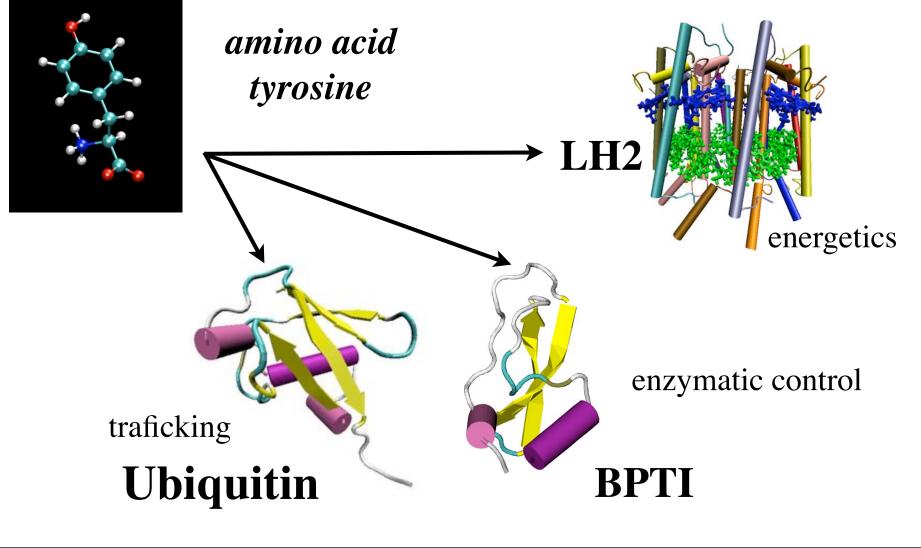
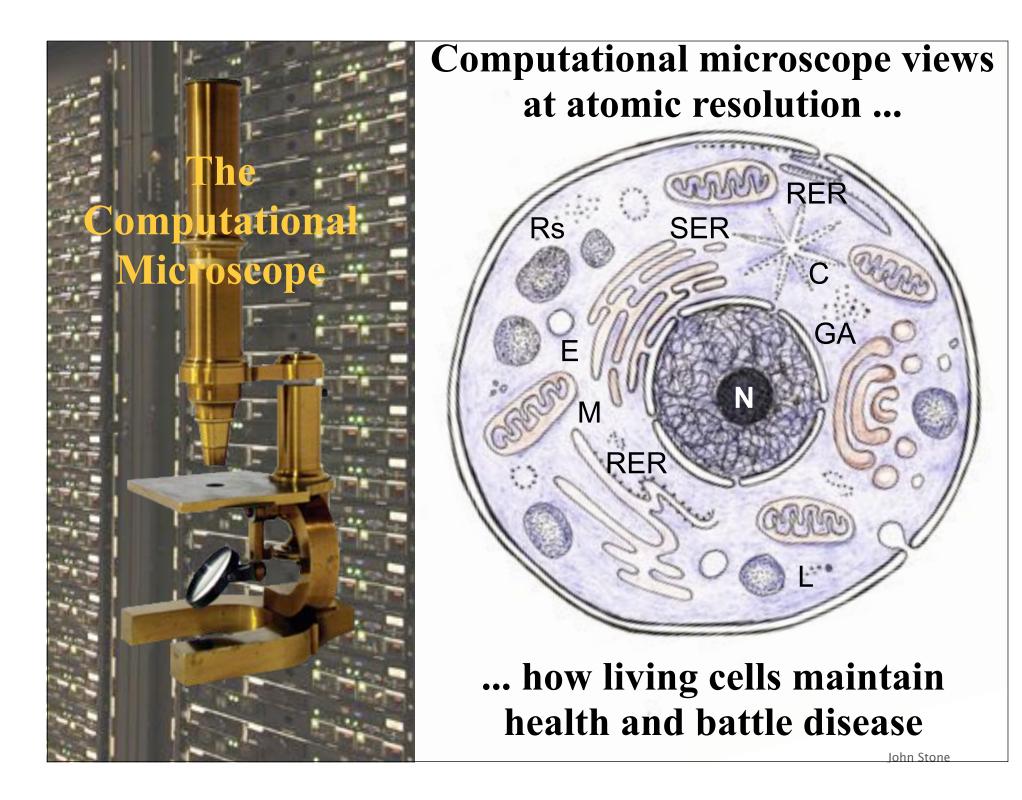
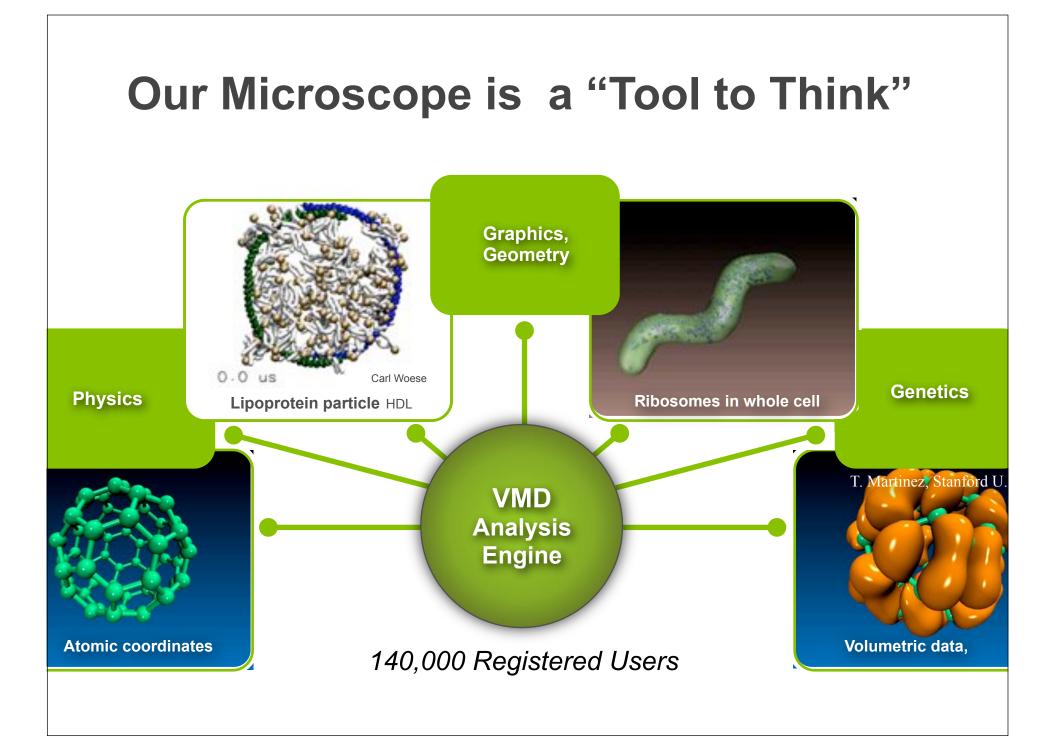
## Lecture 1a Introduction to Protein Structures -VMD Tool



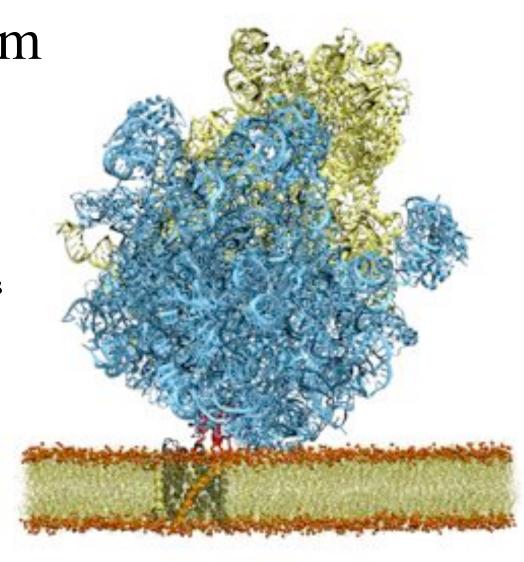


#### **Our Microscope is Made of...** $U(\vec{R}) = \underbrace{\sum_{bonds} k_i^{bond} (r_i - r_0)^2}_{U_{bond}} + \underbrace{\sum_{angles} k_i^{angle} (\theta_i - \theta_0)^2}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihe} [1 + \cos(n_i \phi_i +$ Chemistry Software 100 **10** Virus $\sum_{i} \sum_{j \neq i} 4\epsilon_{ij} \left[ \left( \frac{\sigma_{ij}}{r_{ij}} \right)^{12} - \left( \frac{\sigma_{ij}}{r_{ij}} \right)^{6} \right] + \sum_{i} \sum_{j \neq i} \frac{q_i q_j}{\epsilon r_{ij}}$ 128 256 512 1024 1024 4096 8192 6384 2768 40,000 registered users cores **Physics** $m_i \frac{d^2 \vec{r_i}}{dt^2} = \vec{F_i} = -\vec{\nabla} U(\vec{R})$ 10000 Math $\vec{r}_i(t + \Delta t) = 2\vec{r}_i(t) - \vec{r}_i(t - \Delta t) + \frac{\Delta t^2}{m_i}\vec{F}_i(t)$ ...and Supercomputers (repeat **one billion times** = microsecond)



# Highlights of the VMD Molecular Graphics Program

- > 140,000 registered users
- Platforms:
  - Unix / Linux
  - Windows
  - MacOS X
- Display of large biomolecules and simulation trajectories
- Sequence browsing and structure highlighting
- Multiple sequence structure analysis
- User-extensible scripting interfaces for analysis and customization

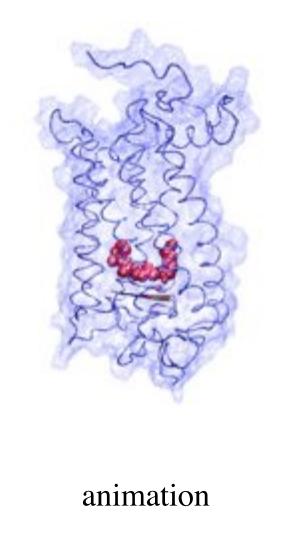


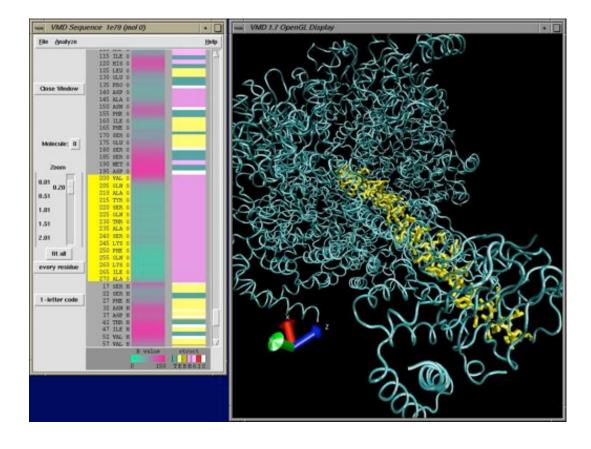
# Key Features of VMD

- General 3-D molecular visualization with extensive drawing and coloring methods
- Extensive atom selection syntax for choosing subsets of atoms for display
- Visualization of dynamic molecular data
- Visualization of volumetric data
- No limits on the number of molecules or trajectory frames, except available memory
- Rendering high-resolution, publication-quality molecule images
- Movie making capability
- Extensions to the Tcl/Python scripting languages
- $\bullet$  Extensible source code written in C and C++
- Built in GPU acceleration support
- Supports cluster computing for extensive analysis runs
- Building and preparing systems for molecular dynamics simulations
- Interactive molecular dynamics simulations

## Molecular Graphics Perspective of Protein Structure and Function

see tutorial at http://www.ks.uiuc.edu/Training/Tutorials/



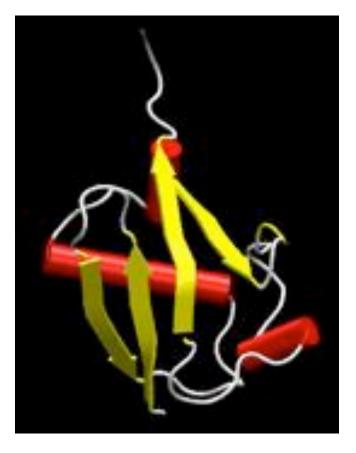


sequence

#### structure

# Ubiquitin

- 76 amino acids
- highly conserved
- covalently attaches to proteins and tags them for degradation
- other cell traficking



• Glycine at C-terminal attaches to the Lysine on the protein by an isopeptide bond.

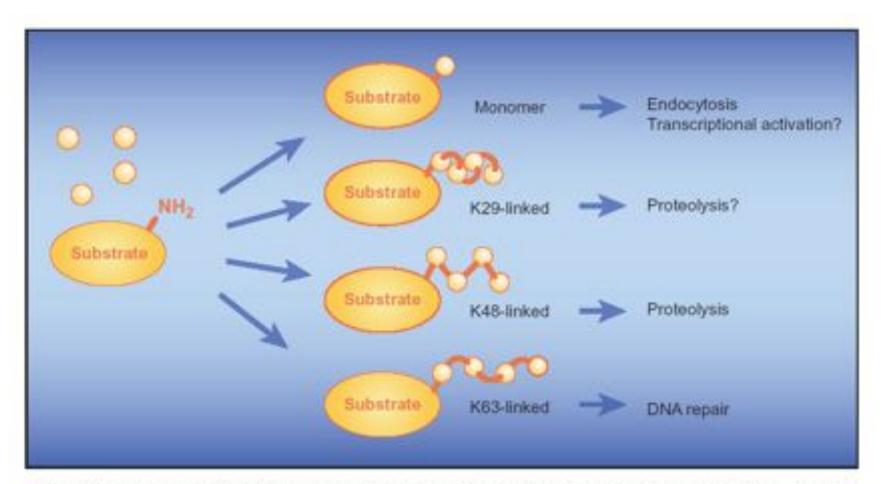
• it can attach to other ubiquitin molecules and make a polyubiquitin chain.

There are 7 conserved lysine residues in ubiquitin.



Two ubiquitins attached together through LYS 48. LYS 63 and LYS 29 are also shown there.

#### Multi-ubiquitylation targets destination of proteins



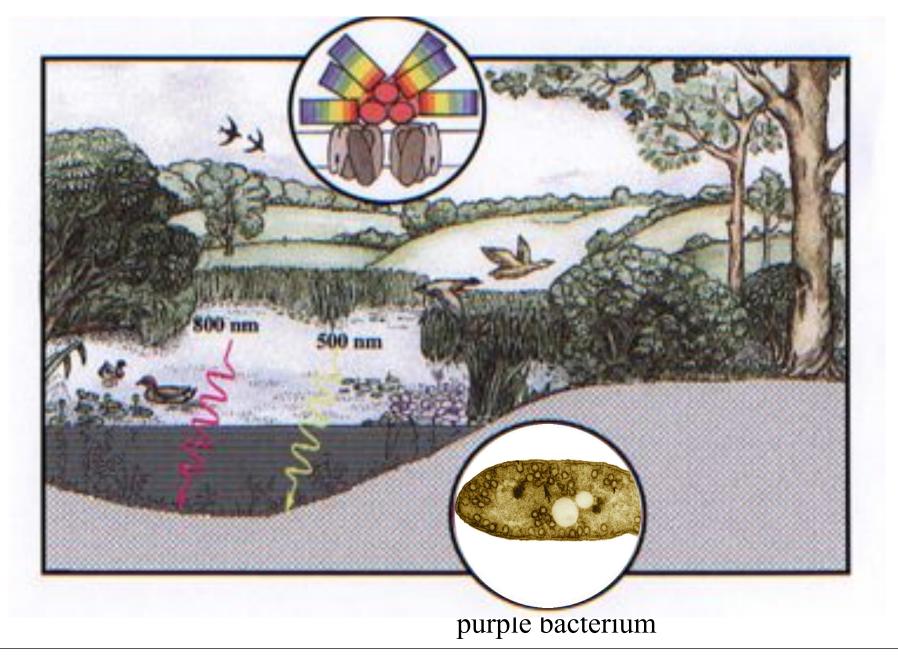
Multifaceted. Ubiquitin can attach to its various substrate proteins, either singly or in chains, and that in turn might determine what effect the ubiquitination has. (K29, K48, and K63 refer to the particular lysine amino acid used to link the ubiquitins to each other.)

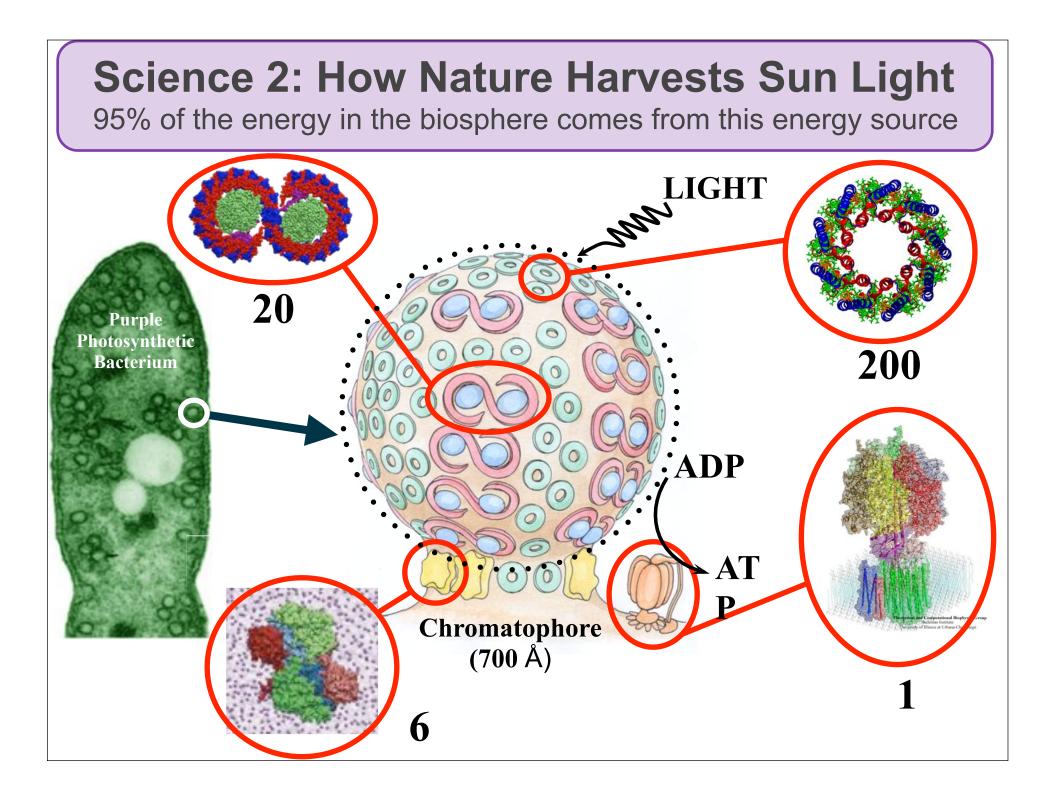
Marx, J., Ubiquitin lives up its name, Science 297, 1792-1794 (2002)

#### VMD session with ubiquitin

# Form-follows-function architecture of purple bacterial light harvesting system

#### Habitats of Photosynthetic Life Forms





#### **Chromatophore of Purple Bacteria** (section of the chromatophore membrane) **ADP** cytoplasm **ATP** H+ membrane Q/QH2/Q **ATPase** hγ bc₁ RC **e**<sup>-</sup> LH-I LH-II cytochrome c<sub>2</sub> periplasm

# Knowing the Atomic Level StructureImage: Structure of the chromatophore, one<br/>can systematically<br/>describe its physical

ATP

mechanism

M. Sener, J. Olsen, N. Hunter, and K. Schulten. *PNAS*, **104**: 15723-15728, 2007

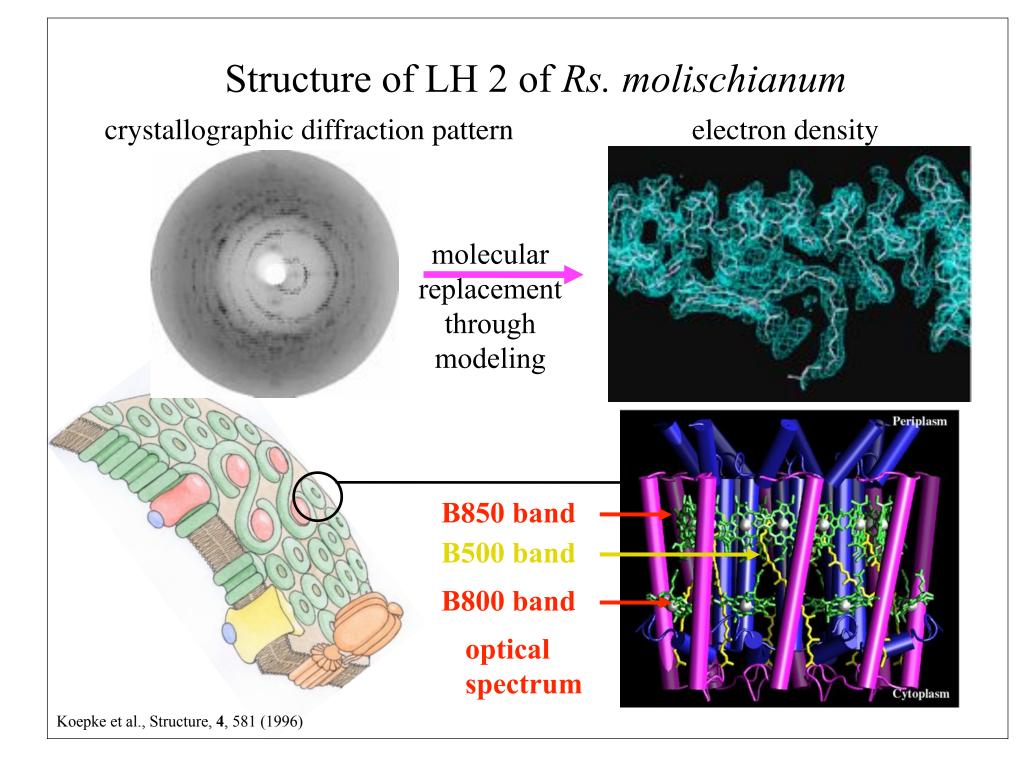
light

#### **How Nature Harvests Sun Light**

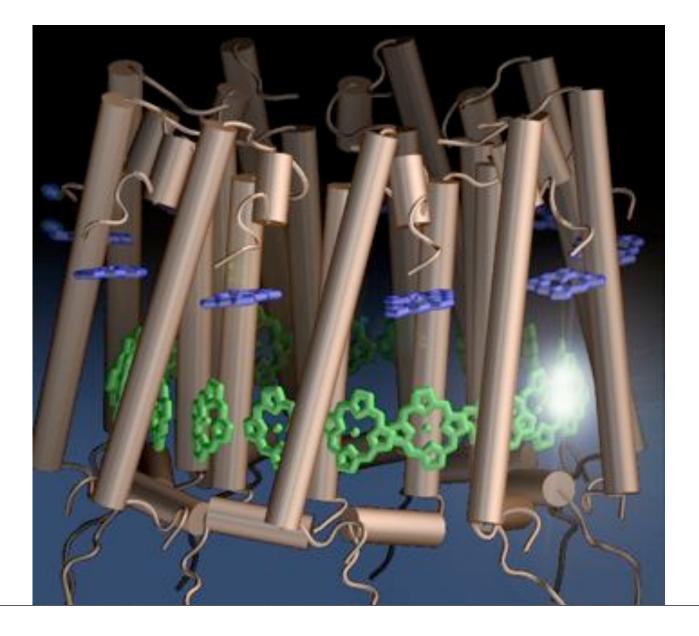
First 23 million atom molecular dynamics simulation

AFM image of flat chromatophore membrane (Scheuring 2009) 10nm

Simulation was realized through new versions of our modeling programs VMD/NAMD, new data storage formats, and parallel I/O.



# **Excitonic dynamics in LH2**



#### VMD session with LH2

#### VMD – "Visual Molecular Dynamics" http://www.ks.uiuc.edu/Research/vmd/

#### Visualization and analysis of:

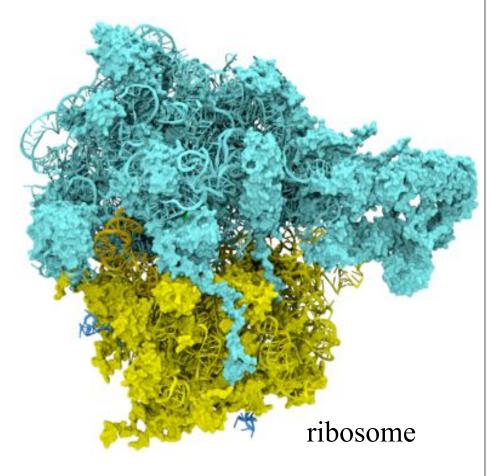
- Molecular dynamics and quantum chemistry simulations
- Sequence data
- Cryo-electron microscopy maps

# User-extensible with built-in scripting and many plugins

Supports very large data sets, batch mode analysis on clusters

# Takes advantage of advanced technological opportunities:

- High quality interactive display, batch mode rendering, movie making
- Supports multi-core processors, GPUs





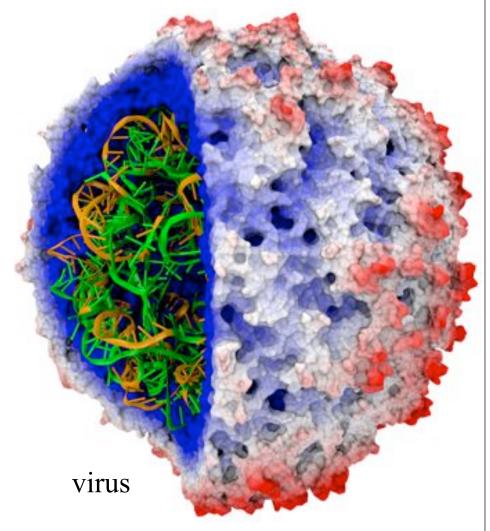
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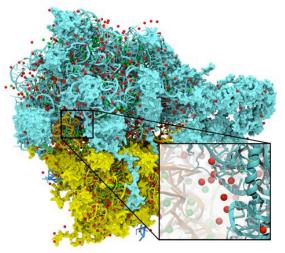
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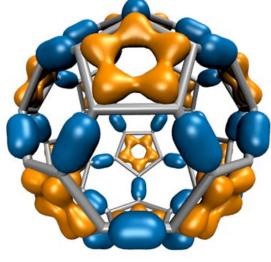


NIH Resource for Biomolecular Modeling and Bioinformatics http://www.ks.uiuc.edu/

## CUDA+OpenCL Acceleration in VMD

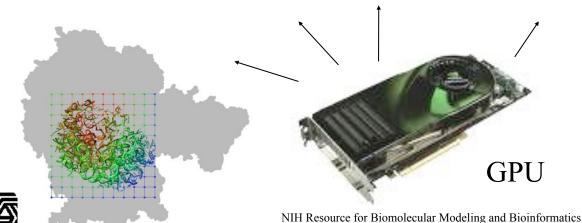


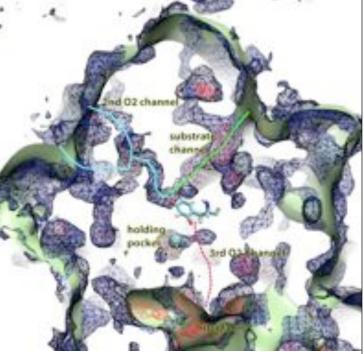
Electrostatic field calculation, Multilevel Summation Method 20x to 44x faster



Molecular orbital calculation and display 100x to 120x faster

http://www.ks.uiuc.edu/





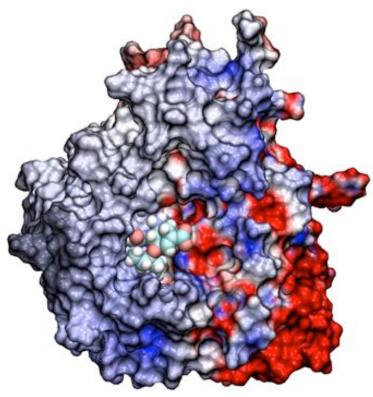
Imaging of gas migration pathways in proteins with Implicit Ligand Sampling (ILS) algorithm

20x to 30x faster

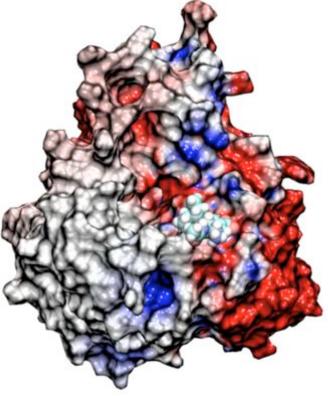


#### Swine Flu Neuraminidase Electrostatics

Mean electrostatic field needed to identify drug binding pathway



Time-averaged electrostatic field of H1N1 neuraminidase calculated from VMD Multilevel Summation Tool: Compution with NVIDIA Tesla C1060 is 20x faster than computation on a single CPU core



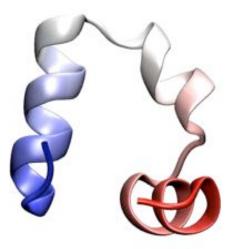
Movie of drug (tamiflu) binding



NIH Resource for Biomolecular Modeling and Bioinformatics http://www.ks.uiuc.edu/ Beckman Institute, UIUC

# Timeline Tool: identify events in an MD trajectory

• We have MD trajectories:

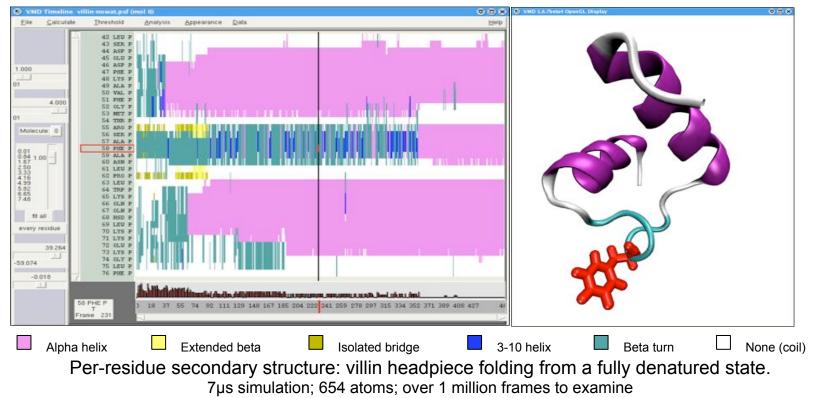


- We want to identify events in a trajectory:
  - 7.1 µs, 600 GB of trajectory data
  - events: 0.5  $\mu$ s, helix 3 forms; 3.0  $\mu$ s, helix 1 forms; etc.
  - How long would it take an expert user to visually inspect this trajectory to find motional changes of events?

2 days! (plus: tiring task; one is liable to miss much)

# Timeline: a graphing and analysis tool to identify events in an MD trajectory

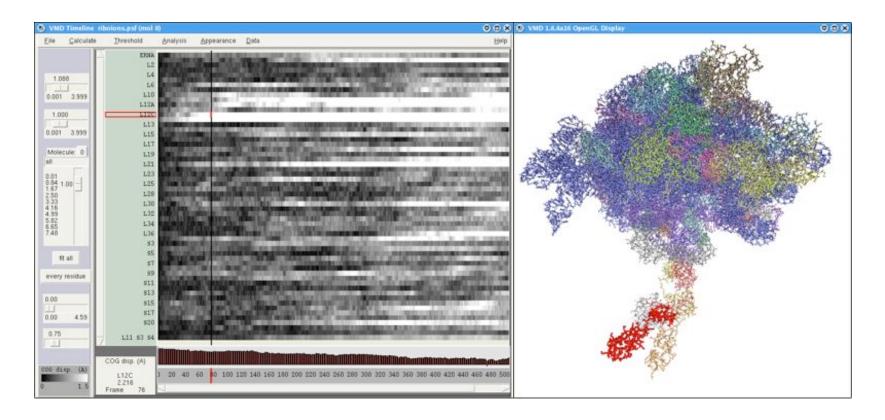
Events during 7 µs villin headpiece folding



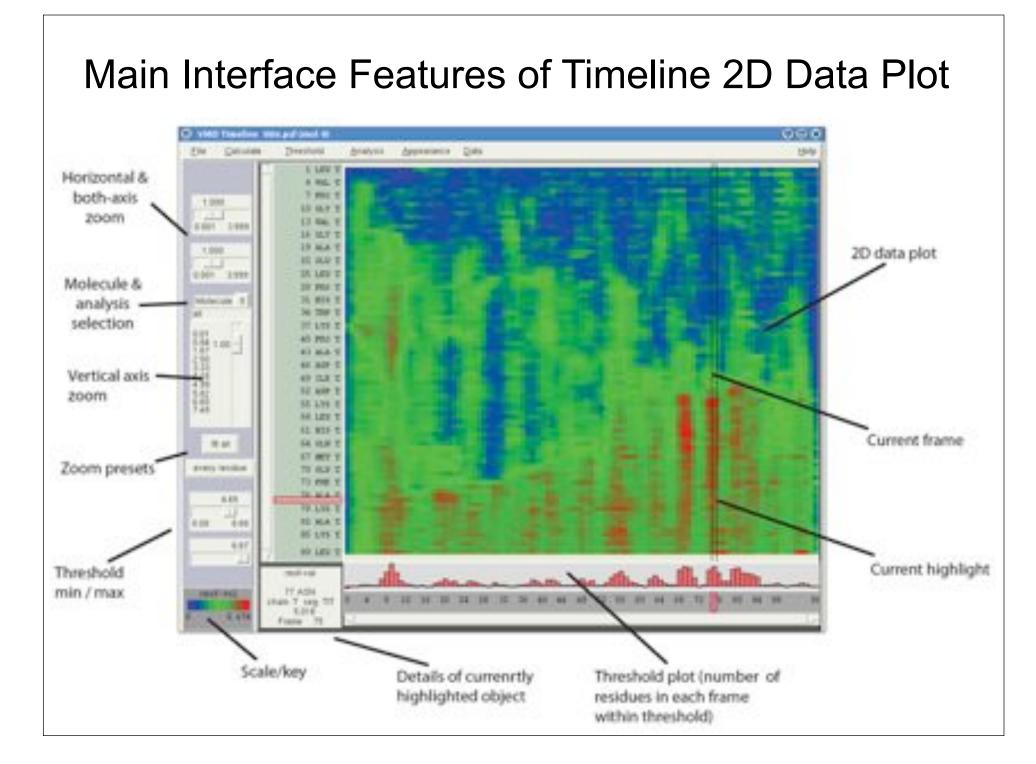
VMD Timeline plug-in: live 2D plot linked to 3D structure

- a single picture shows changing properties across entire structure, entire trajectory.
- explore time vs. attribute (per-residue or per-selection) linked to molecular structure
- many analysis methods available; user-extendable

# Timeline and large structures: events during ribosome equilibration



Ribosome equilibration, 17,000+ protein/nucleic residues + ions *Example analysis*: displacement (Å) of center-of-geometry of each component protein (calculation here is per-component-protein, not per-individal-residue) *Finding:* peripheral proteins show greater displacement than core proteins





## Acknowledgements

VMD team J. Stone (leader) D. Hardy B. Isralewitz J. Saam K. Vandivoort





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