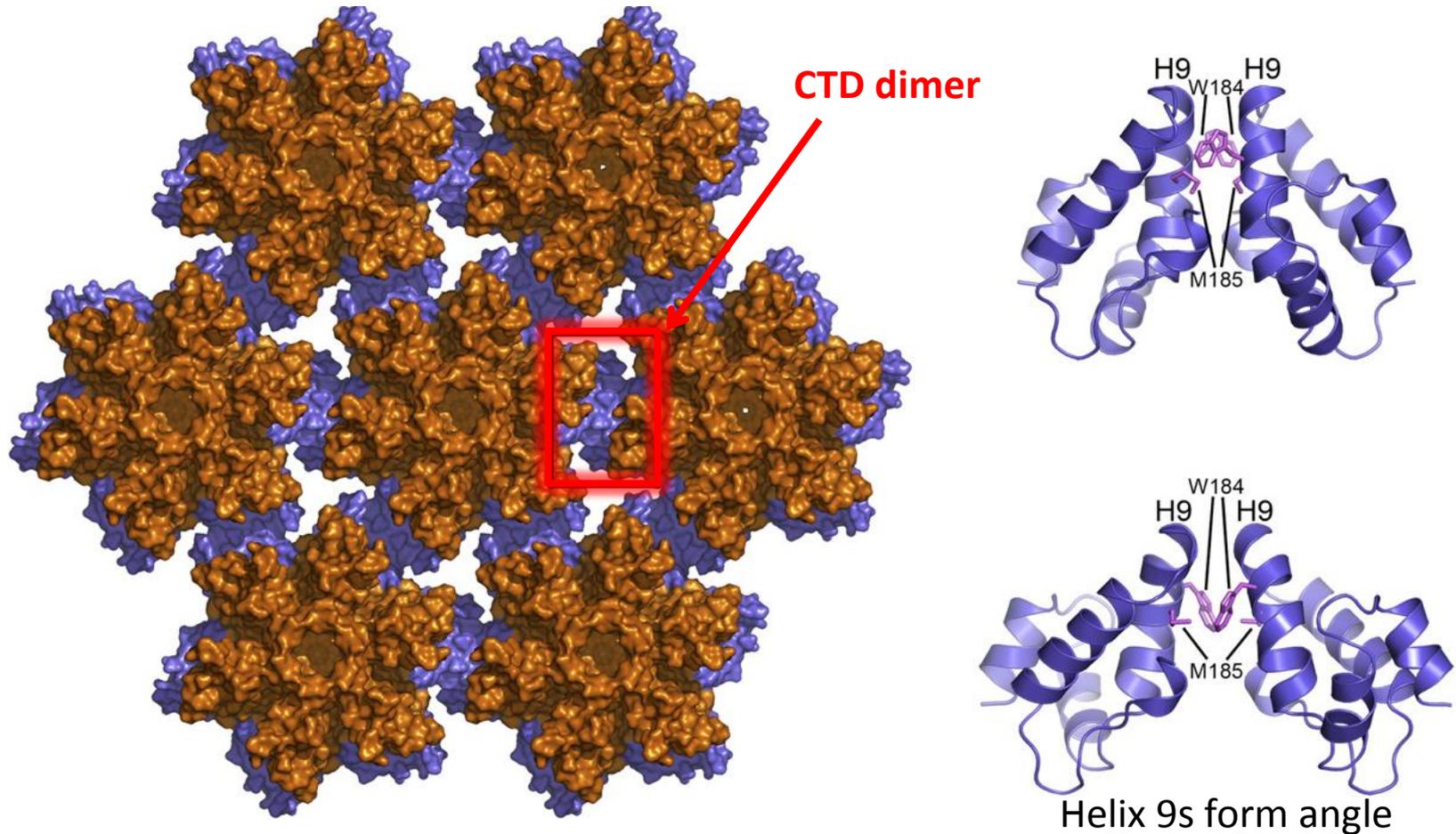


CA-hexameric **planar** lattice in crystal

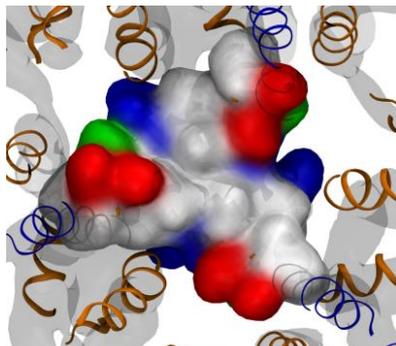
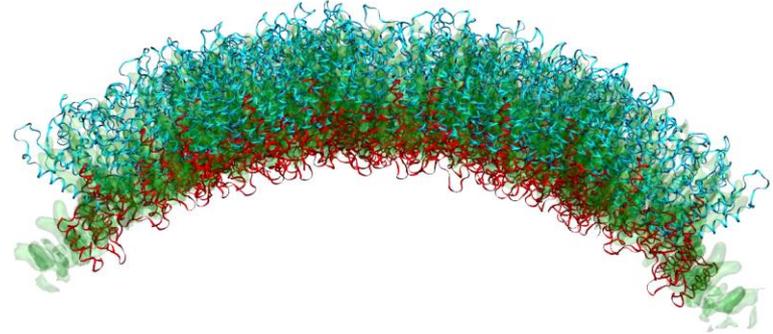
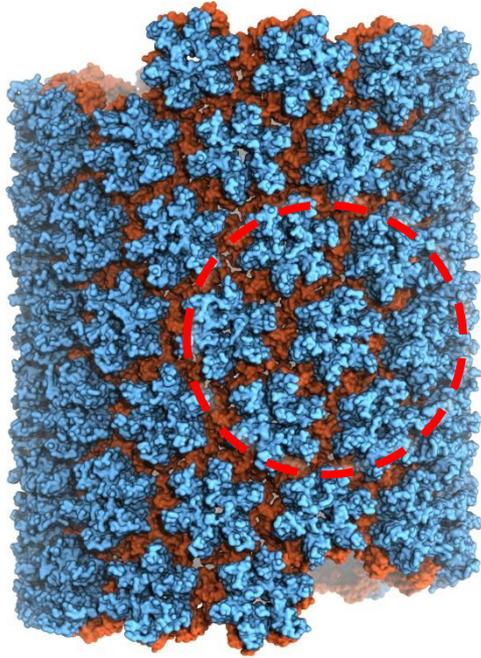
**NTD** fills inside space of hexamers, **CTD** fills outside.

Ganser-Pornillos, B. K. et al. (2007). *Cell*, 131(1), 70–9.

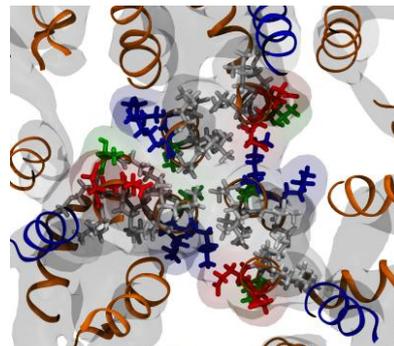
# NMR structures for the CTD dimers



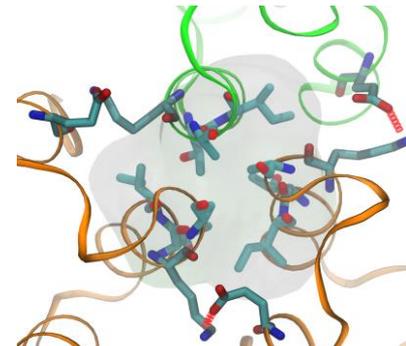
# MDFF Structural Refinement



hydrophobic interaction at trimeric interface, surr. by polar residues

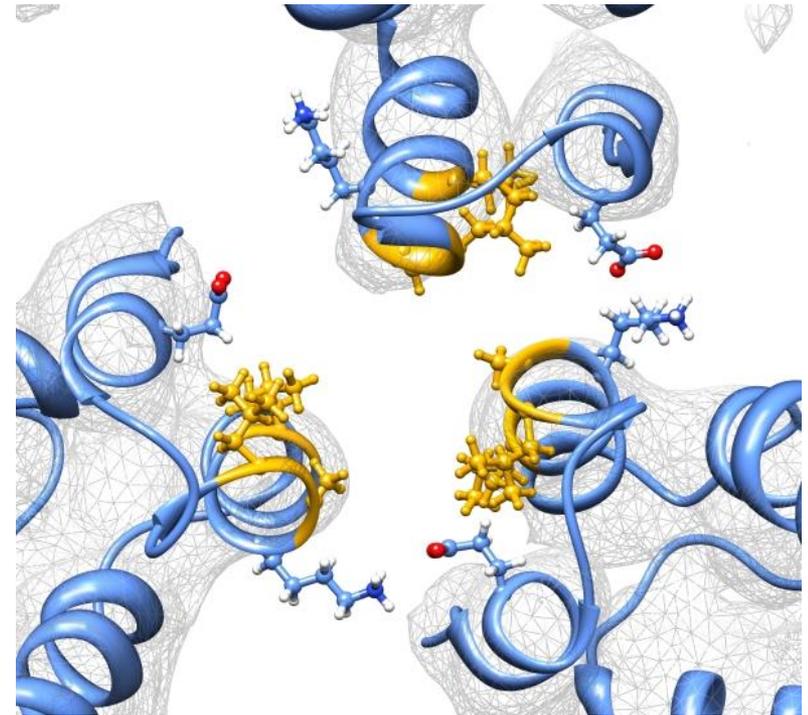
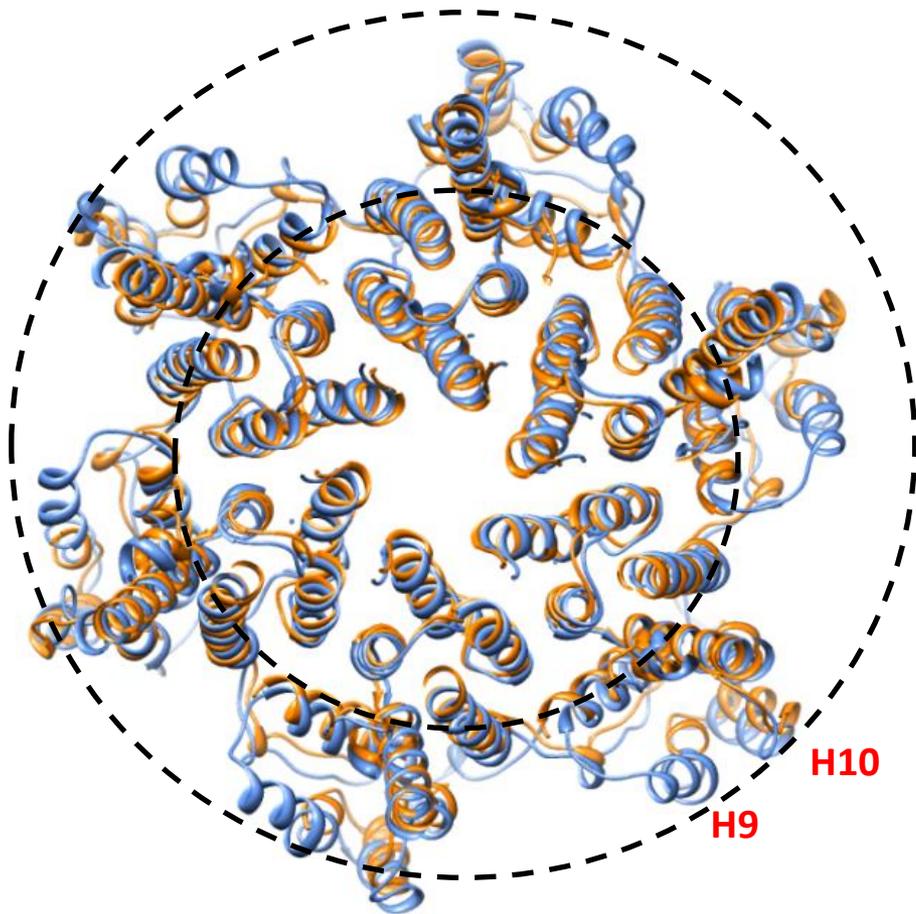


a view into the trimeric interface, contacts between three helices\_10

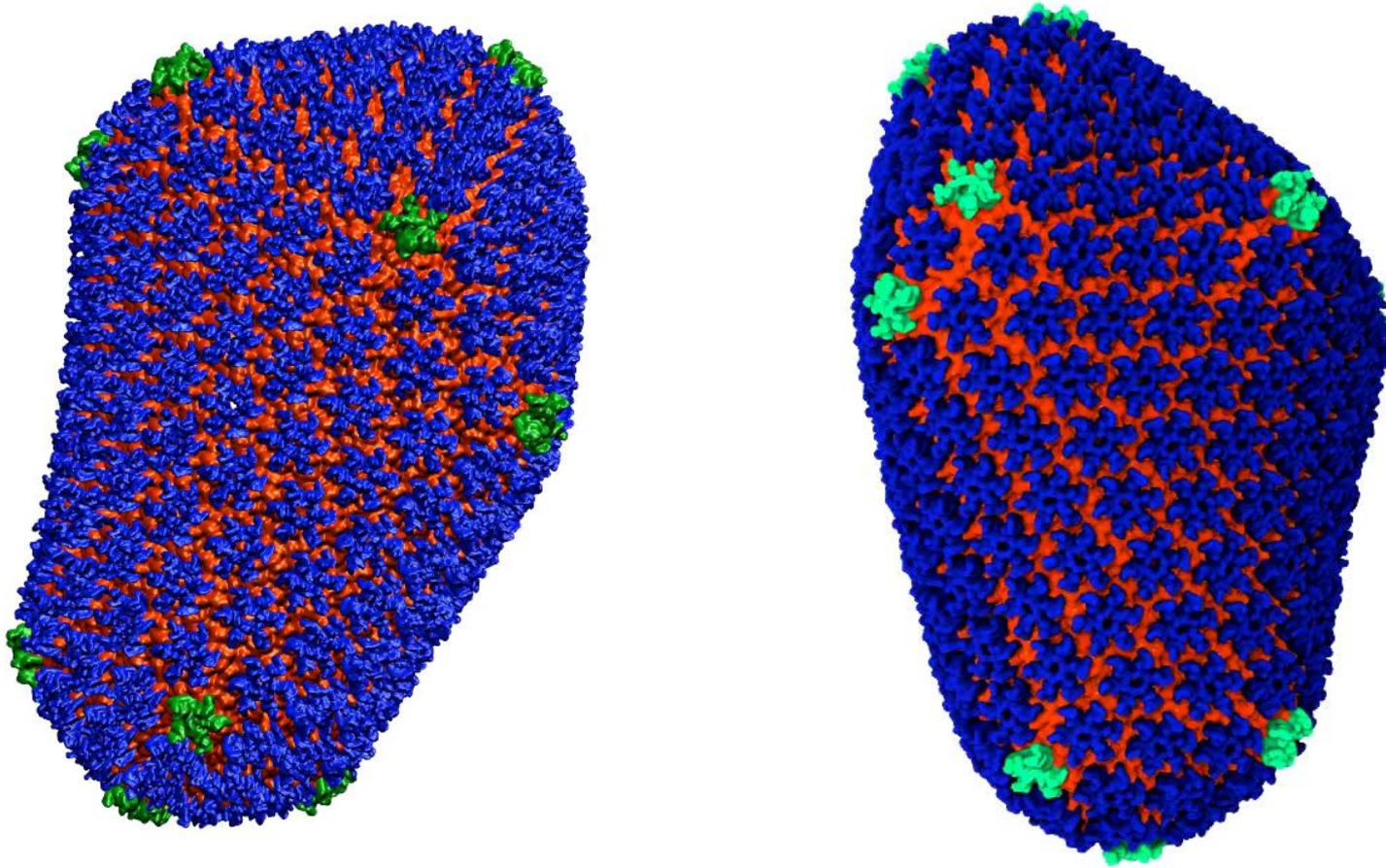


formation of peripheral hydrogen bonds

# MDFF reveals a different hexameric conformation and a novel trimeric interface

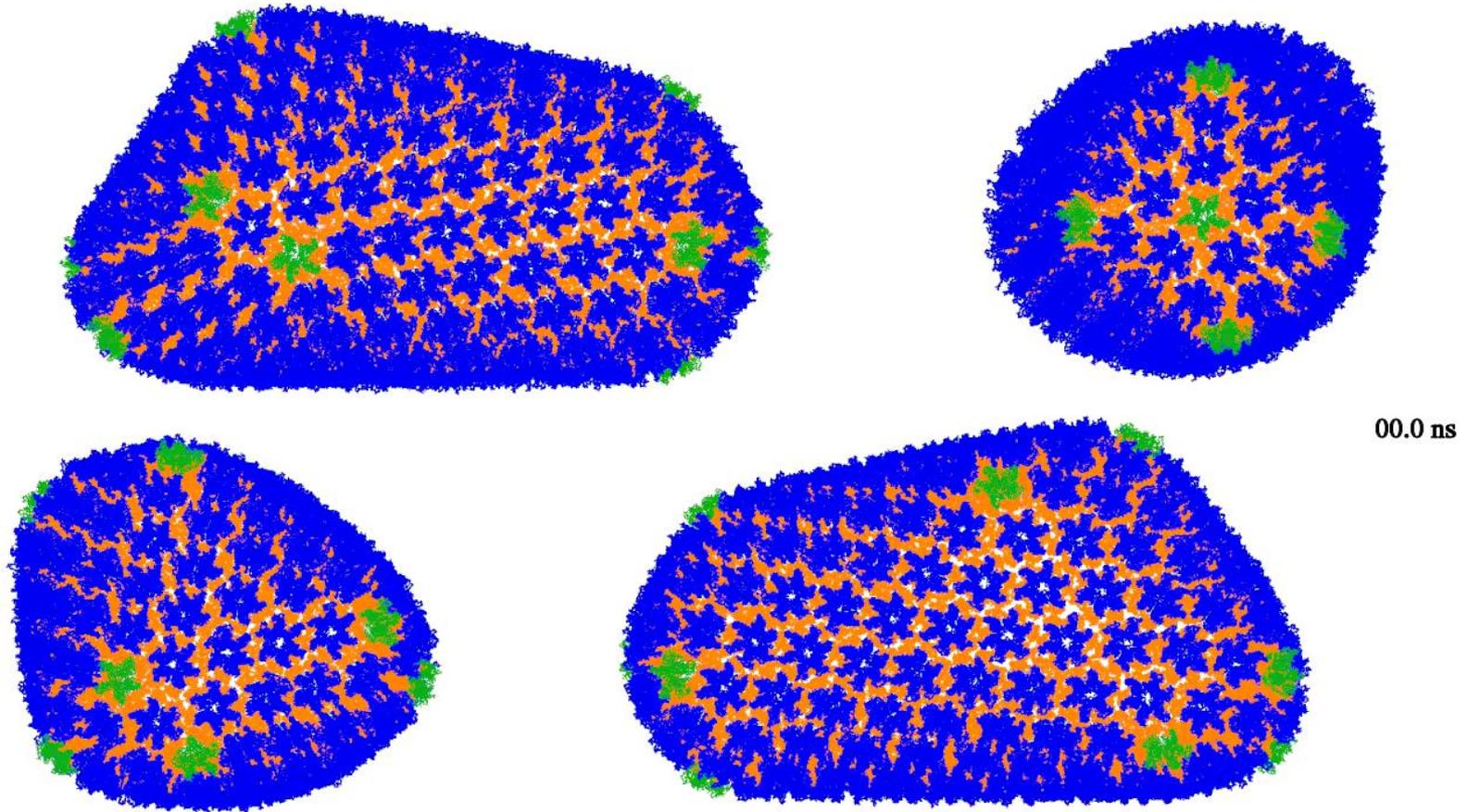


# All-atom MD simulation of mature HIV-1 capsid



- 216 hexamers +12 pentamers (13.4 M),
- 64 million atoms total including solvent
- 100 ns running NAMD on 4000 Cray-XE nodes, 128000 cores
- 6 ns/day

# All-atom MD simulation of mature HIV-1 capsid



- 186 hexamers +12 pentamers (12 M),
- 64 million atoms total
- 500 ns running *NAMD* on NCSA Blue Waters
  - 2000 to 3500 XK Nodes equipped with NVIDIA Tesla K20X
- 9 to 14 ns/day



# Solvation of large systems

Avoid GUI altogether. Scripts offer more control over the modeling process.

**Use JS file format.** Available in all version of **psfgen** and **developmental versions of VMD.**

- **PROBLEM:**
  - Solvate GUI/script performs several atom selections that result in poor performance for large systems. Solvating a large structure can take **days**.

**# Create an empty water box of 100nm x 100nm x 100nm**

```
solvate -o solvent.js -minmax {{0 0 0} {1000 1000 1000}}
```

**# Load solute and solvent and save as a single js file with clashes.**

```
package require psfgen
```

```
readjs solute.js
```

```
readjs solvent.js
```

```
writejs clashes.js
```

**# Save non clashing atoms**

```
mol new clashes.js
```

```
set good [atomselect top "not same residue as within 2.4 of not segname \"W.*\""]
```

```
$good writejs final.js
```

```
$good writenamdbin final.coor
```

# Ionization of large systems

Autoionize does NOT handle large structures properly.

Limited to one segment (9999 atoms).

Also a problem for high ionic concentrations in medium sized systems.

Default parameters for ions are CHARMM22 unless manually changed.

## # Get water segments

```
set water [atomselect top "water"]
```

```
set segments [lsort -unique [$water get segnames]]
```

## # Loop over water segments and ionize each segment independently

```
foreach watseg $segments {
```

### #Select current water segment

```
set atomsel [atomselect top "segname ${watseg}"]
```

### #Write segment as a pdb

```
atomsel writepdb water/${watseg}.pdb
```

### #Create a psf for water segment

```
segment ${watseg} {
```

```
  auto none
```

```
  pdb watert/${watseg}.pdb
```

```
} ; coordpdb water/${watseg}.pdb $watseg
```

```
writepdb water/${watseg}.pdb
```

```
writesf watert/${watseg}.psf
```

# Ionization of large systems

# Get new segment name for ionized water box

```
set number [string trimleft ${watseg} W]  
set segment I${watseg}
```

# Ionize each water segment

```
    autoionize -psf water/${watseg}.psf -pdb water/${watseg}.pdb -sc $sc  
              -o ions/$segment -seg $segment  
}
```

# Load solute and ionized water box

```
resetpsf  
readjs solute.js  
foreach watseg $segments {  
    set number [string trimleft ${watseg} W]  
    set segment I${watseg}  
    readpsf ions/$segment  
    coordpdb ions/$segment  
}  
writejs ionized.js  
writenamdbin ionized.coor
```

**IMPORTANT:** Make sure the system is neutralized, add more ions if necessary.

# Special considerations for simulations of large systems

- Use the new minimizer. Older versions of NAMD require the use of velocity quenching.
- Make use of the multi-time stepping algorithm available in NAMD  
MTS 2-1-3 (2fs inner loop, 2 fs non-bonded, 6fs electrostatics)  
**shake** must be enabled.
- Set **PME** grid spacing to **2Å** and increase **interpolation order** to **8**.
- Make use of the **hybrid** load balancer
- Use the **memory optimized** version of NAMD.  
<http://www.ks.uiuc.edu/Research/namd/wiki/?NamdMemoryReduction>  
Different handling of fixed and restrained atoms.  
Supports TMD, TCL forces, and other features.
- Limit the number of I/O nodes. NAMD uses parallel I/O.
- If running in NPT, set the period of the barostat to a large value (e.g. ~ 15ps).
- Watch out for excluded volumes for the appearance of bubbles.