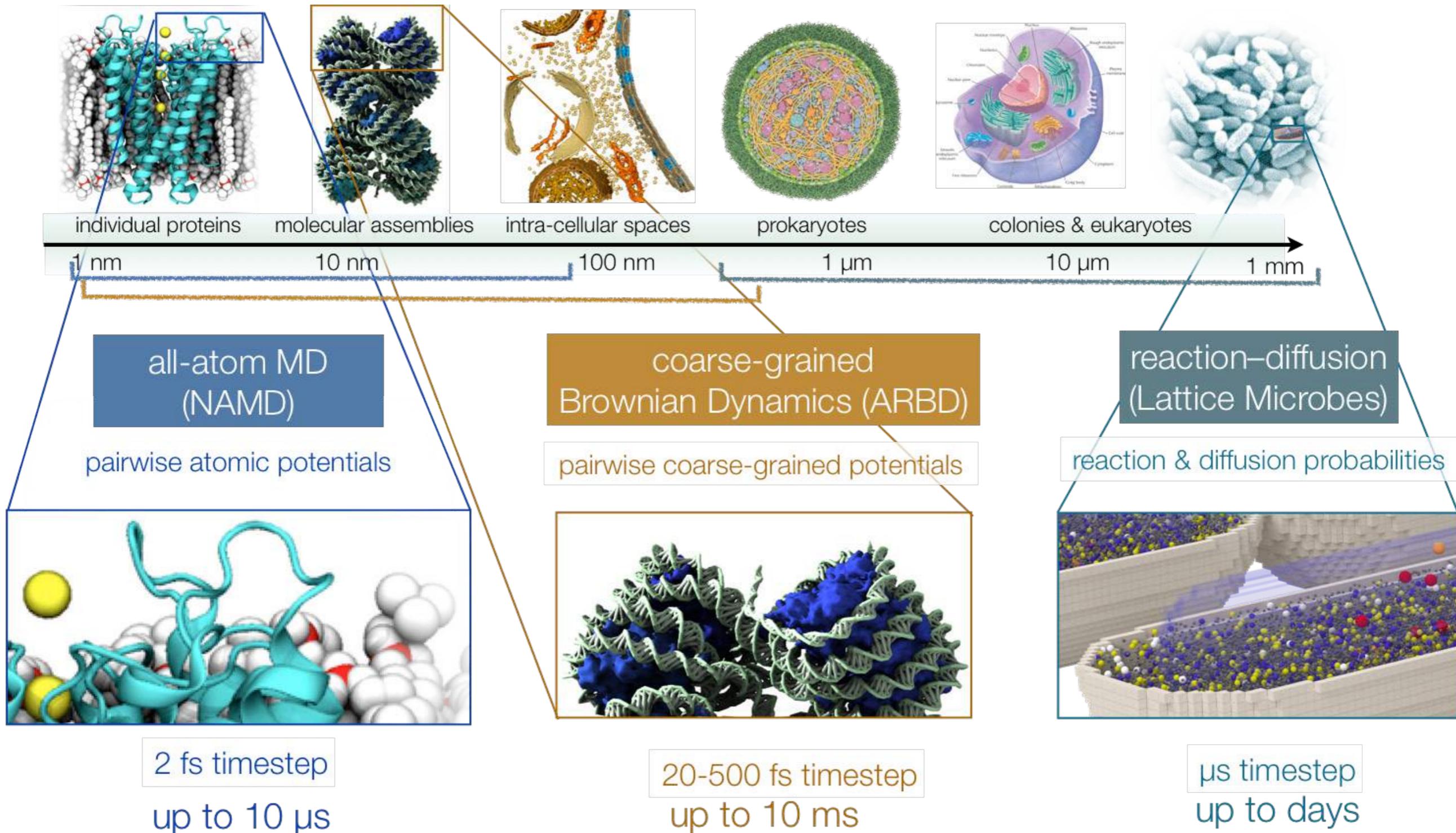


# Introduction to MD simulations of DNA systems

Aleksei Aksimentiev

# Biological Modeling at Different Scales

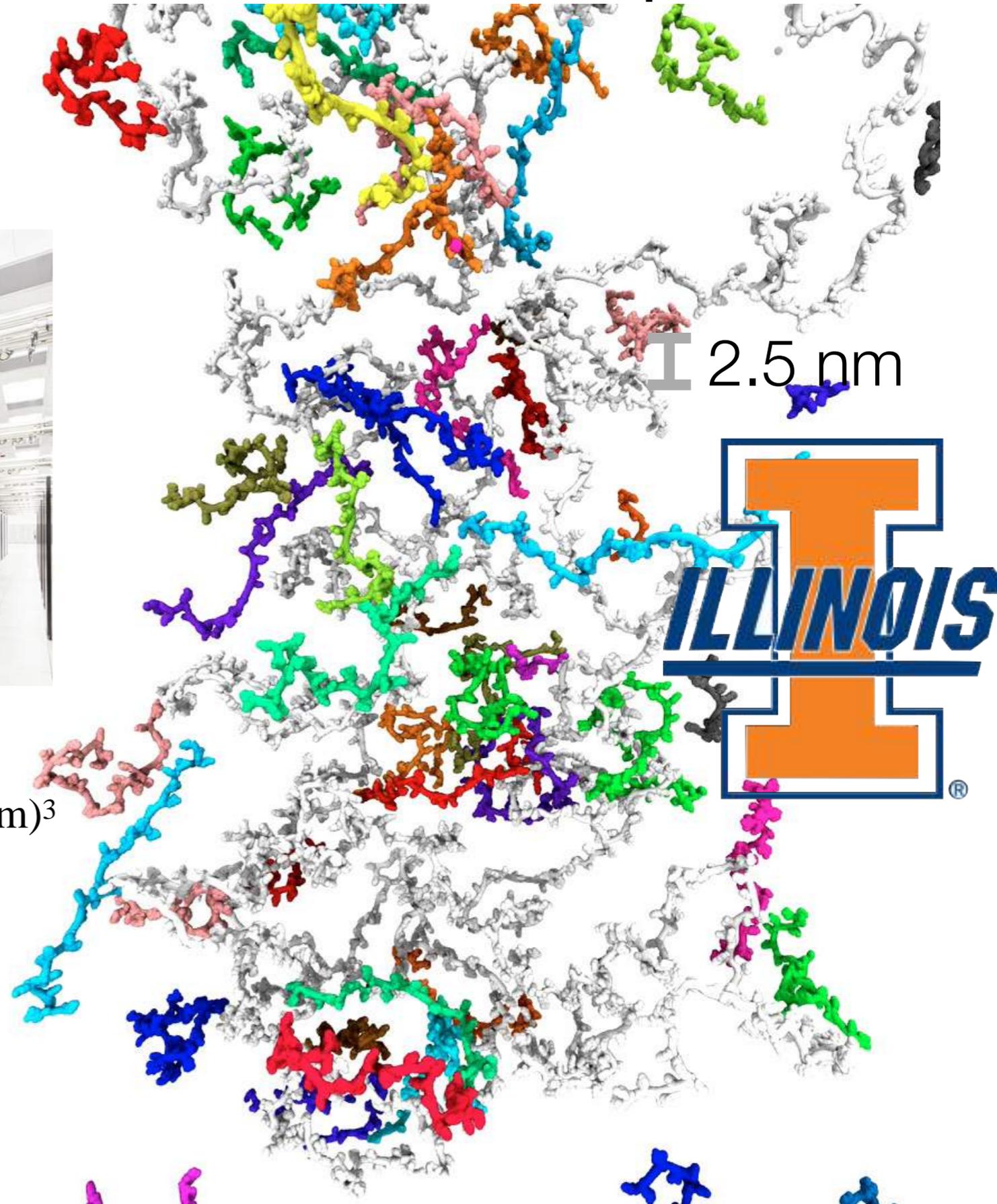
spanning orders of magnitude in space and time



# The computational microscope

Massive parallel computer

Blue Waters (UIUC): ~200,000 CPUs



Time scale: ~ 0.1-100  $\mu$ s

Length scale: 10K - 1,000M atoms or ( $< 70$  nm)<sup>3</sup>

Time resolution: 2 fs

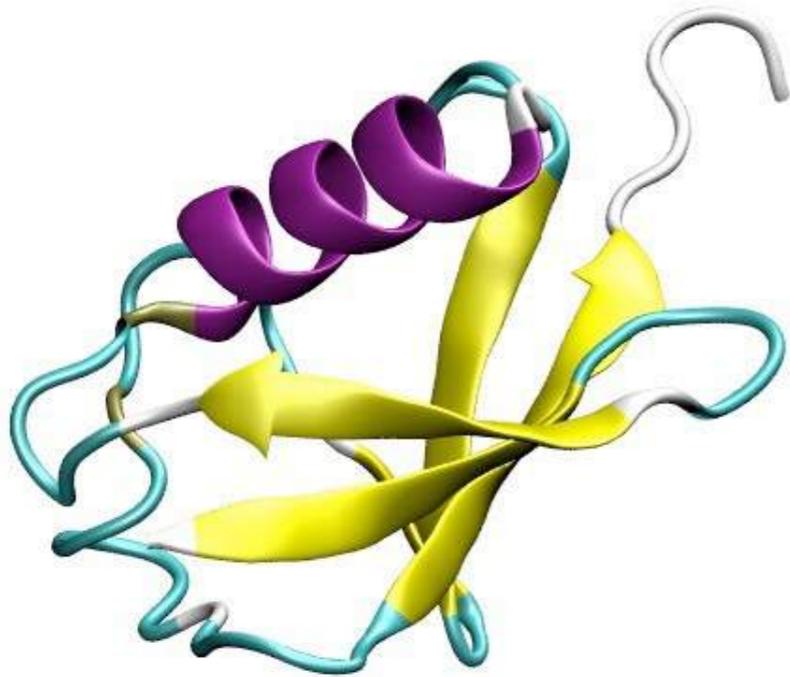
Spacial resolution: 0.1 Å

Atoms move according to classical mechanics  
( $F = ma$ )

Interactions between atoms are defined by  
the molecular force field

# What is Molecular Dynamics?

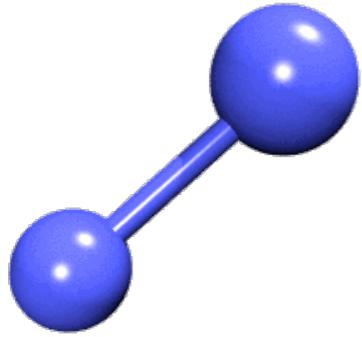
$$m_i \frac{d^2 \vec{r}_i}{dt^2} = \vec{F}_i = -\vec{\nabla} U(\vec{R})$$



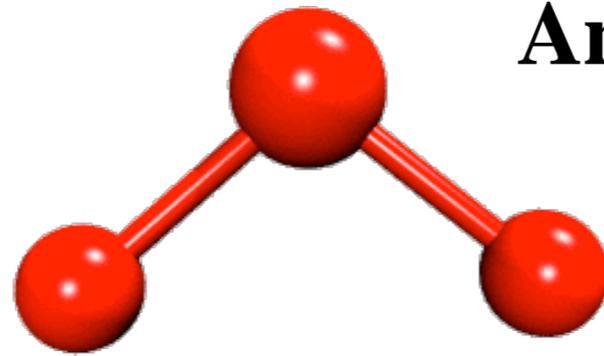
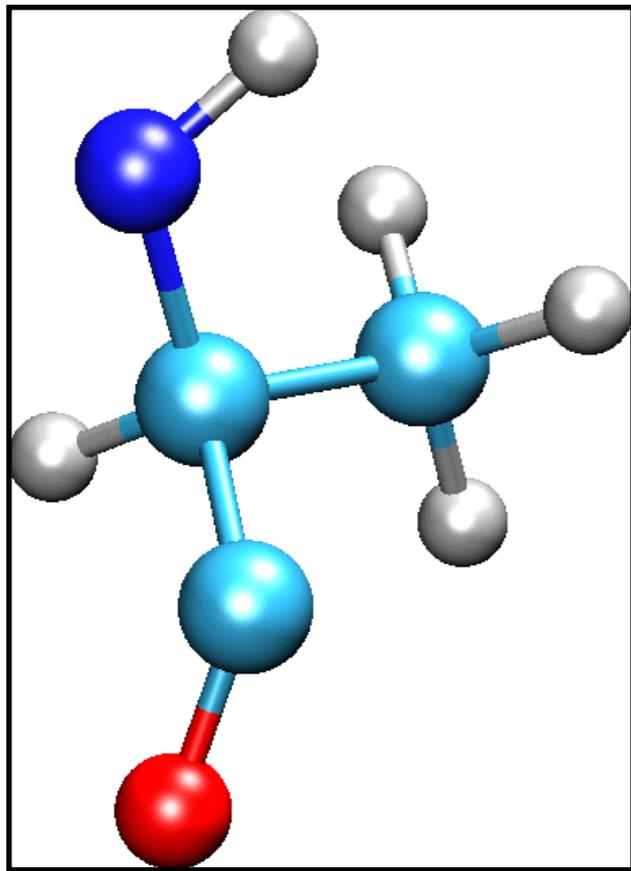
Energy function has two parts:

- chemical bond interactions
- non-bonded interactions

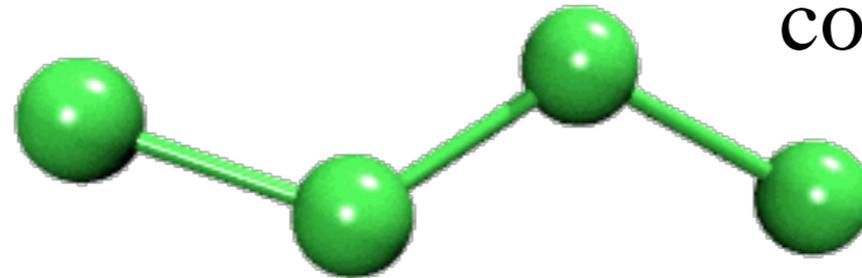
# Chemical Structure



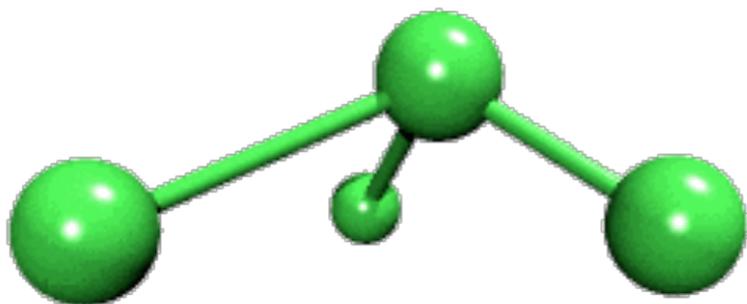
**Bonds:** Every pair of covalently bonded atoms.



**Angles:** Two bonds that share a common atom form an angle.



**Dihedrals:** Two angles that share a common bond form a dihedral.

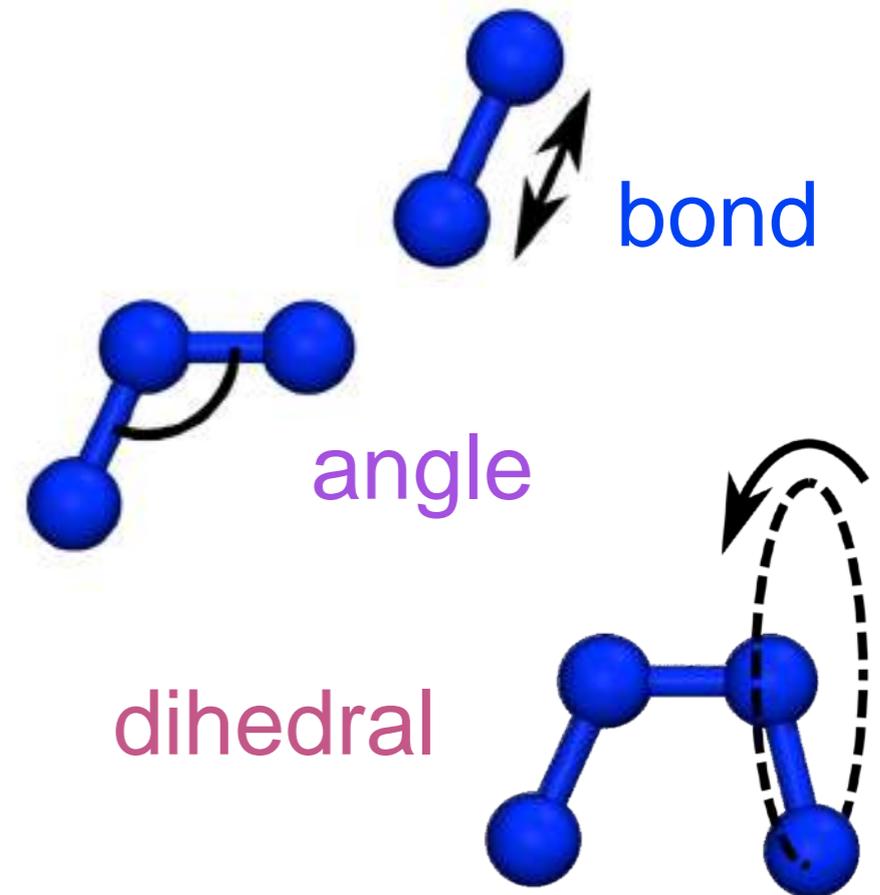


**Improper:** Any *planar* group of four atoms forms an improper.

$$m_i \frac{d^2 \vec{r}_i}{dt^2} = \vec{F}_i = -\vec{\nabla} U(\vec{R})$$

The MD method models atoms with point particles interacting through classical potentials

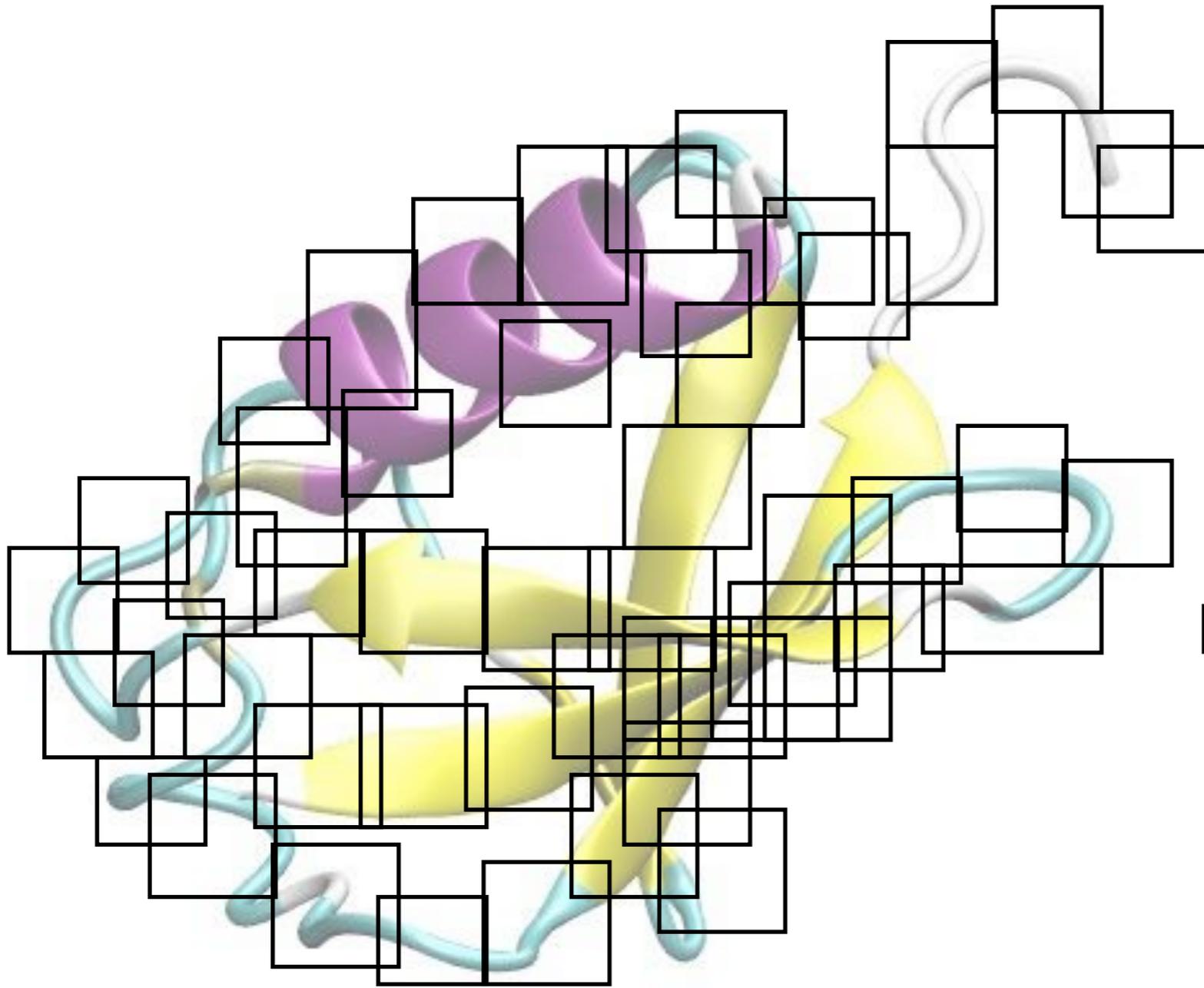
$$U = \sum_{\text{bonded}} \left\{ \begin{aligned} &k(r_{ij} - r_0)^2 \\ &+ k_\theta(\theta - \theta_0)^2 \\ &+ k(1 + \cos(n\psi + \phi)) \end{aligned} \right\} \\ + \sum_{i>j} \left\{ \begin{aligned} &-U_{\min} \left[ \left( \frac{R_{\min}}{r_{ij}} \right)^{12} - 2 \left( \frac{R_{\min}}{r_{ij}} \right)^6 \right] \\ &+ \frac{Cq_i q_j}{\epsilon_0 r_{ij}} \end{aligned} \right\}$$



Lennard-Jones  
(van der Waals)

electrostatic

# MD force field: a approach



- Parametrize parts of the structure
- Assemble the parts

Example: a protein is a collection of 20 amino acids

Bonding cannot change(!)

# To run MD you'll need...

Computer (multi-core/GPU or cluster)

MD code (NAMD, Gromacs, Charmm, ...)

Input coordinates (typically .pdb): specifies initial state of the system

Protein structure file (typically .psf): specifies chemical bond information

Parameter file: provides atom-type specific values to interacting potentials

Configuration file (.namd or similar): run-type parameter, temperature, etc

Constraints or other simulation-specific files (typically .pdb)



Output: MD trajectory (.dcd file): describes how coordinates change in time

Analysis tools (VMD, python, etc.)

# What is MD good for?

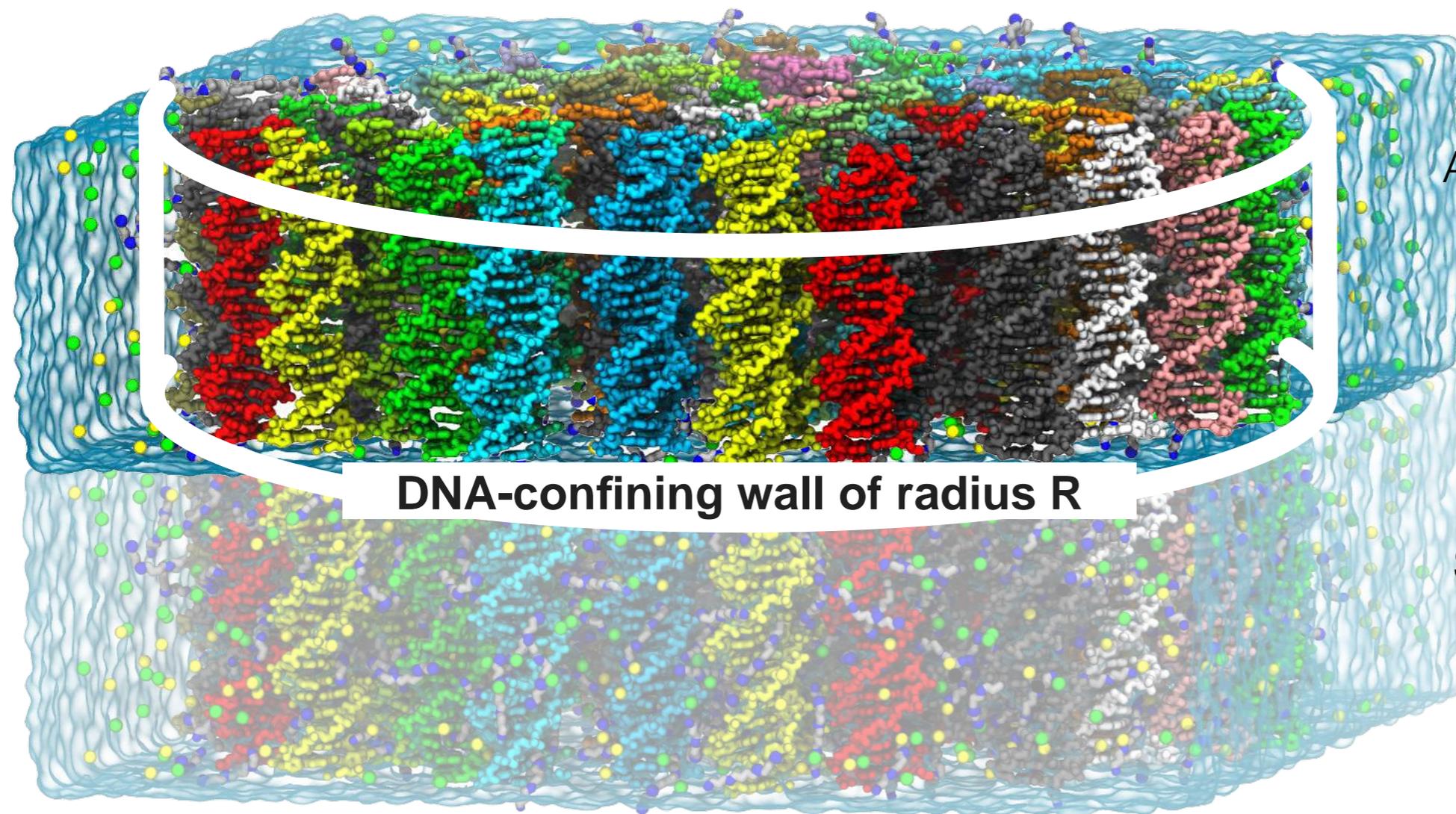
## Minimum requirements:

- Well-defined question
- Atomic-detail structure (or disordered state)
- Minimum chemistry
- Volume of 60 x 60 x 60 nm<sup>3</sup> or less
- Processes taking less than 100 microseconds

## Typical questions:

- What is the equilibrium structure in physiological environment?
- What is the free-energy cost of going from A to B?
- What is the path from A and B?
- How does the structure respond to external force or field?
- What is going on in my experiment?

# All-Atom Molecular Dynamics Simulation of DNA Condensates



Add **64 DNA** helices

Add **polyamine cations (+4)**

Add 150 mM **NaCl**

Add explicit **water**

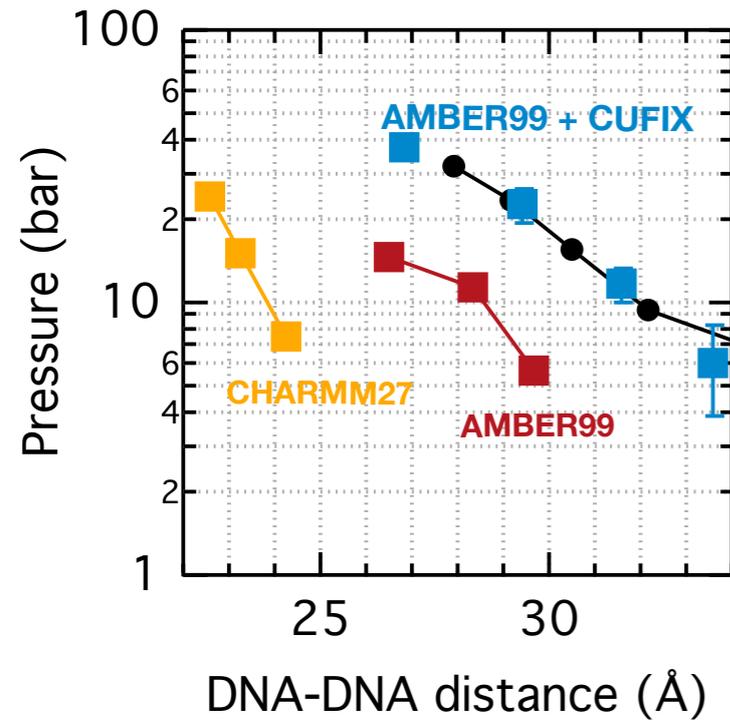
Apply a half-**harmonic wall potential** only to DNA

Solve the equation of motion ( $F=ma$ ) under **periodic boundary condition** in all directions

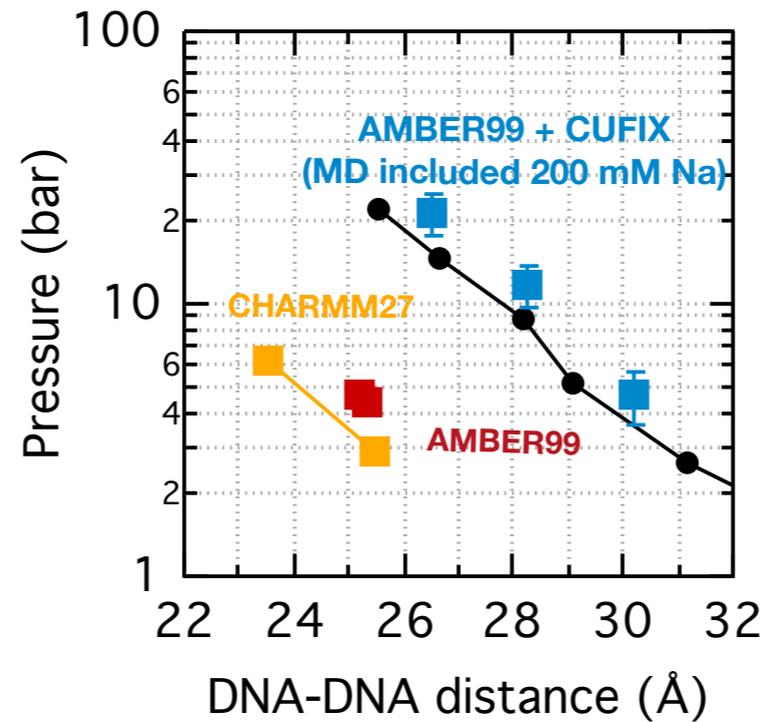
# CUFIX Improves Simulations of DNA Condensates

<http://bionano.physics.illinois.edu/CUFIX>

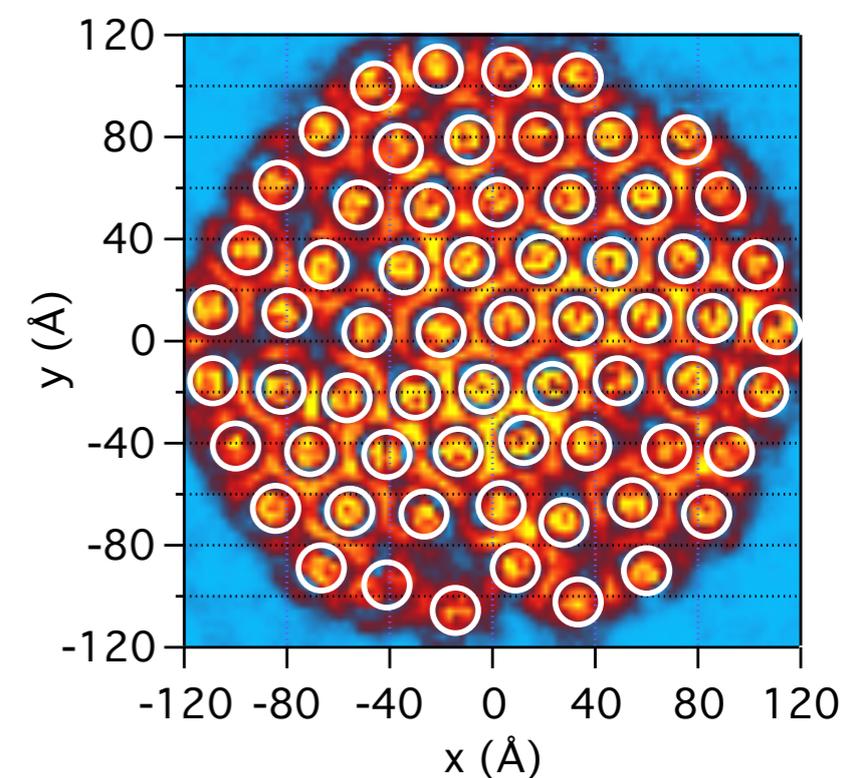
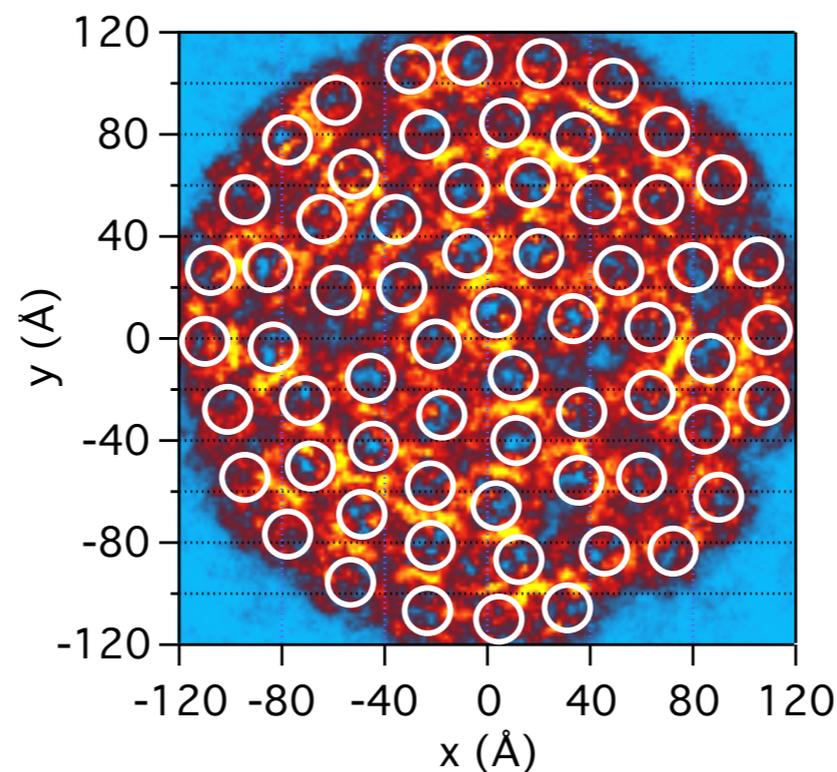
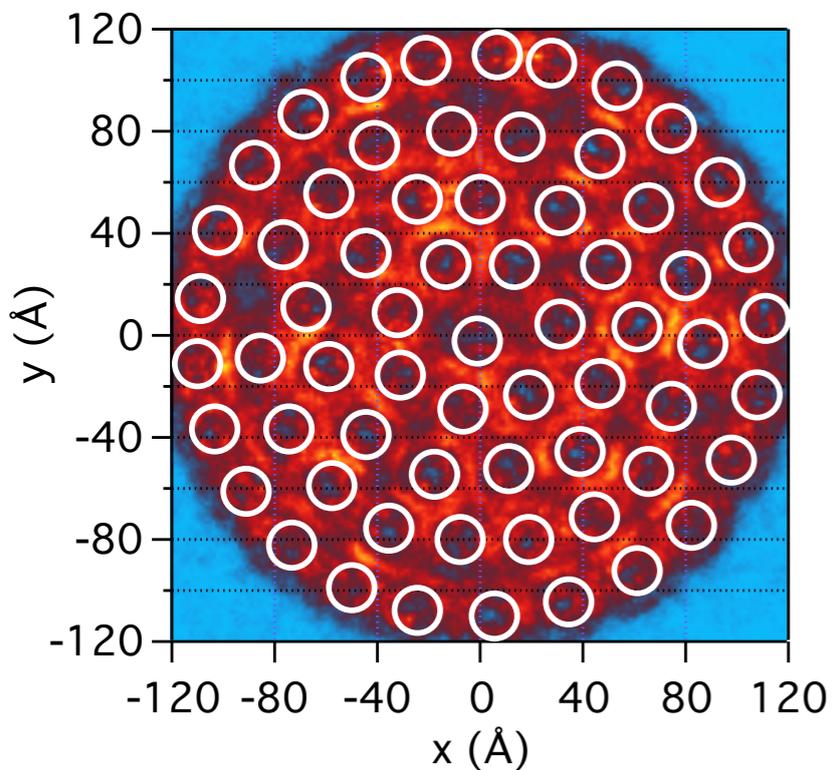
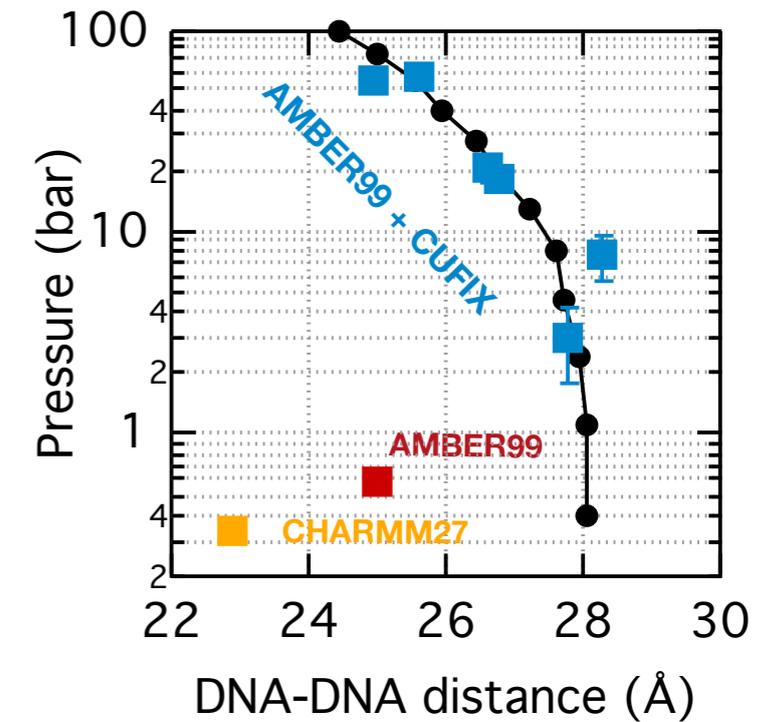
[Na] = 250 mM



[Mg] = 20 mM



[spermine] = 2 mM

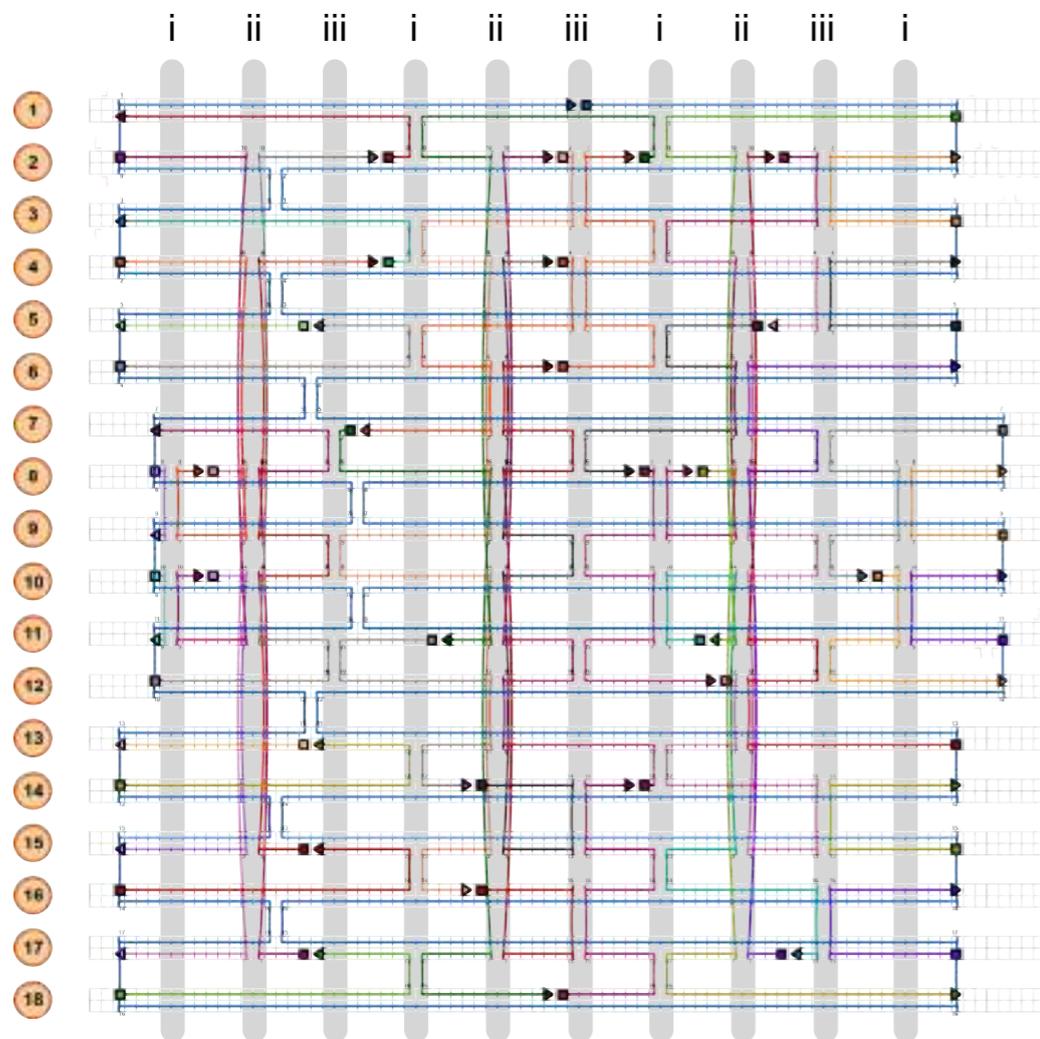


# From caDNAno to all-atom



Jejoong Yoo

## Cadnano design

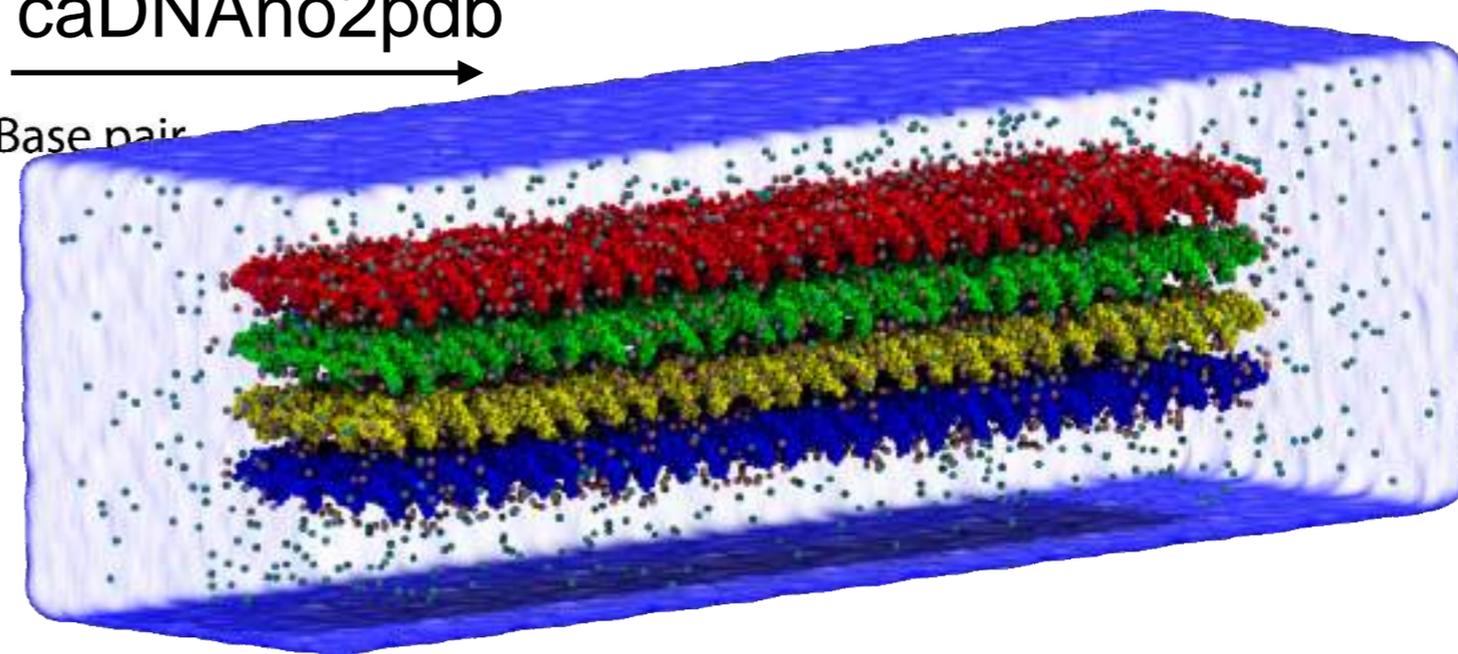


SM. Douglas, ..., WM. Shih,  
Nucleic Acids Res., 2009

## Slow water structure

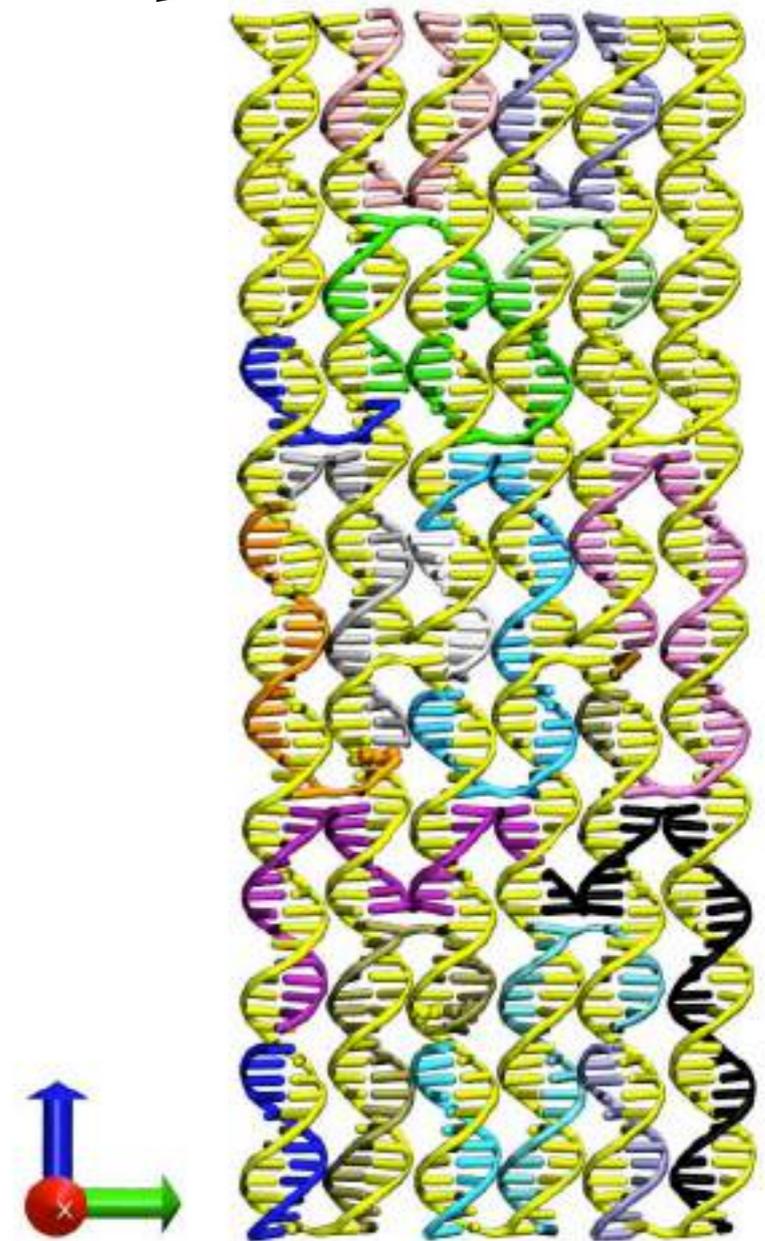
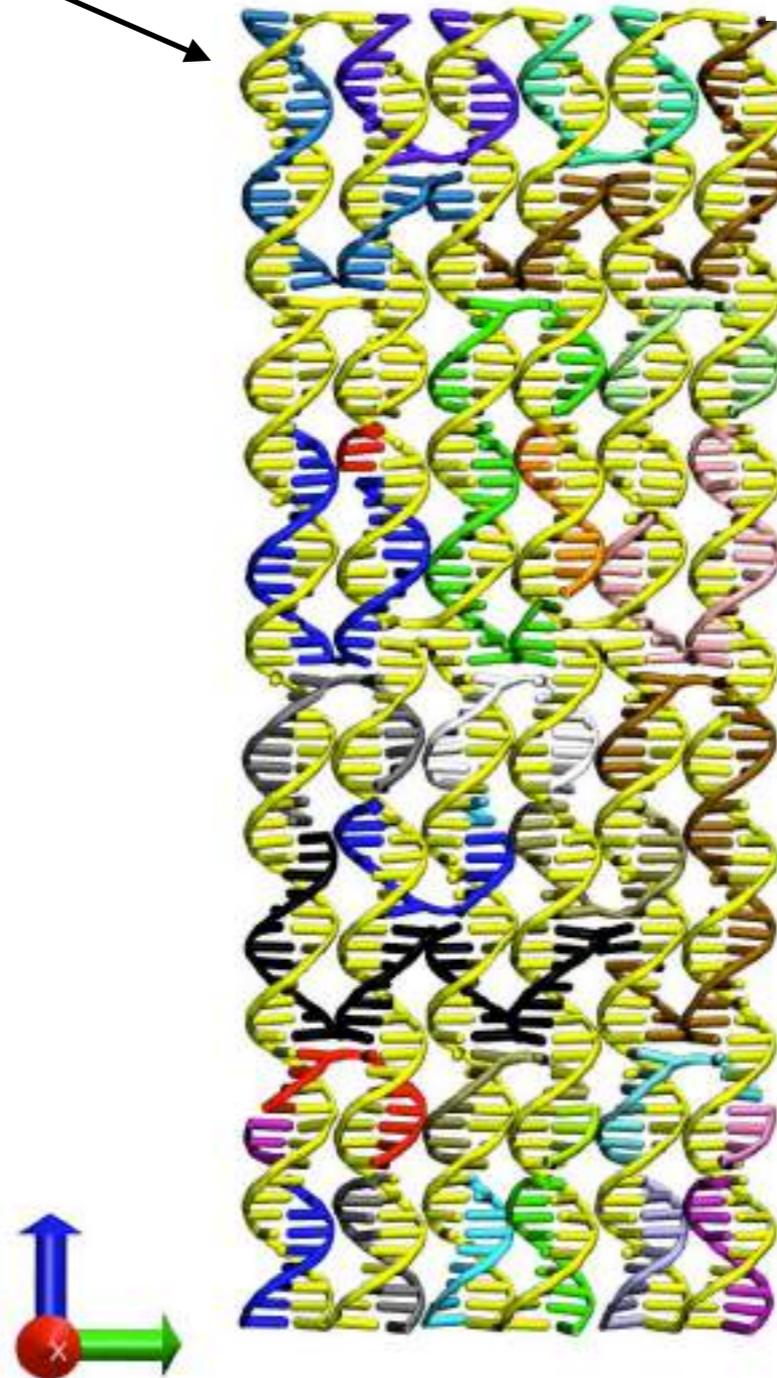
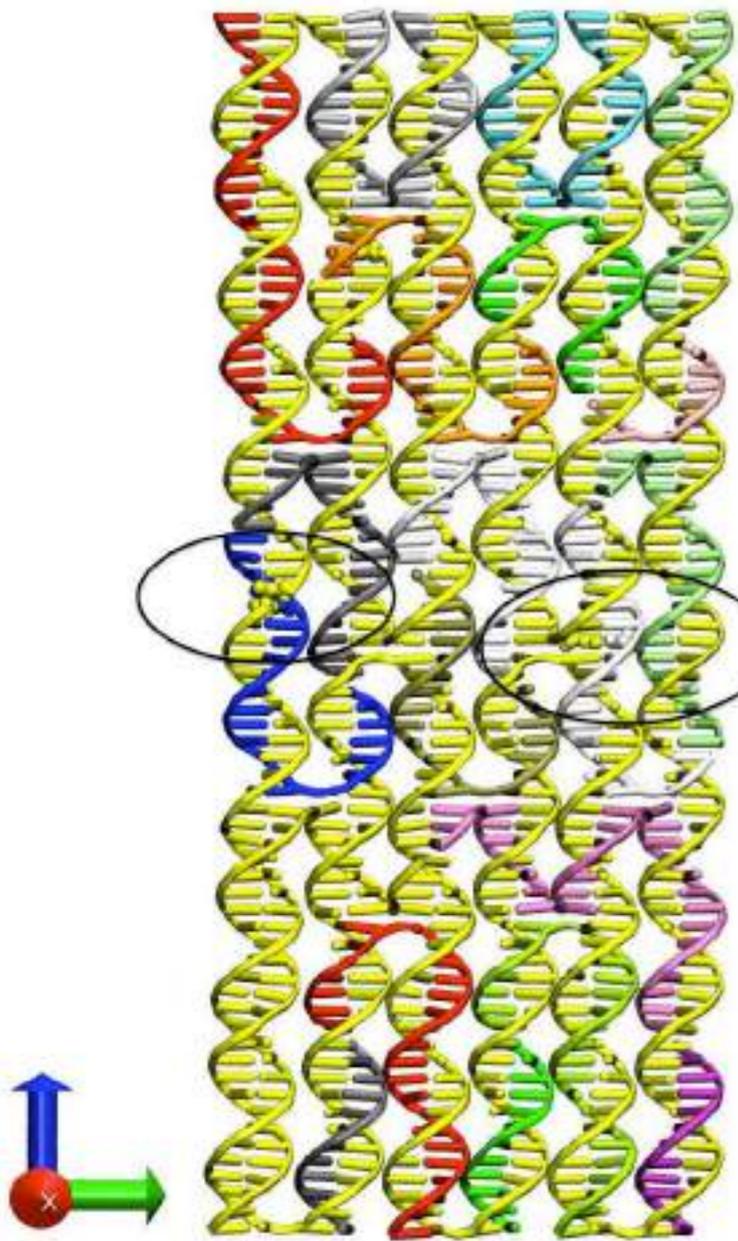
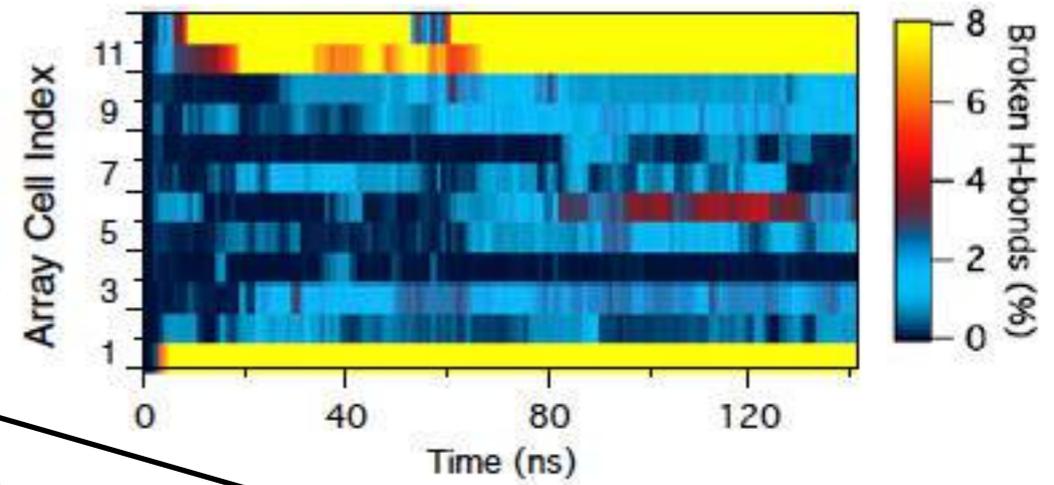
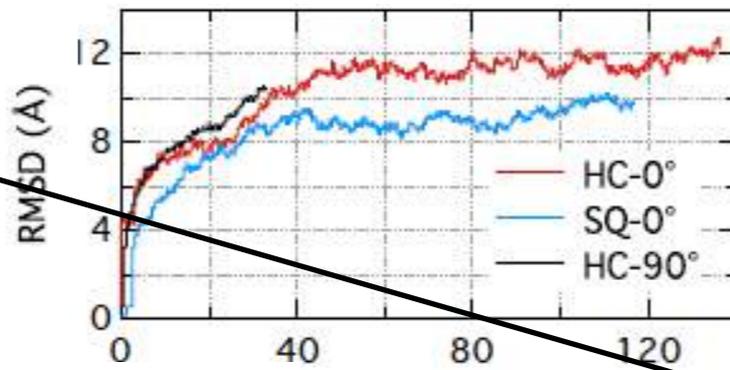
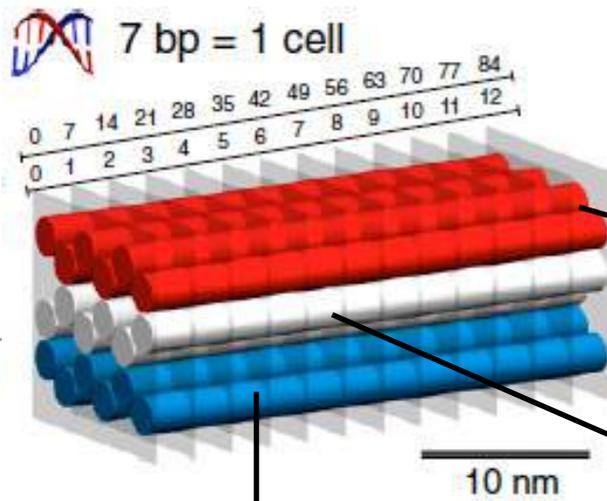
caDNAno2pdb

Base pair

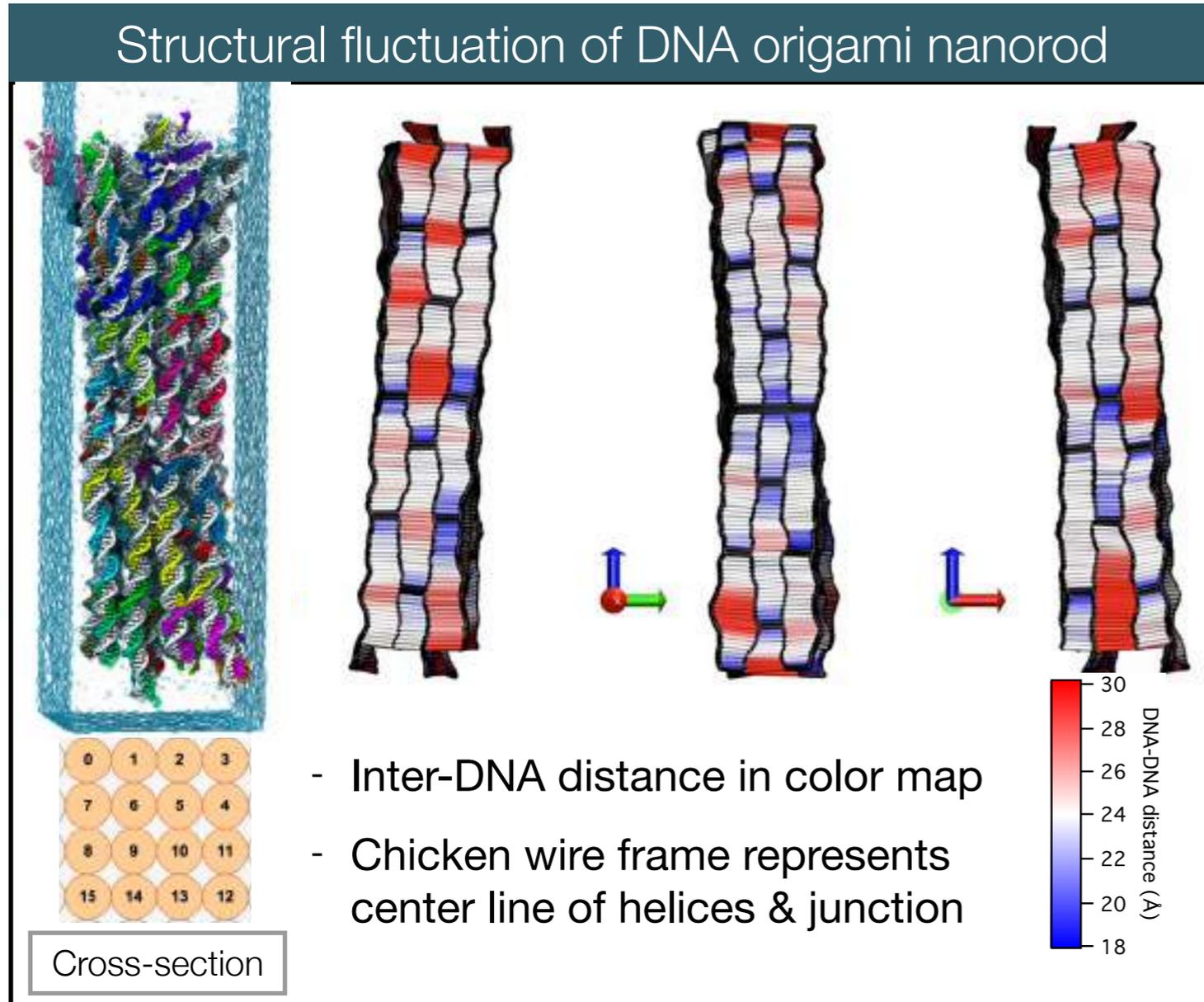


- \* CHARMM36 force field
- \* Explicit water
- \* [MgCl<sub>2</sub>] ~ 10 mM
- \* NAMD
- \* 1 to 3M atoms
- \* 500 to 1,000 CPUs

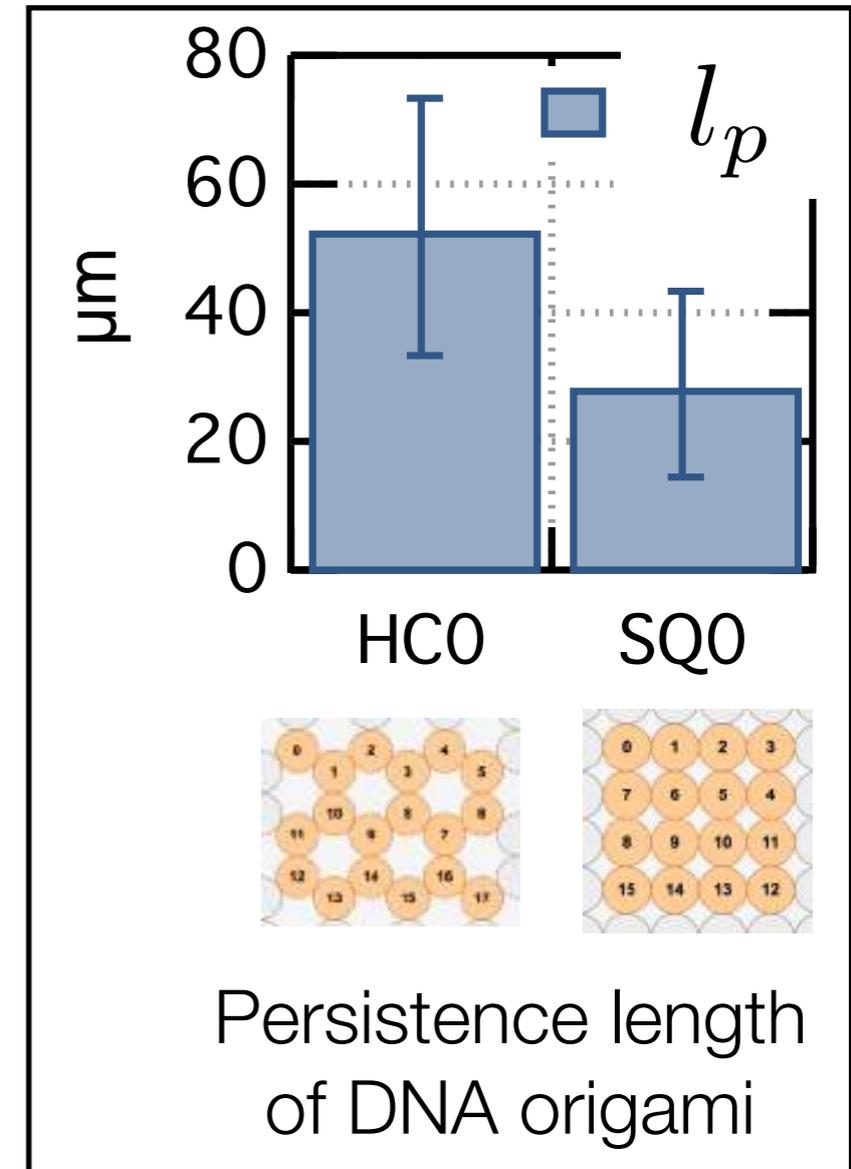
# Structural dynamics



# Structural fluctuations reveal local mechanical properties



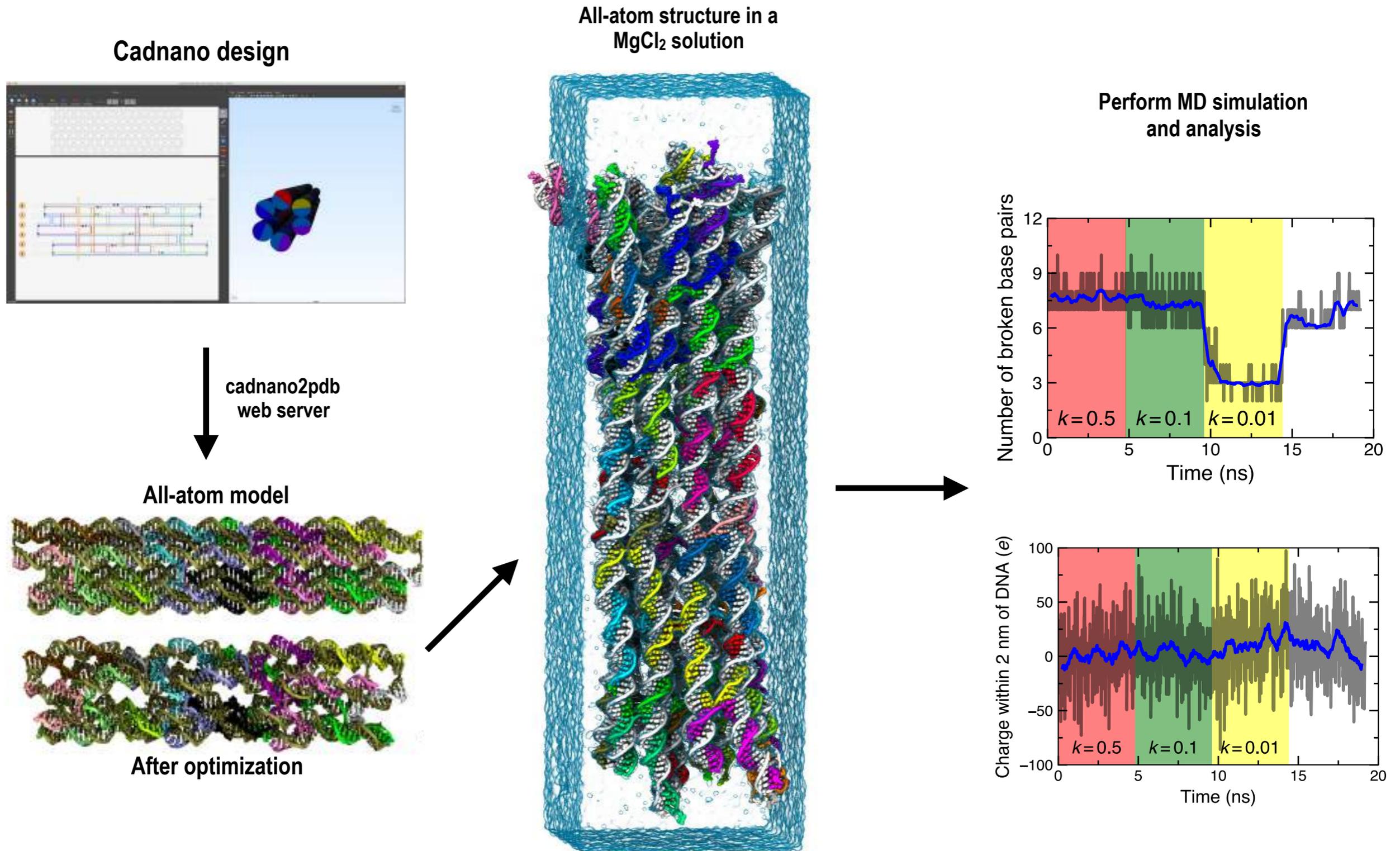
Yoo and Aksimentiev, PNAS 110:20099 (2013)



Simulations predict higher rigidity for honeycomb-lattice design

# A Practical Guide to DNA Origami Simulations Using NAMD

— Walk through the protocol for all-atom simulations of DNA origami using the NAMD package

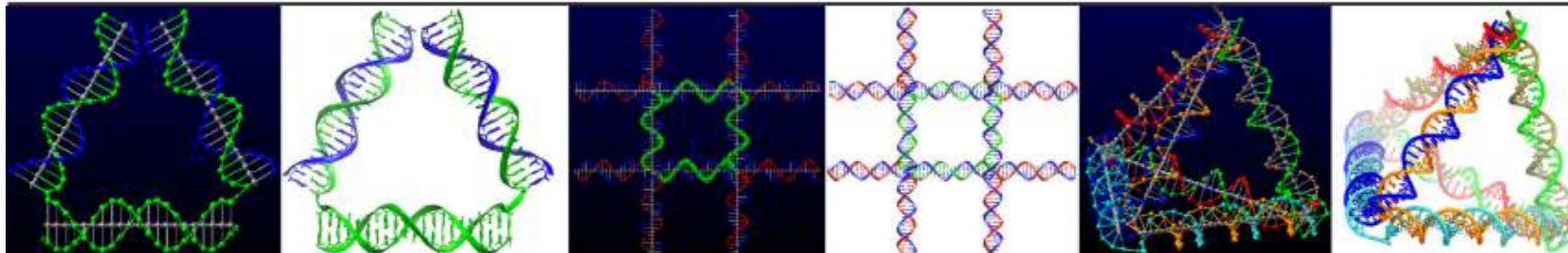
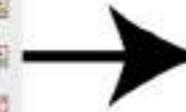
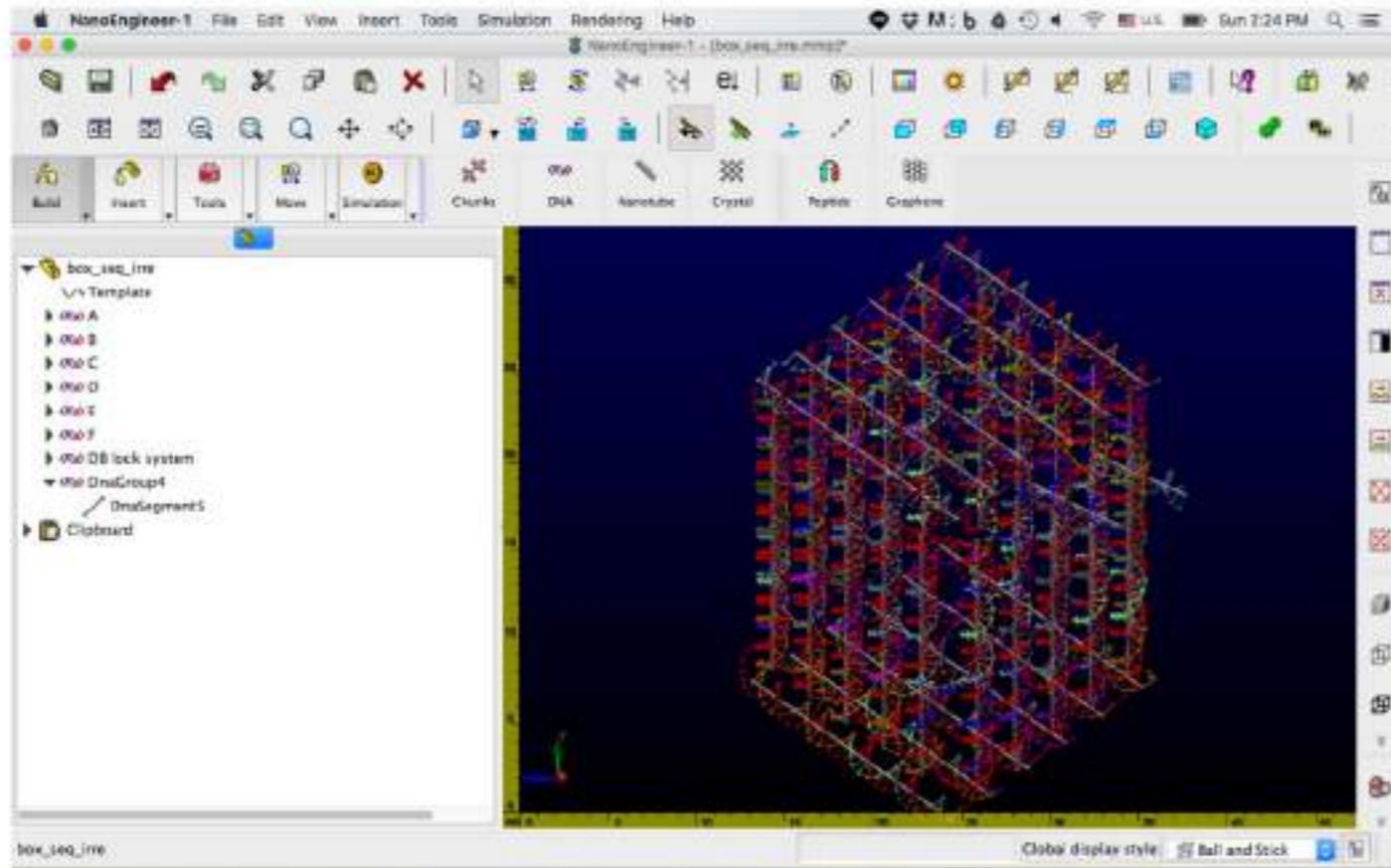


# NanoEngineer-1

All-atom PDB

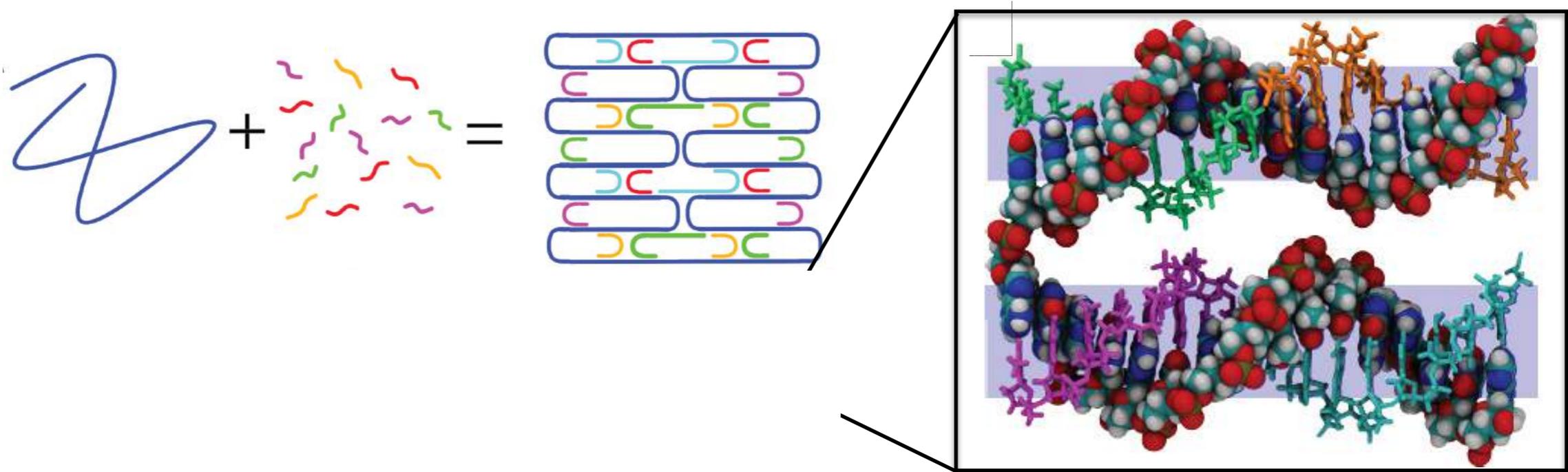


Chen-Yu Li

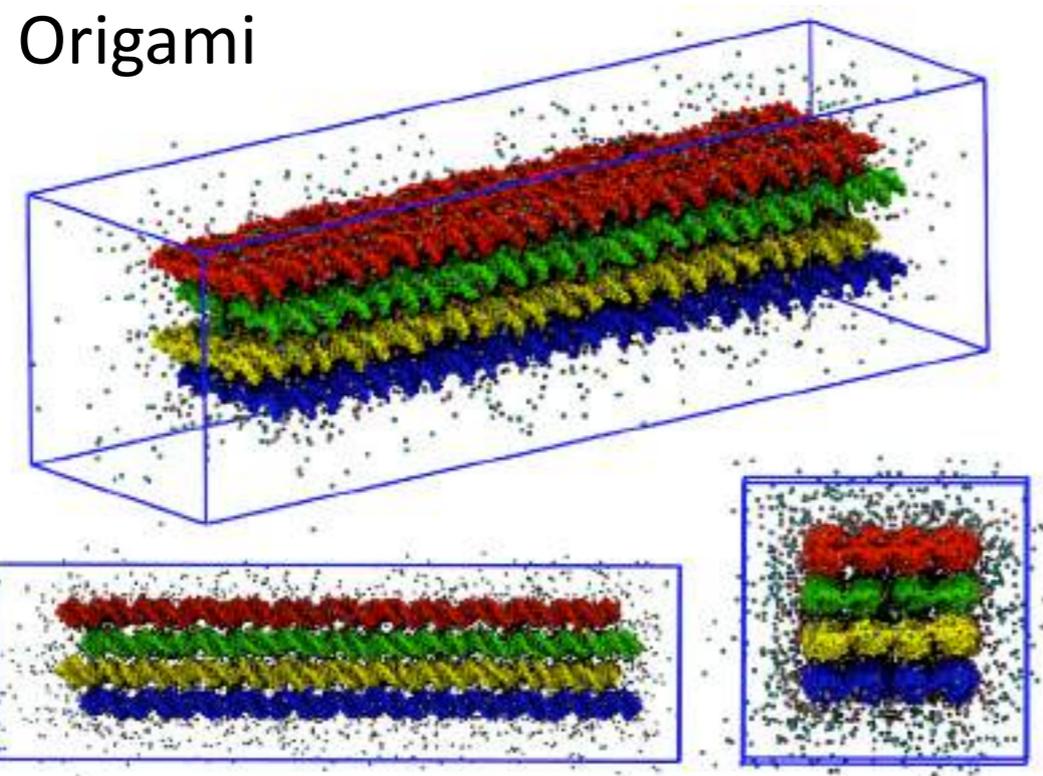


# Tiled DNA nanostructures

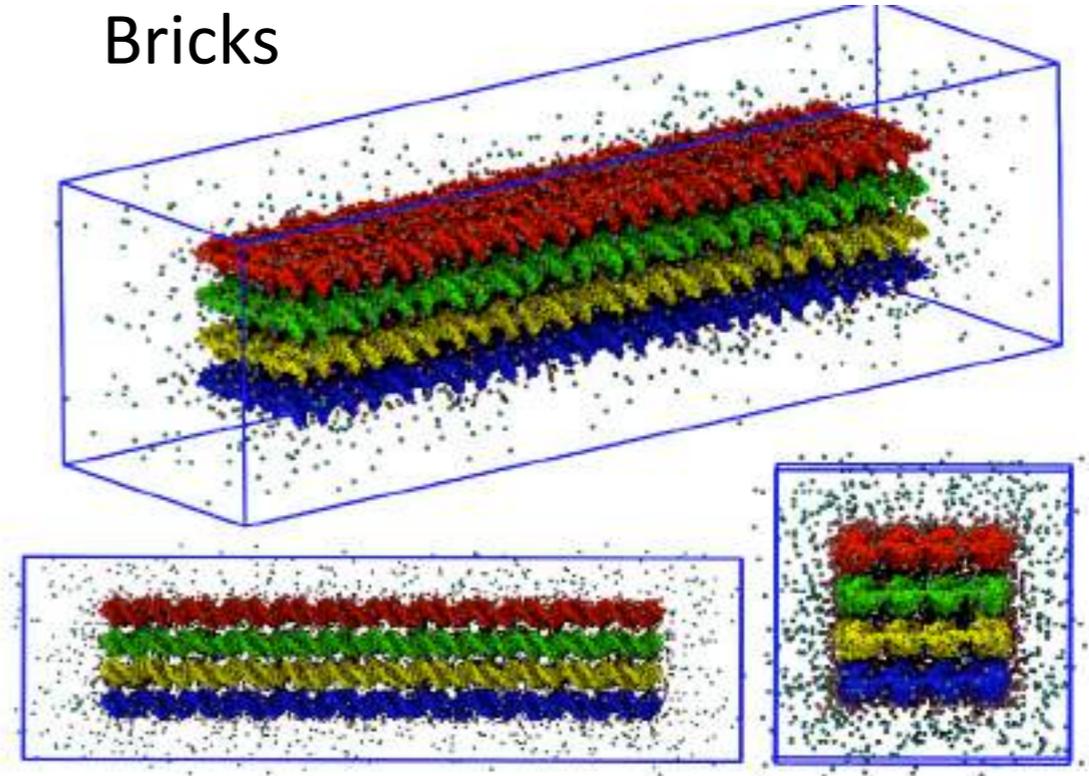
DNA  
Origami



Identical  
nucleotide  
sequence  
in both  
structures



Bricks

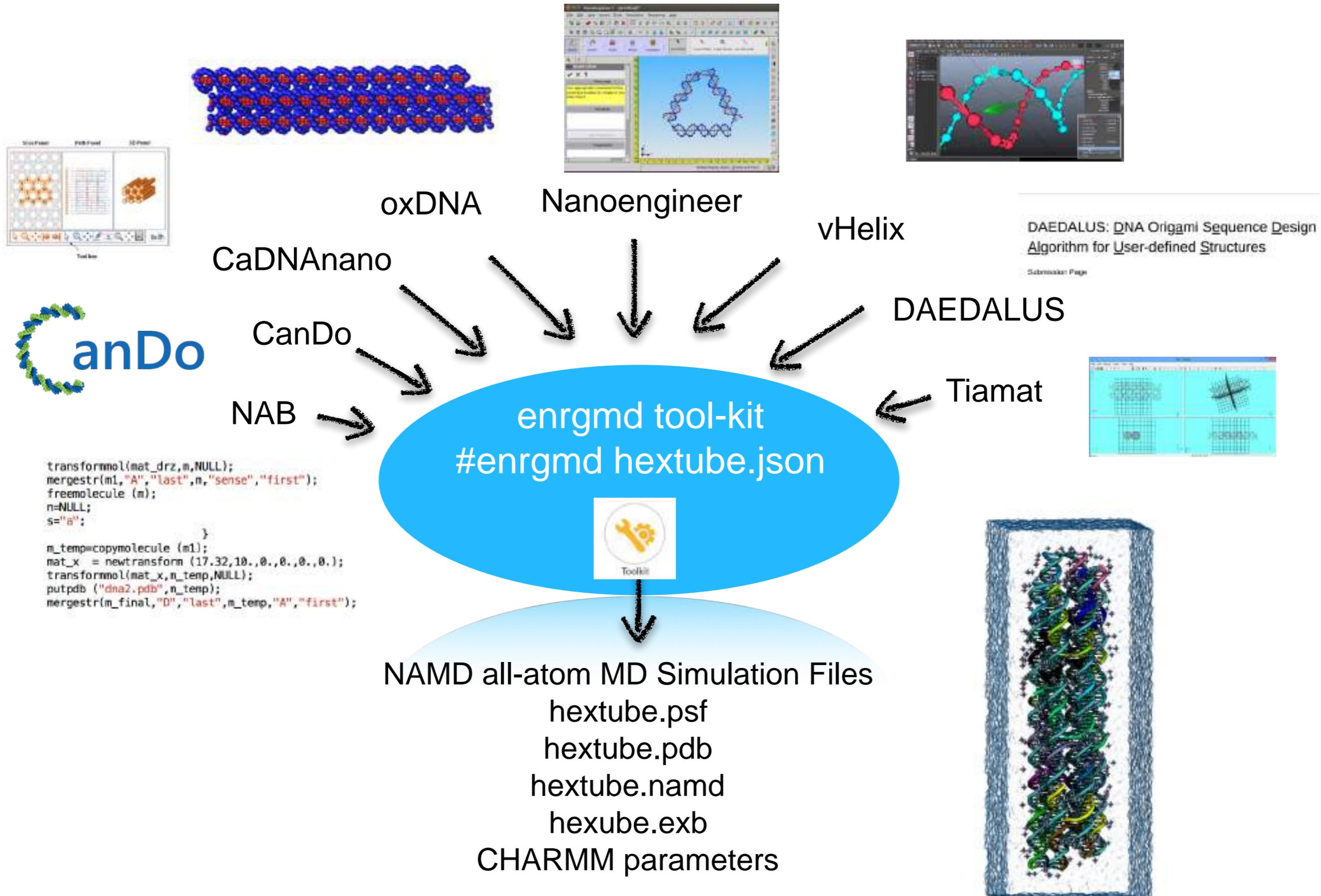


All-atom MD  
165 ns each

Slone et al., New J. Phys. 18:055012 (2016)

[nanohub.org/resources/legogen](http://nanohub.org/resources/legogen)

# ENRG MD tool kit : a universal all-atom structure converter



```

transformmol(mat_drz,m,NULL);
mergestr(m1,"A","last",n,"sense","first");
freemolecule (m);
n=NULL;
s="a";
}
n_temp=copymolecule (m1);
mat_x = newtransform (17.32,10.,0.,0.,0.,0.);
transformmol(mat_x,n_temp,NULL);
putpdb ("dna2.pdb",n_temp);
mergestr(m_final,"D","last",n_temp,"A","first");
    
```

DAEDALUS: DNA Origami Sequence Design Algorithm for User-defined Structures  
Submitter Page



# Cryo-EM reconstruction versus all-atom simulation

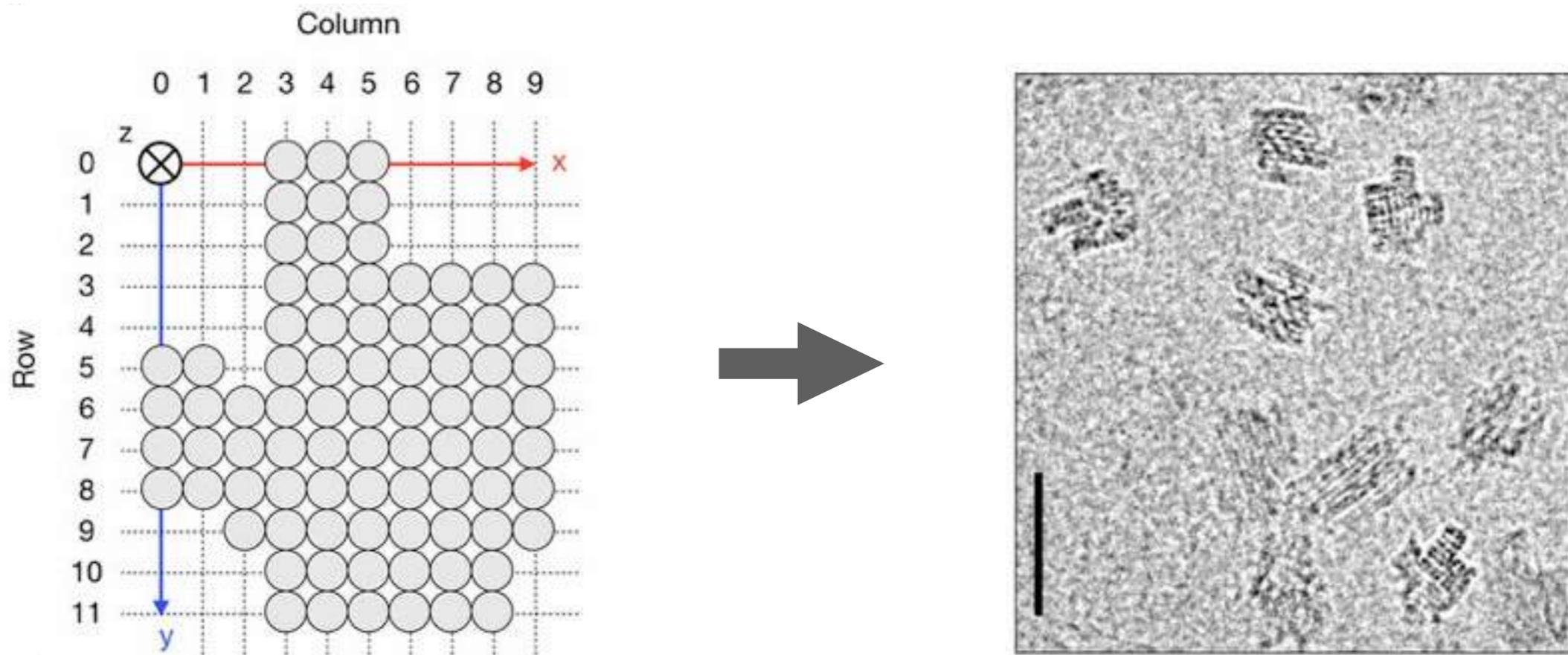


High-resolution cryo-electron microscope



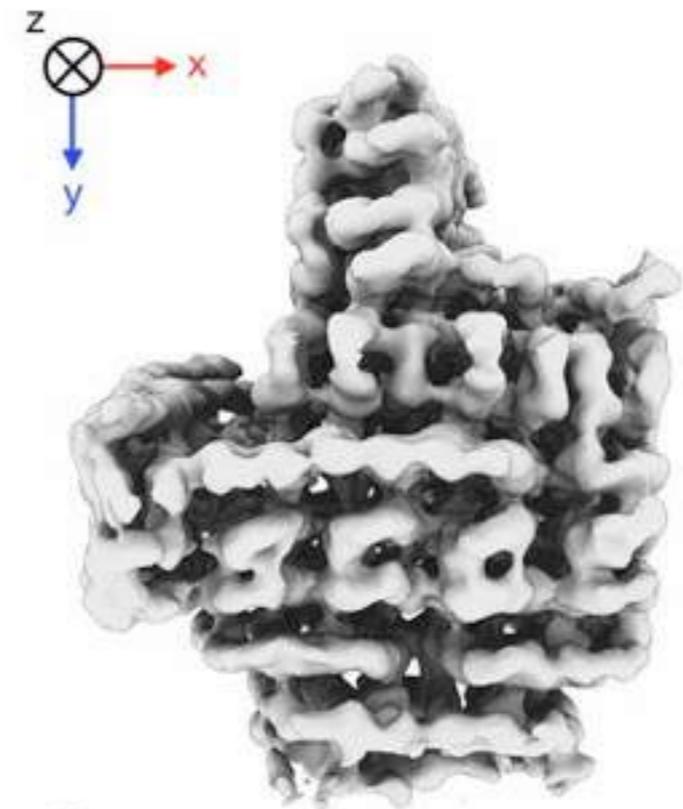
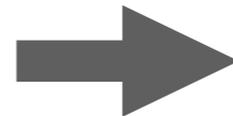
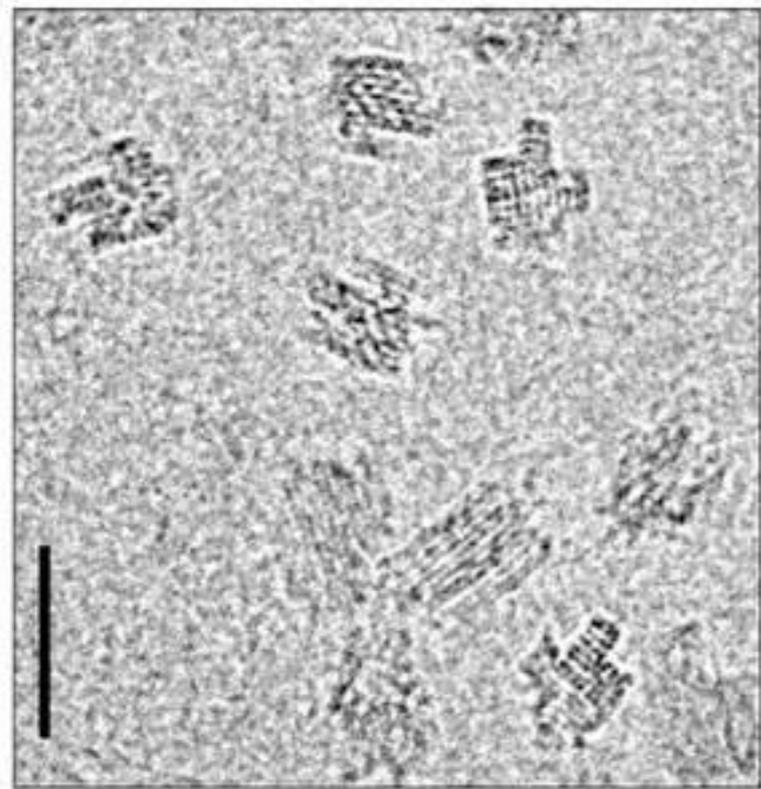
Petascale computer system

# Cryo-EM reconstruction versus all-atom simulation



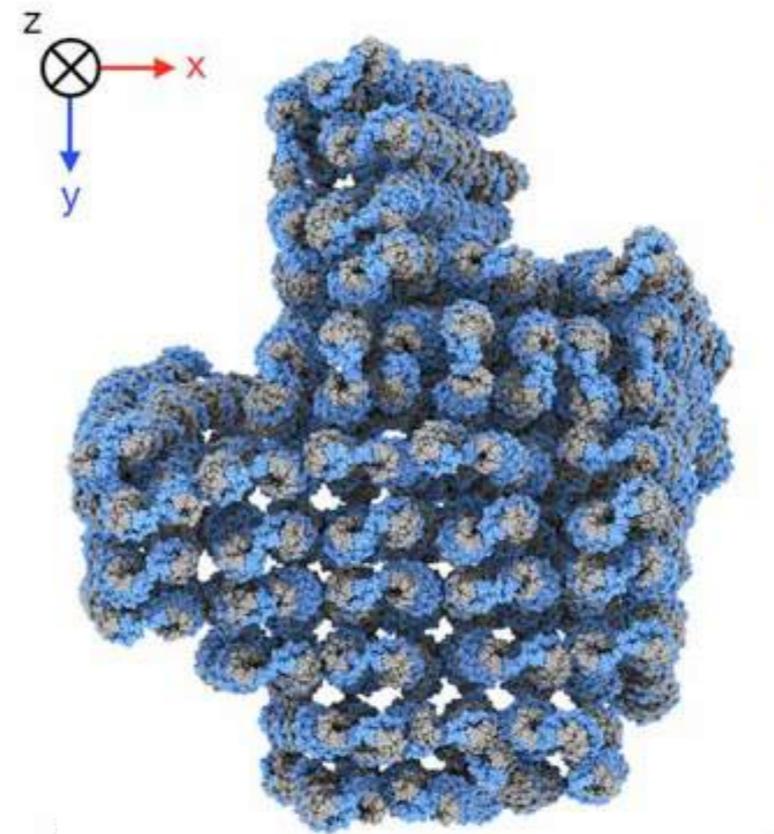
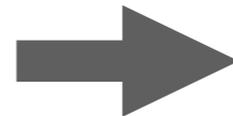
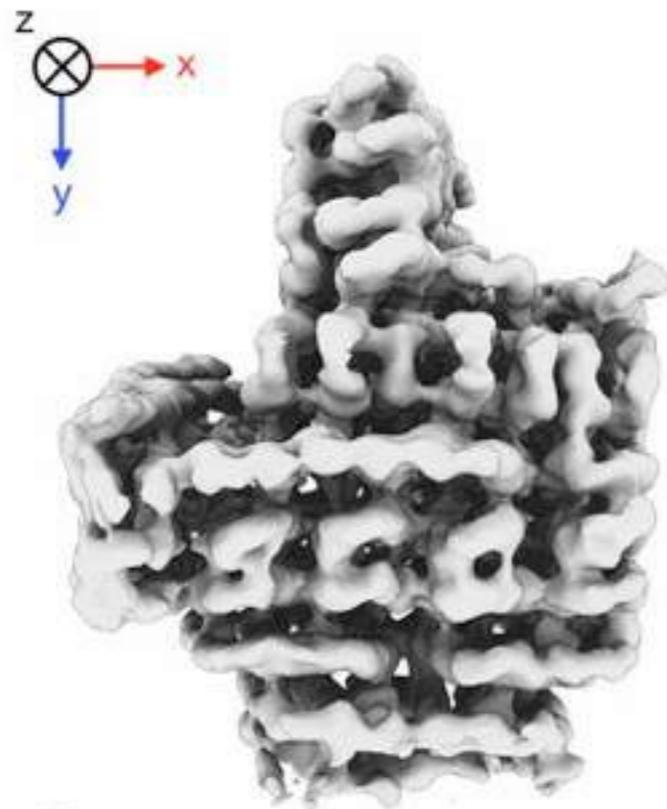
Bai *et al*, PNAS 109:20012 (2012)

# Cryo-EM reconstruction versus all-atom simulation



Bai *et al*, PNAS 109:20012 (2012)

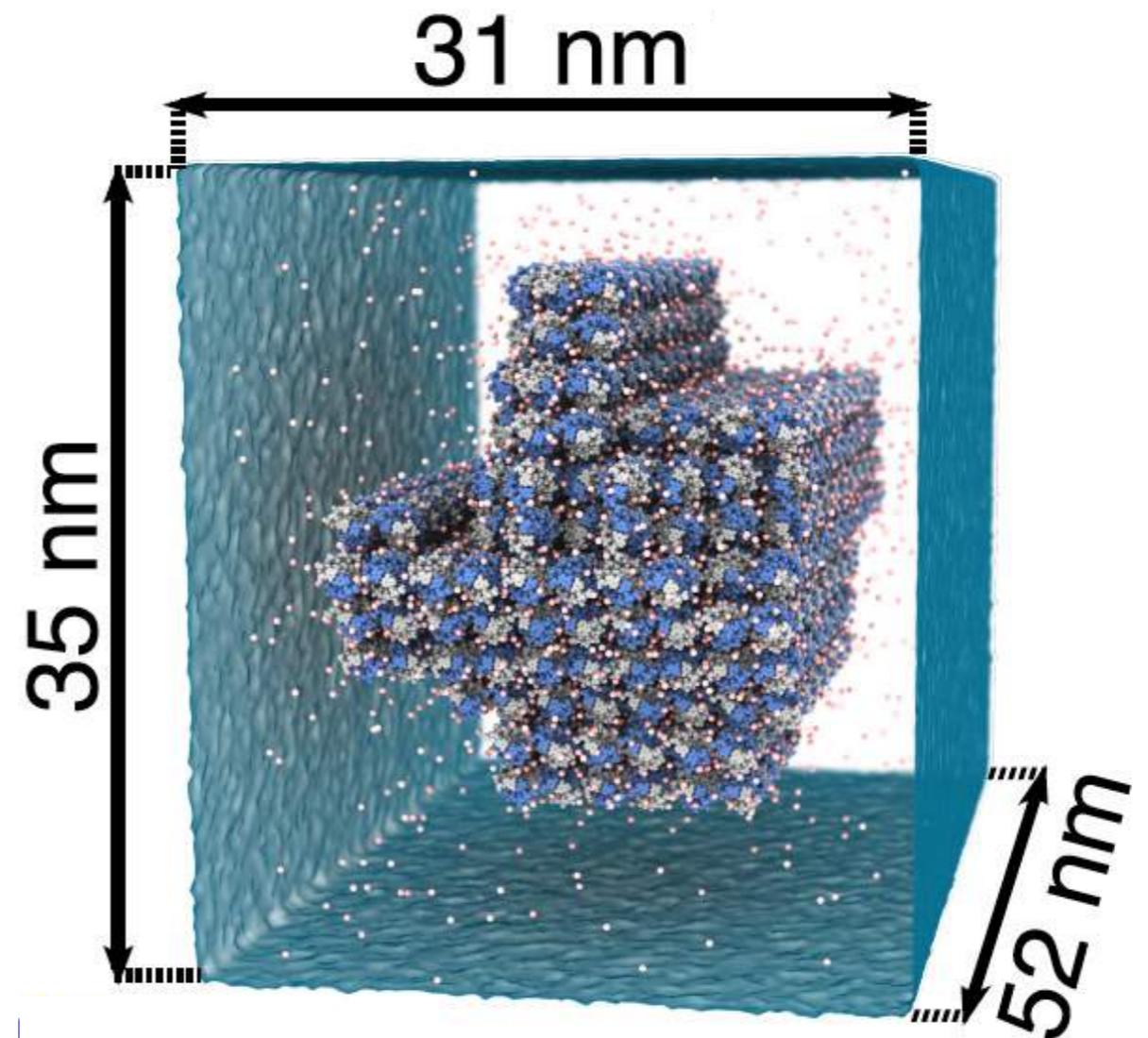
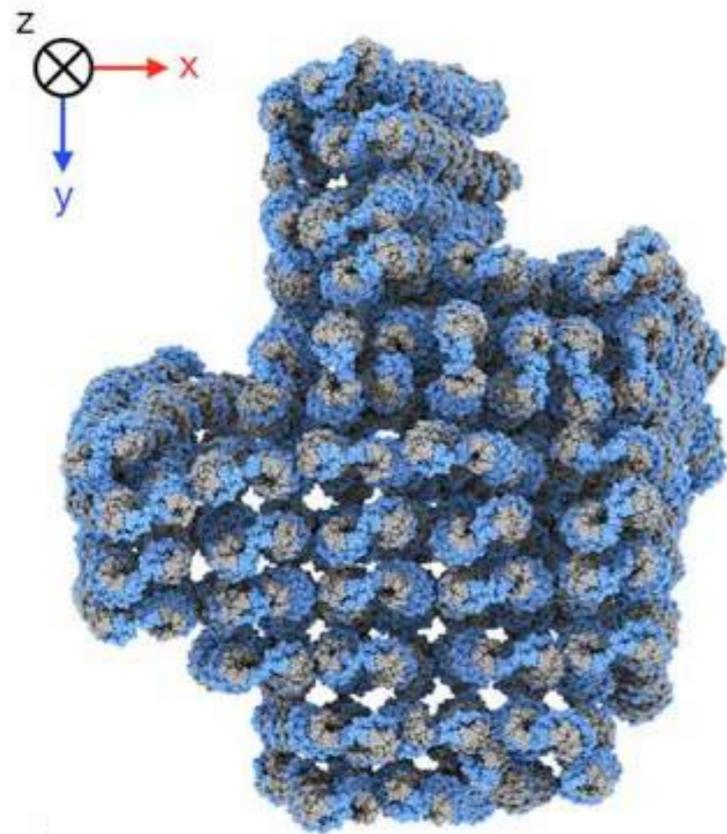
# Cryo-EM reconstruction versus all-atom simulation



Pseudo-atomic model

Bai *et al*, PNAS 109:20012 (2012)

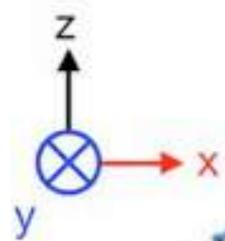
# MD simulation of the cryo-EM object starting from a caDNAano design



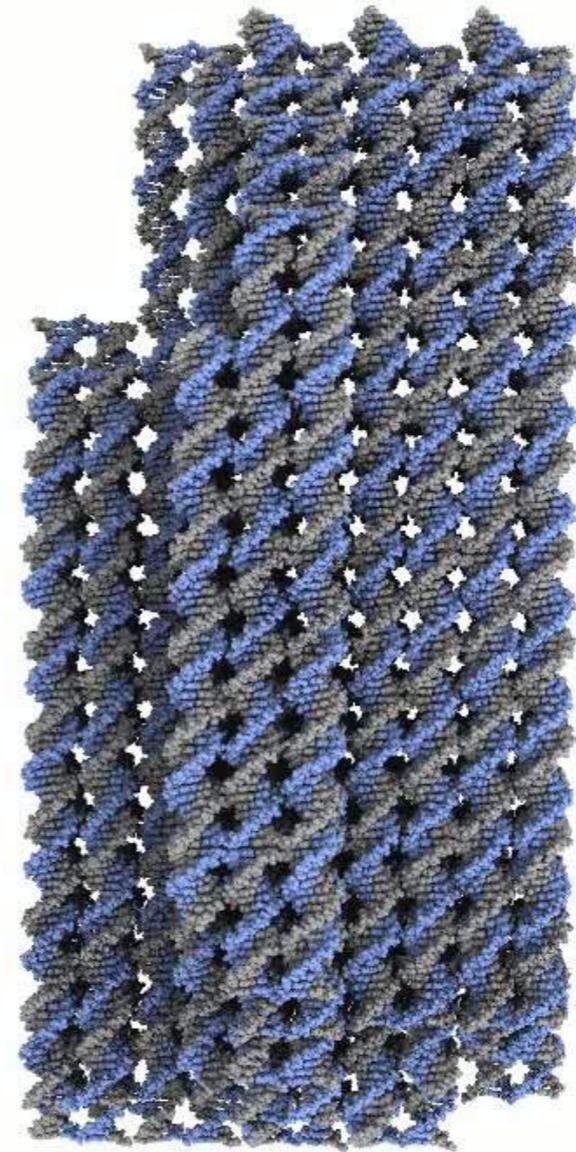
Bai *et al*, PNAS 109:20012 (2012)

7M atom solvated model  
~200 ns MD trajectory

# MD simulation of the cryo-EM object starting from a caDNAano design

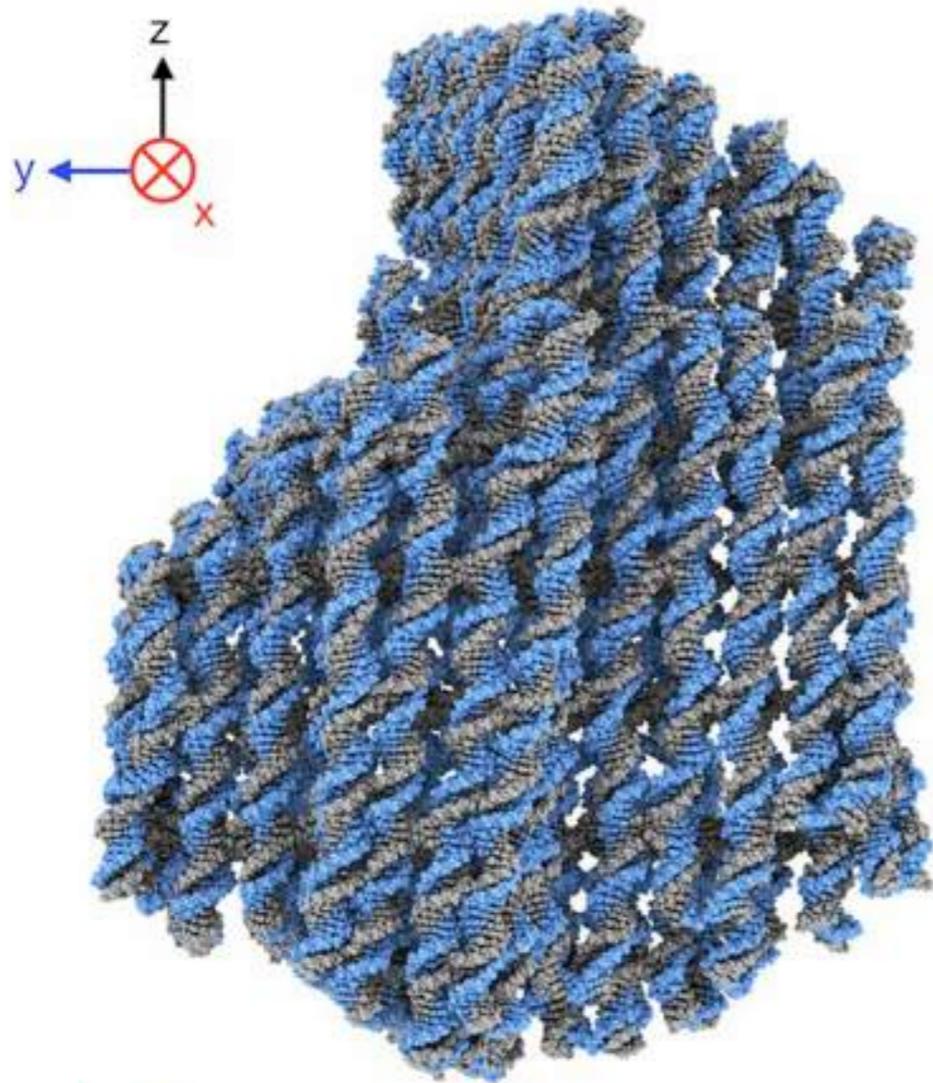


Bai *et al*, PNAS 109:20012 (2012)

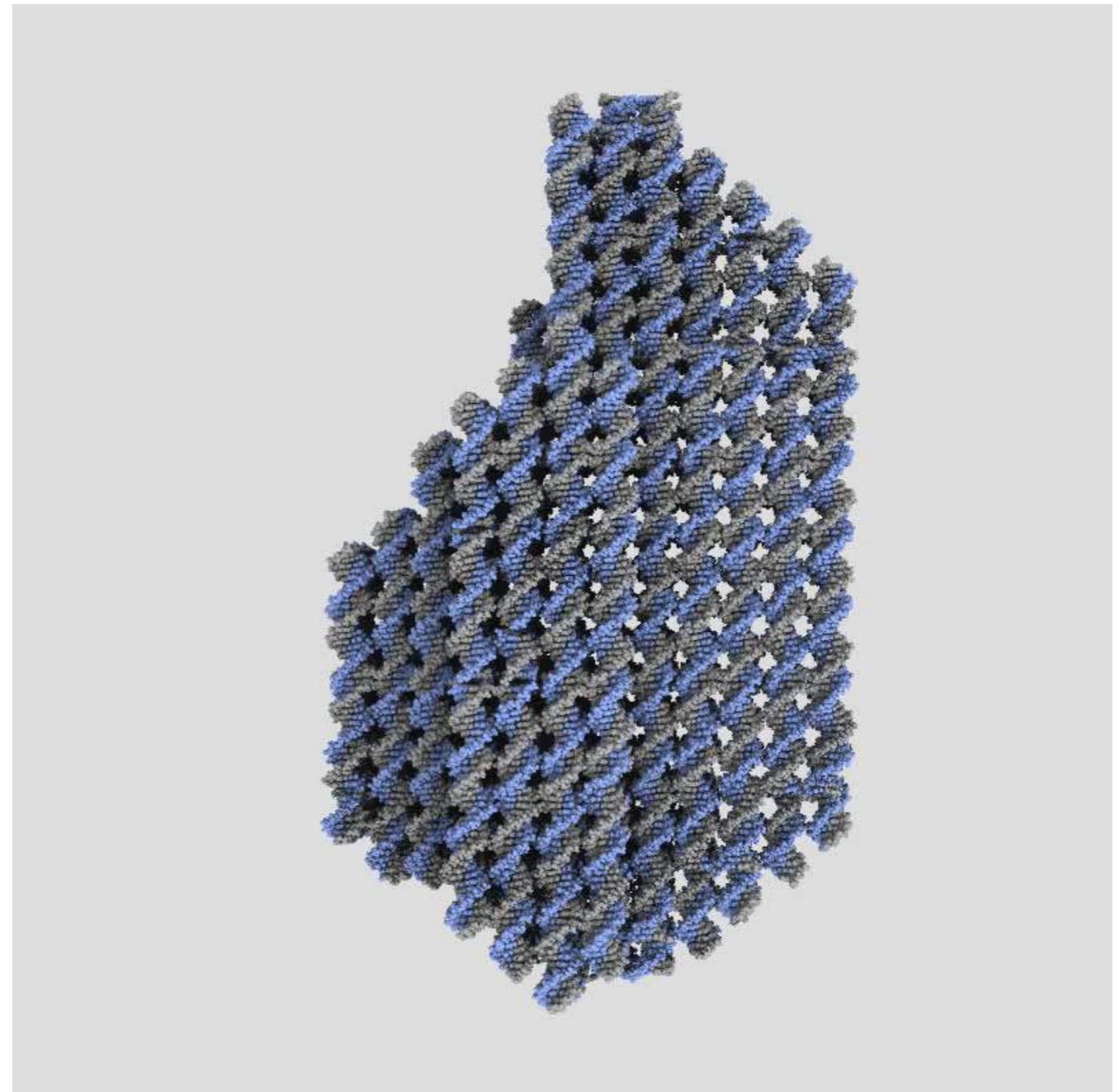


7M atom solvated model  
~200 ns MD trajectory

# MD simulation of the cryo-EM object starting from a caDNAano design

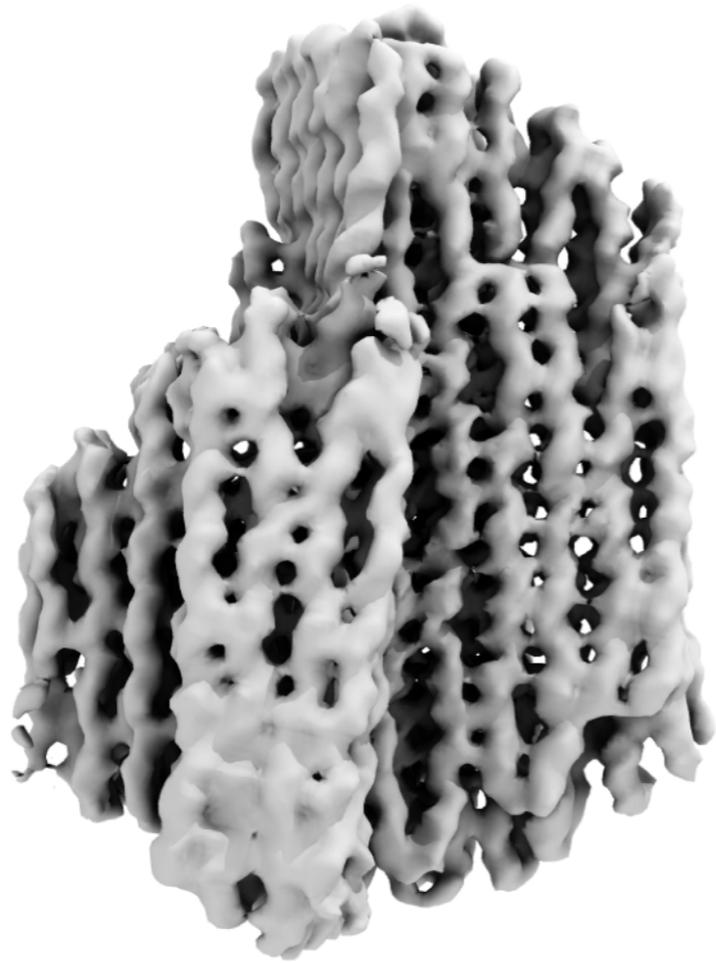


Bai *et al*, PNAS 109:20012 (2012)

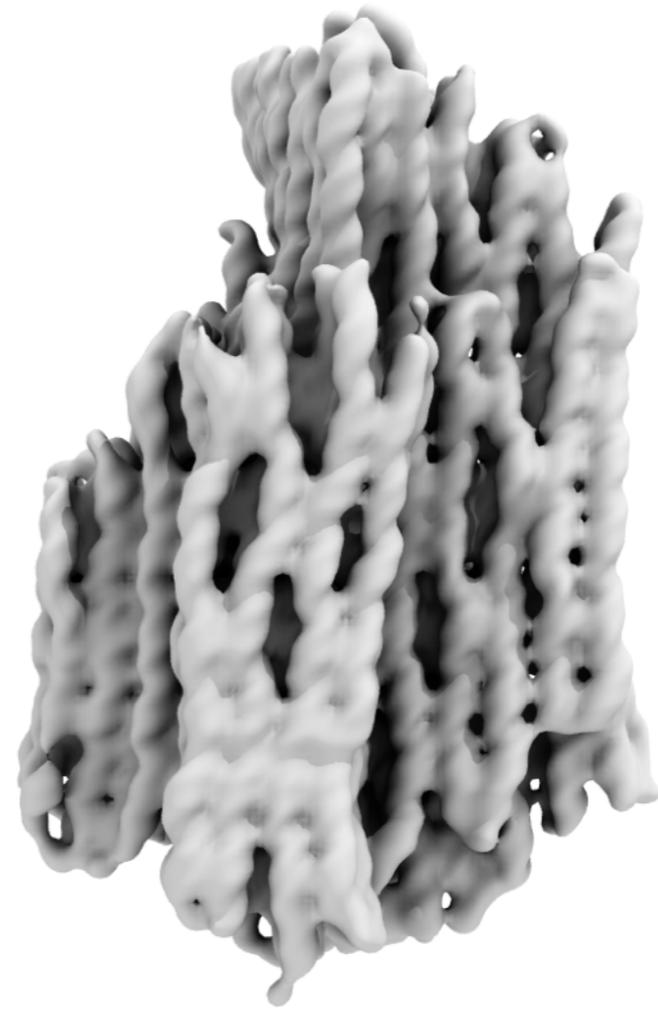


7M atom solvated model  
~200 ns MD trajectory

# Electron density maps



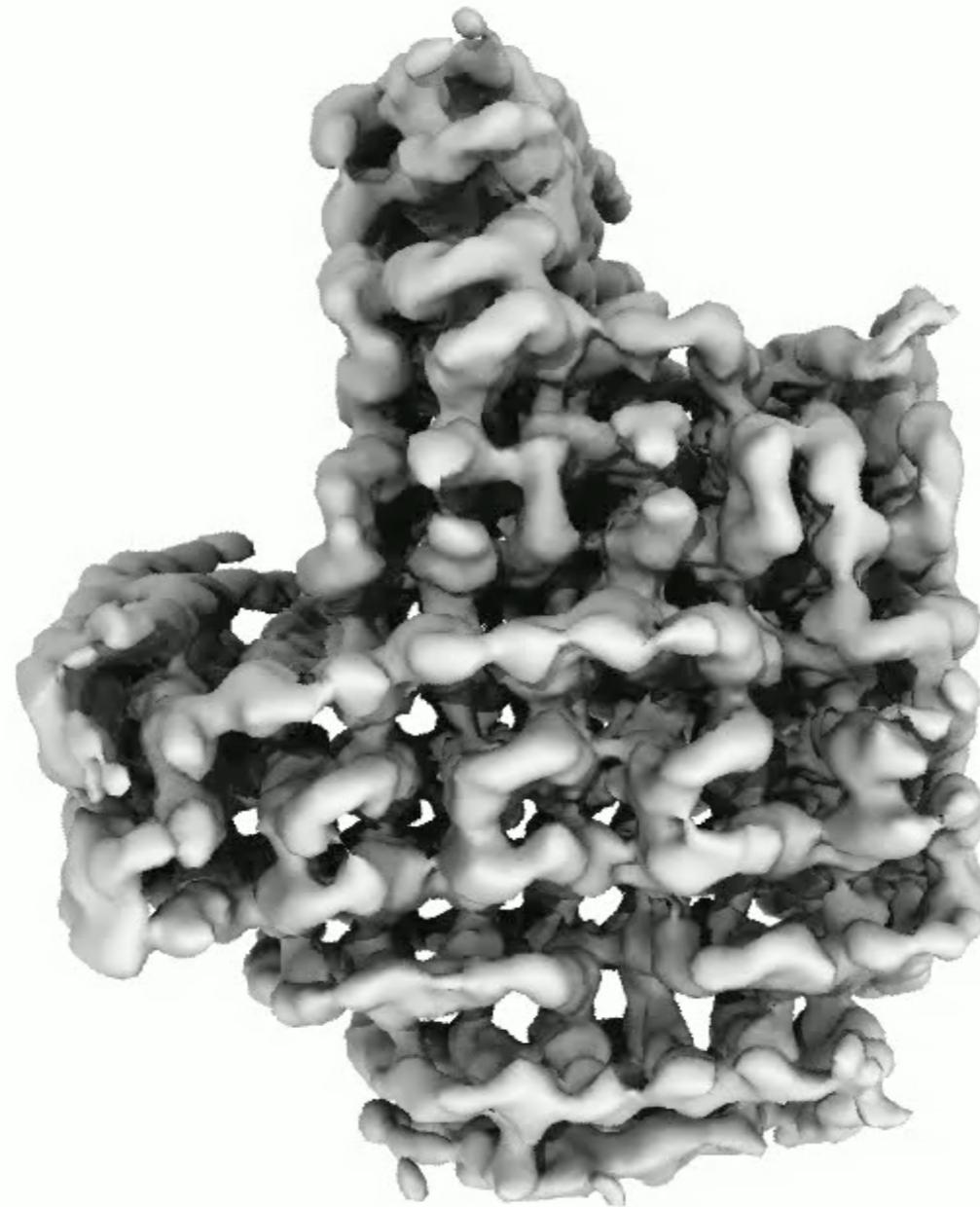
Cryo-EM reconstruction



All-atom MD simulation

# Comparison with experiment

Maffeo, Yoo & Aksimentiev, *NAR* 44: 3013 (2016)



EM density

pseudo-atomic model

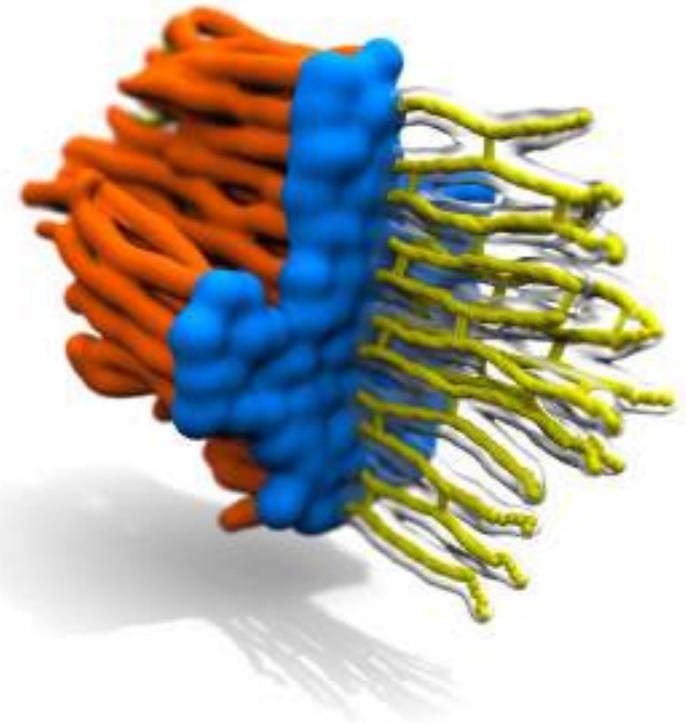
simulation

# Making images and animations of DNA nanostructures with VMD

Images from a single structure

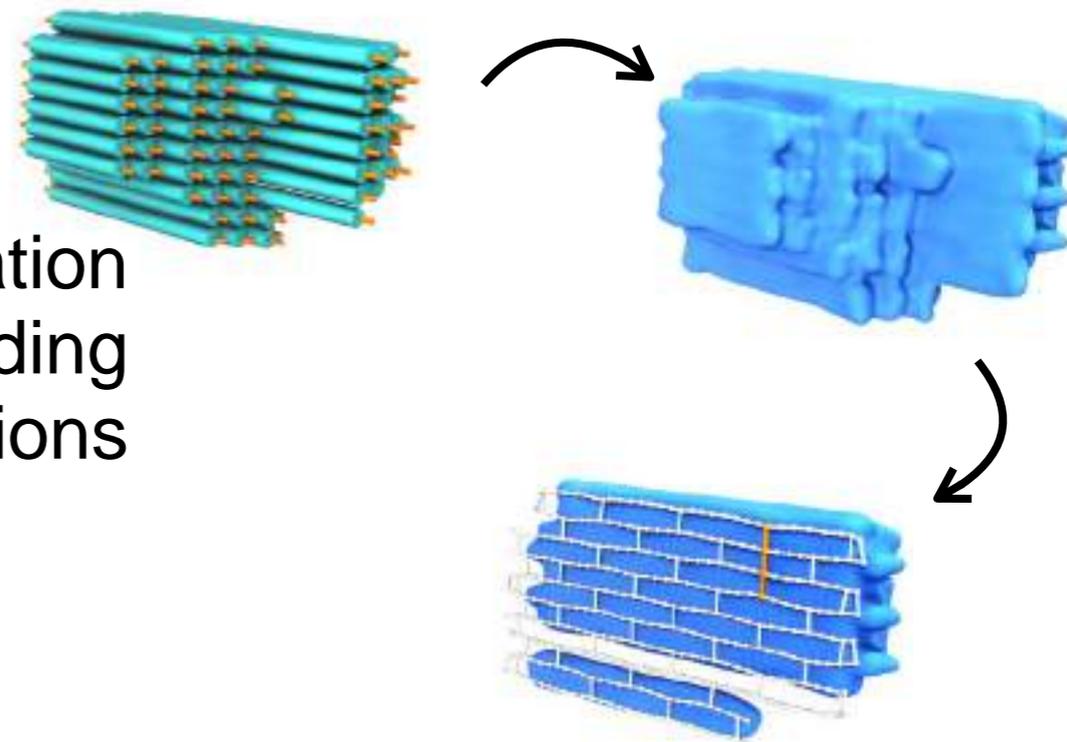


Image with multiple representations

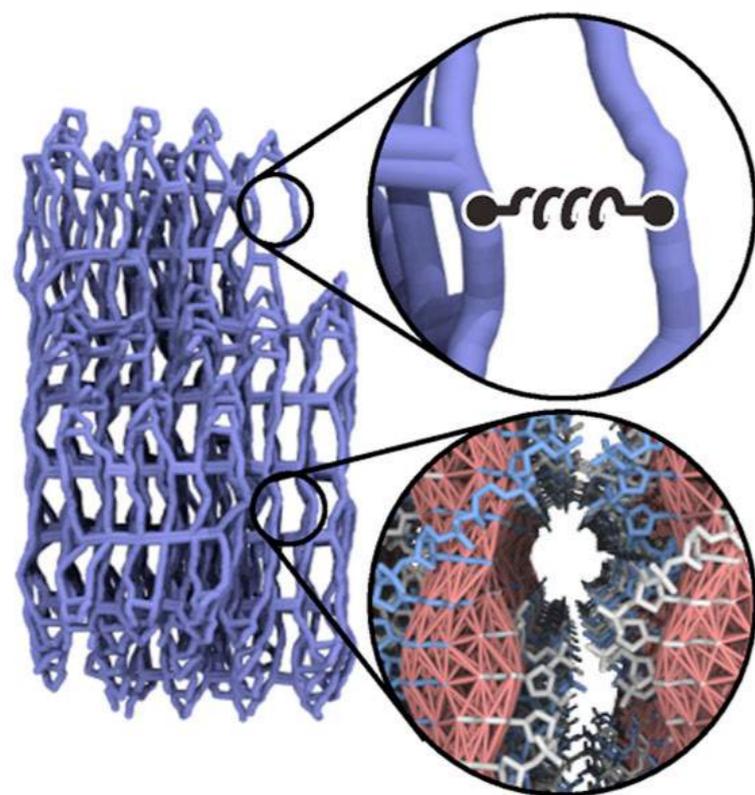


Animation based on a trajectory

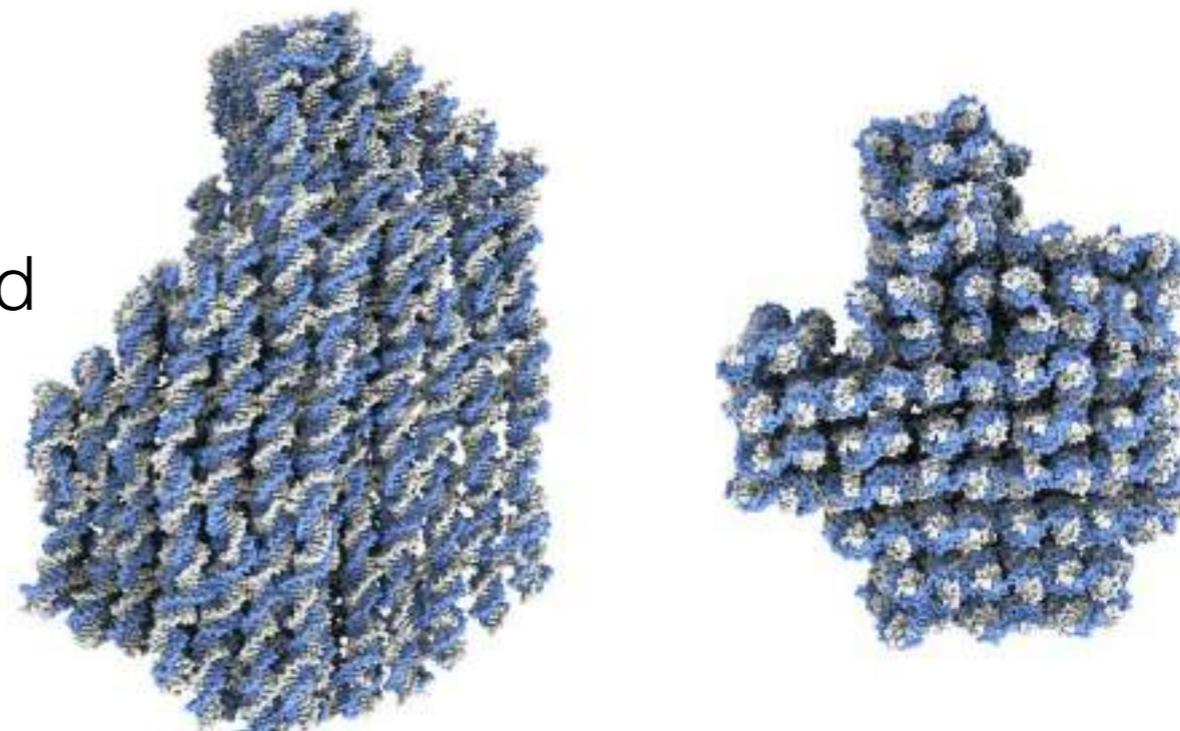
Animation including transitions



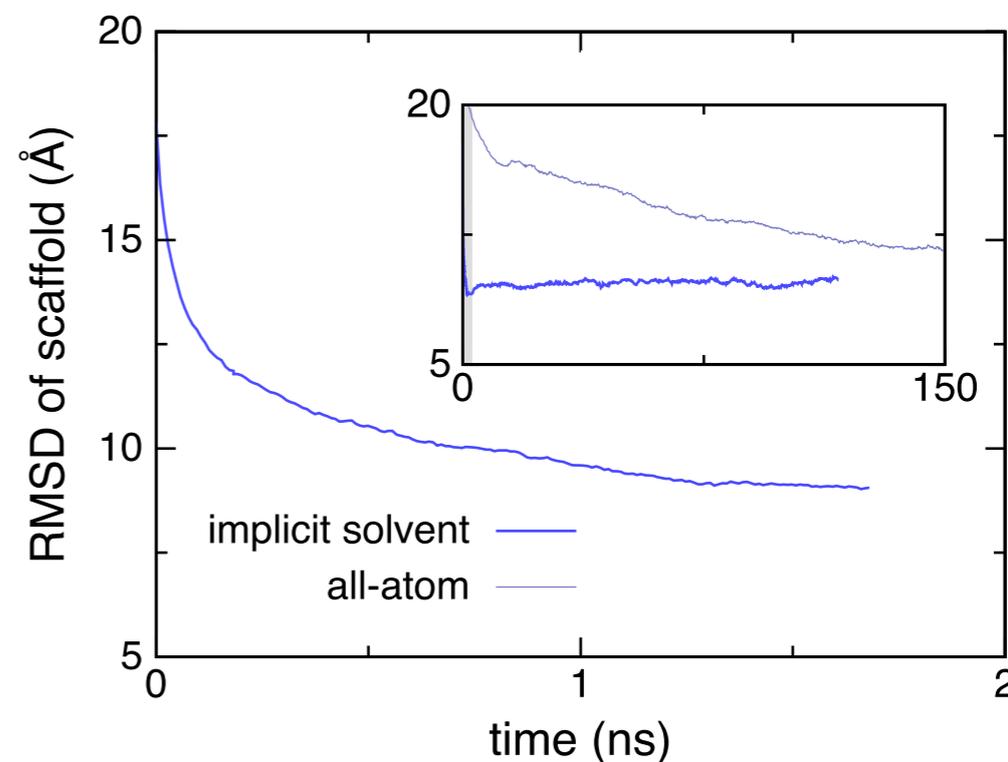
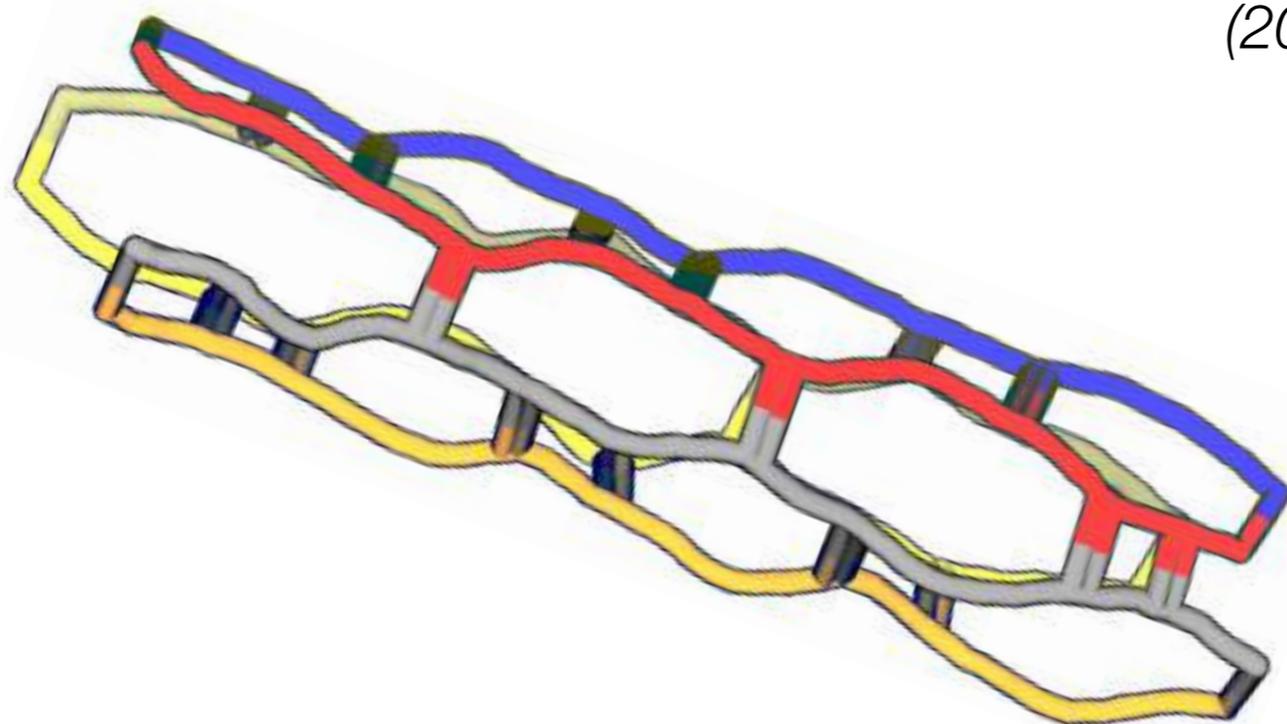
# Elastic network of restraints guided MD (ENRG MD) ~10,000 times more efficient



Solvent replaced  
with elastic  
network



Maffeo, Yoo & Aksimentiev, *NAR* 44: 3013  
(2016)



# ENRG MD For Origami Structure Prediction

Upload a DNA origami design .json file

No file chosen

Select the origami lattice. \*

Square

Honeycomb

Select the scaffold sequence. \*

m13mp18 (up to 7,249 bases)

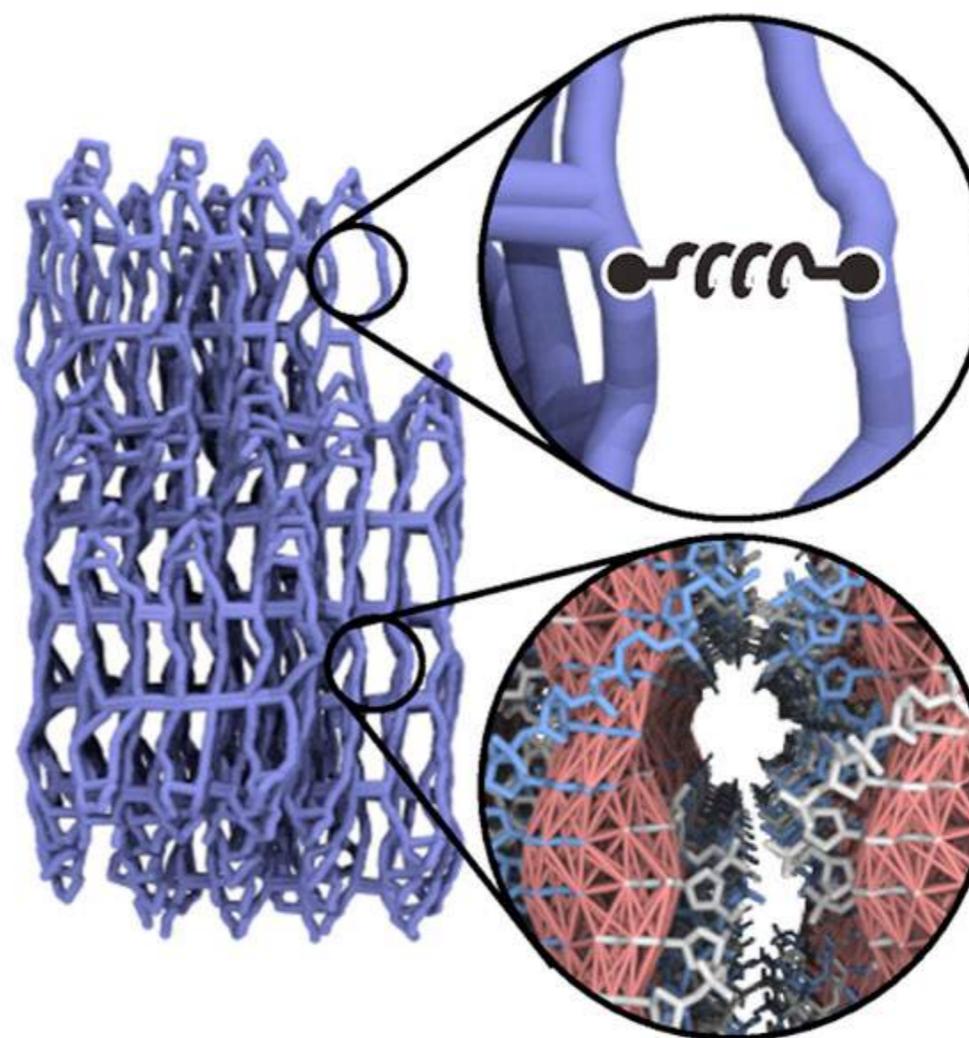
Custom

Simulation package \*

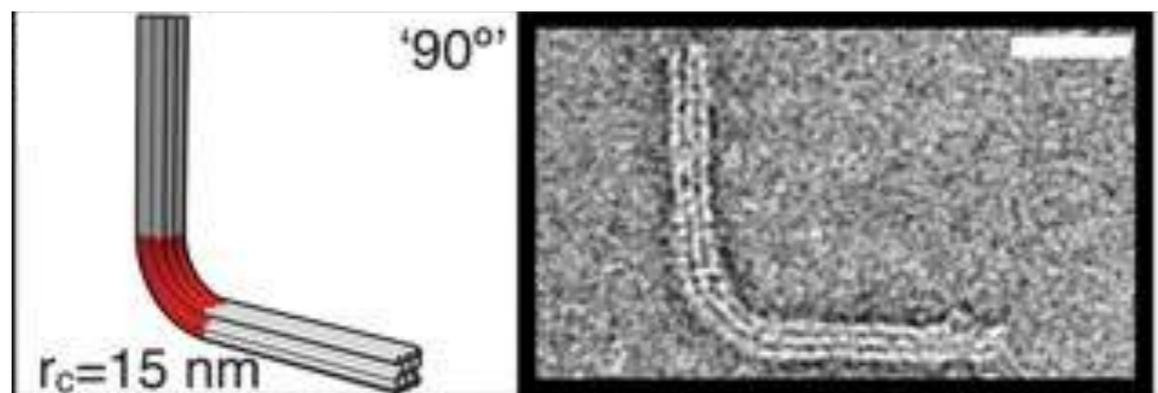
NAMD (CHARMM FF)

Gromacs (AMBER FF; beta coming soon)

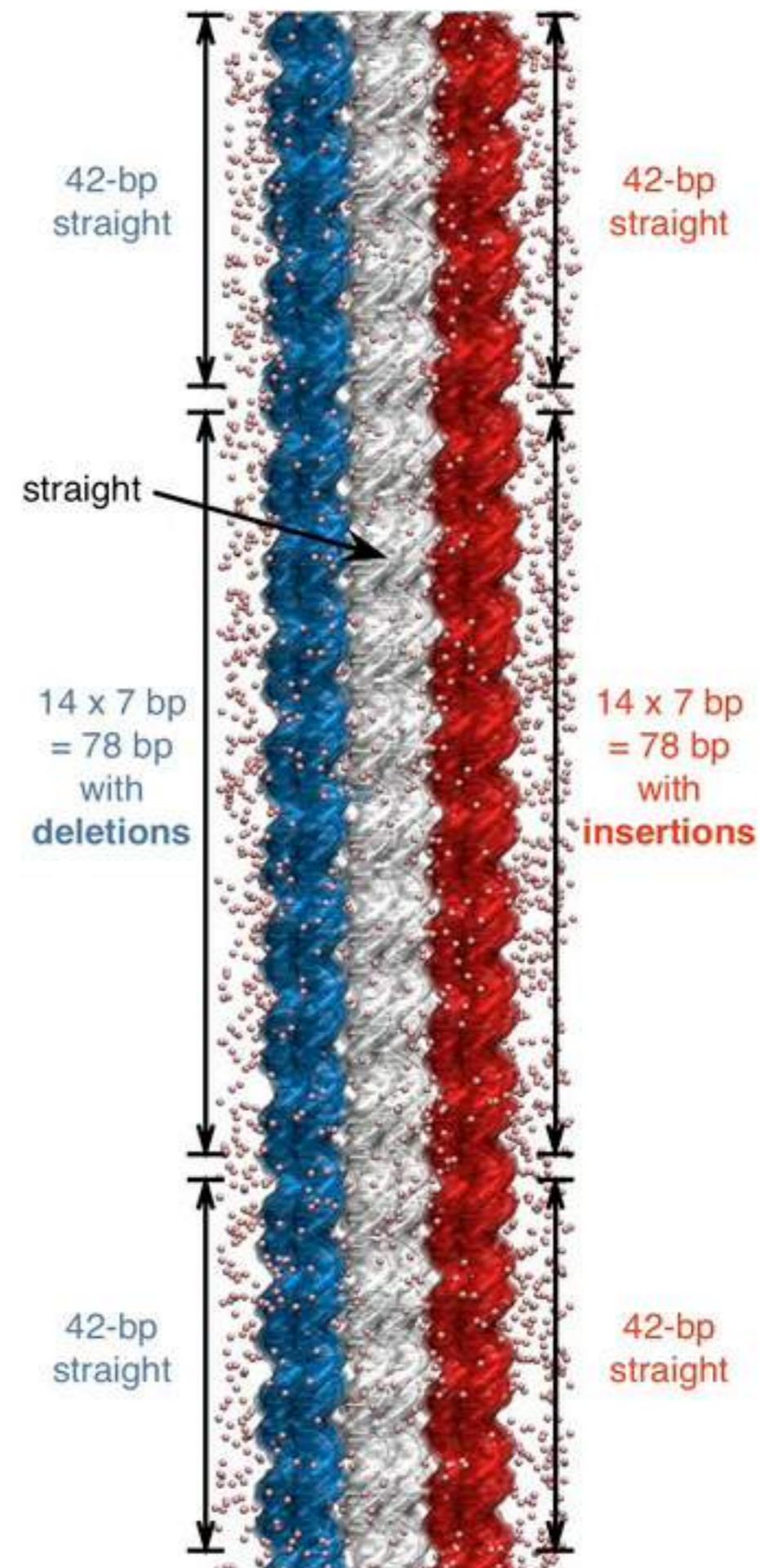
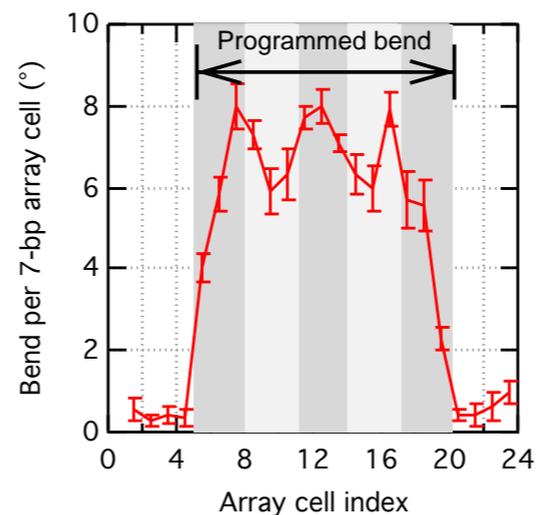
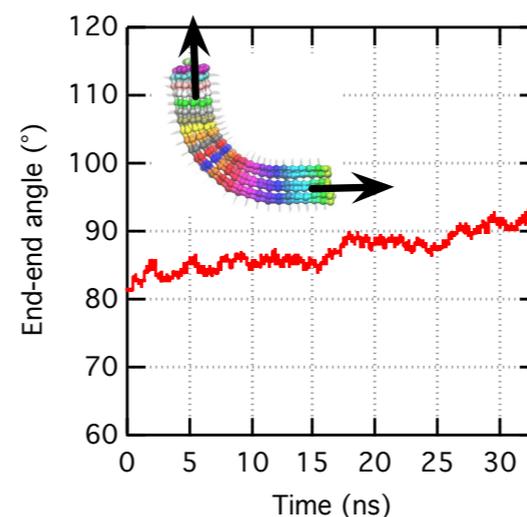
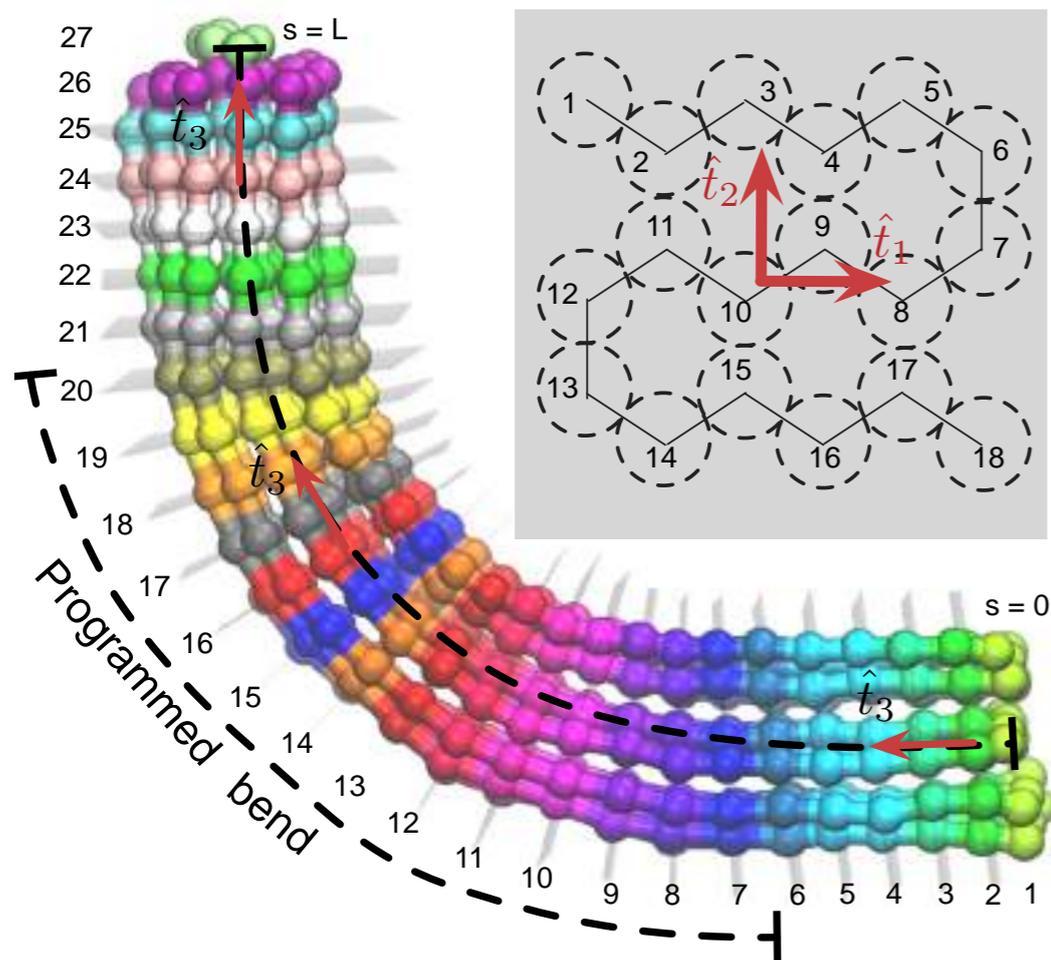
Create simulation files



# Curved DNA origami systems



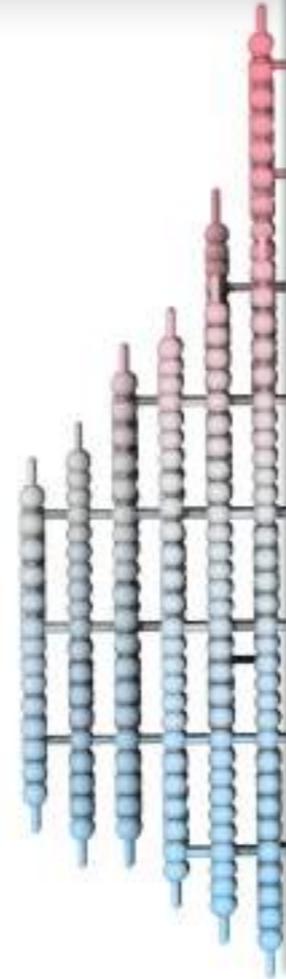
Experiment  
 Dietz, H. et al,  
*Science*, 325



Yoo and Aksimentiev, PNAS 110:20099 (2013)

# Multi-resolution simulations of self-assembled DNA nanostructures

```
9. dna-nano  
~/dna-nano mrdna pointer_v1_1
```



**Setting up a simulation to capture a DNA origami object in a nanopipette**

Here we will recreate a portion of a recent [study](#) of a voltage-sensitive DNA origami object that is captured at the opening of a glass nanopipette.

**Load the object into cadnano**

First, let's have a look at the DNA origami object. When you load the object into cadnano, you can see that it is a two-layer plate with a hole in the middle. If you inspect the design carefully you might notice that there is an insertion of nearly 800 bp near the hole. We call this long stretch of DNA the "leash". This long flexible piece of DNA is expected to result in the capture of the plate object by the nanopipette.

Notice the large insertion on helix 36.

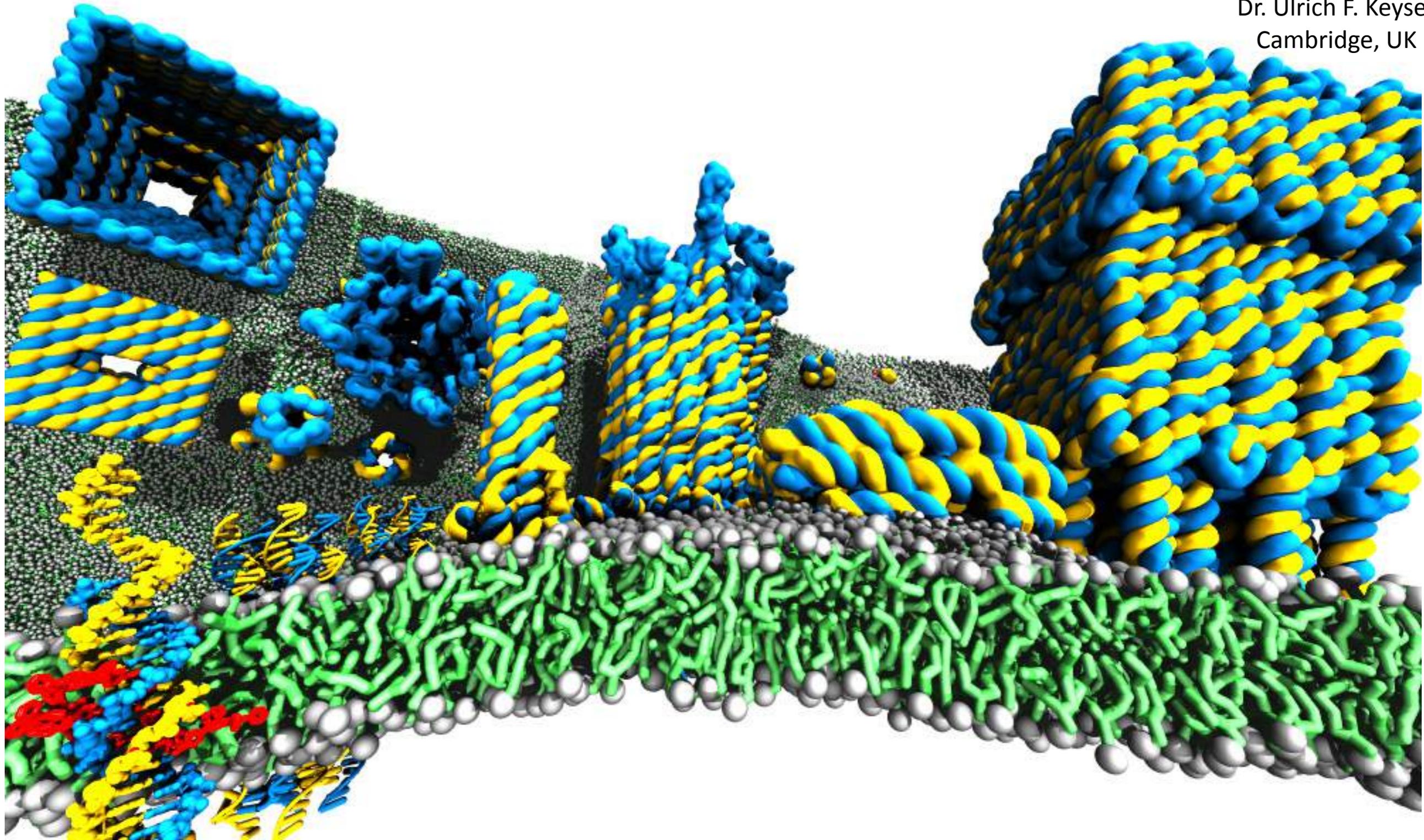
**Examine the potential derived from a continuum COMSOL model**

Our simulation engine can apply forces to an object using a potential described in a regular three-dimensional grid. The engine reads OpenDX grid files for this purpose. Such files can be generated using VMD or the `gridDataFormat` Python module. It is possible to convert the output of some continuum models to the DX format, but this is beyond the scope of this tutorial.

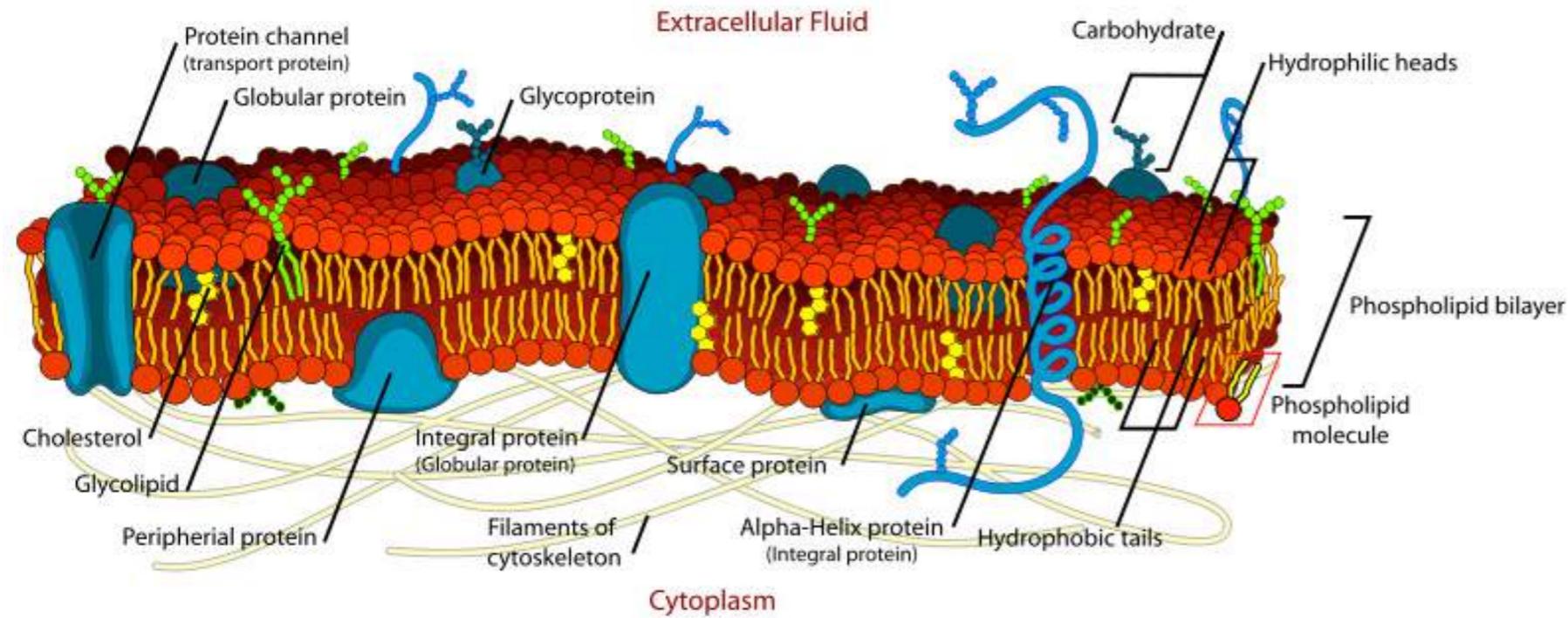
# DNA membrane channels



Dr. Ulrich F. Keyser  
Cambridge, UK

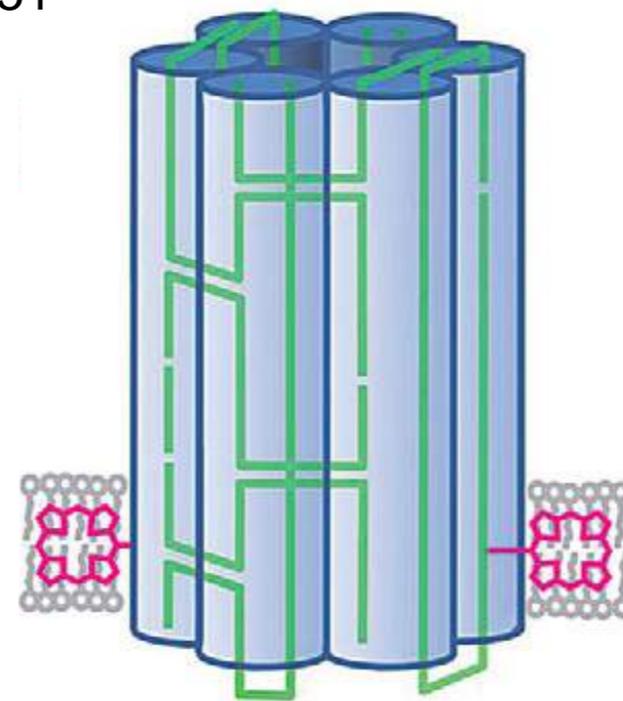
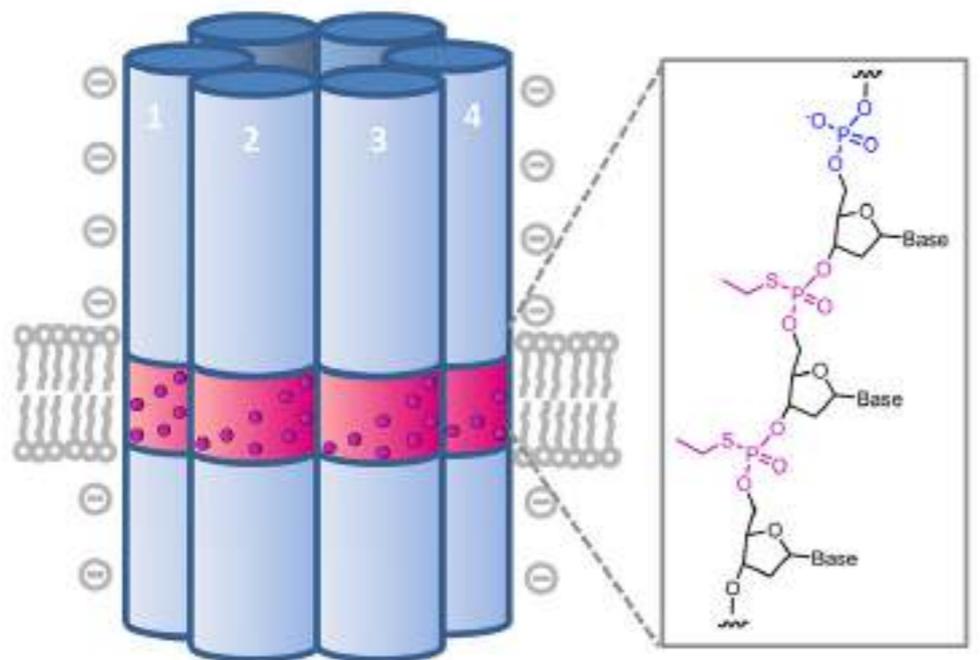


# DNA Ion Channels

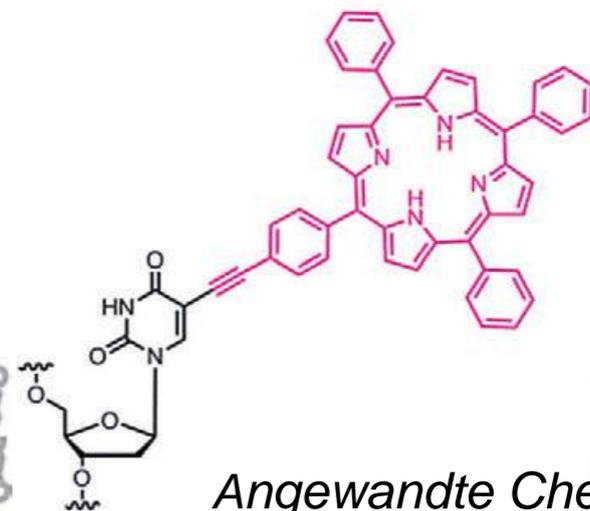


Langecker, M. *et al.*, *Science*: 338, 932-936.

*Nano Letters*, 13: 2351

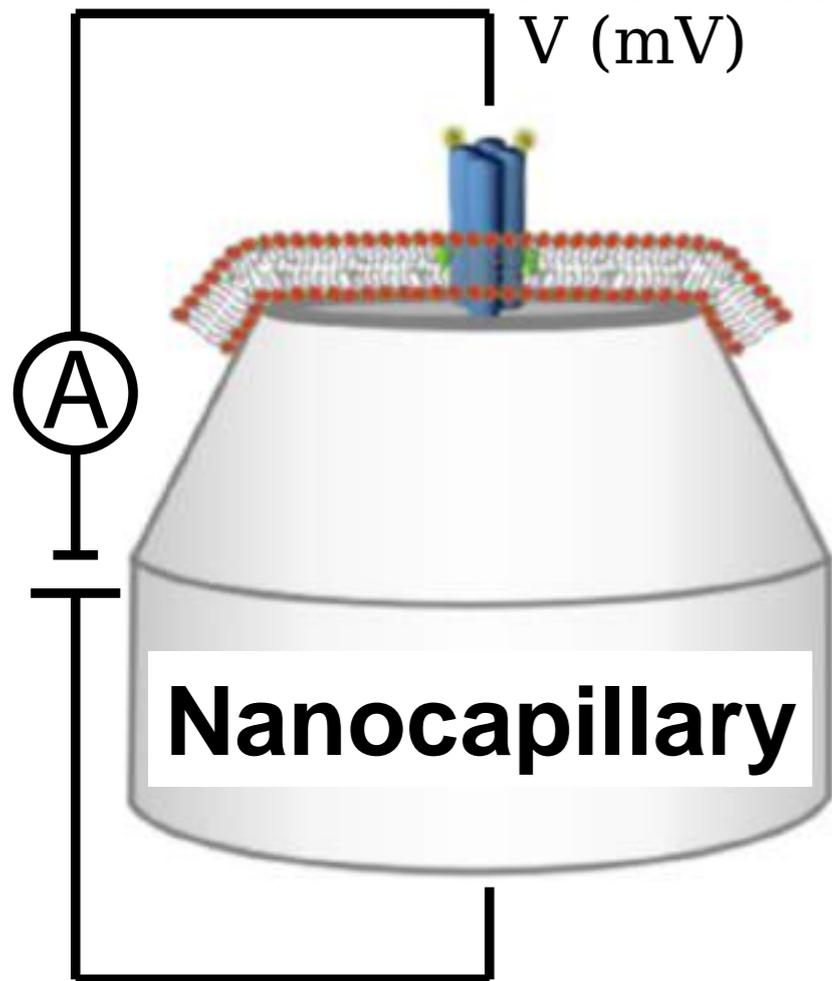
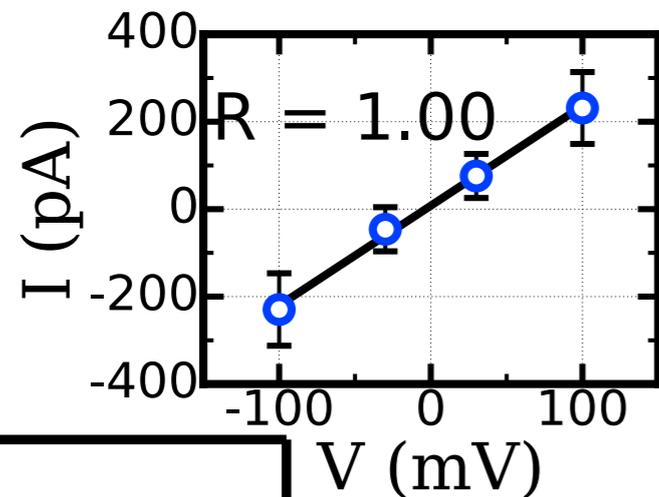


**~1 nS conductance**

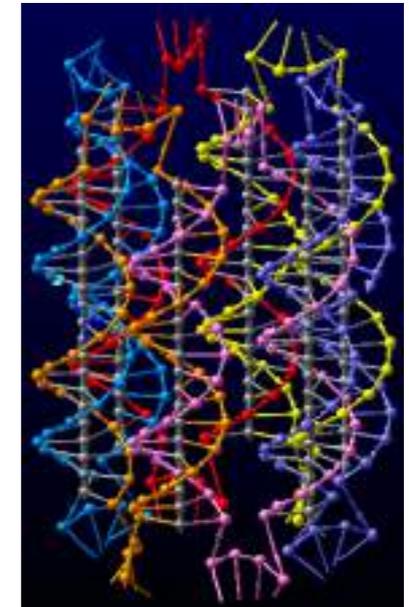
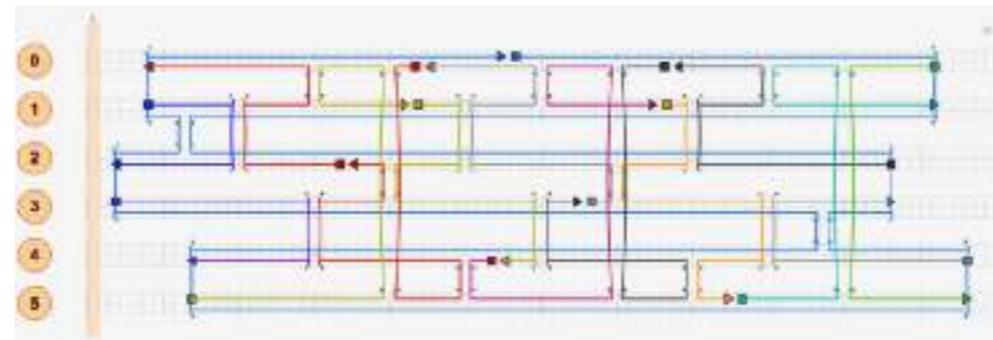


*Angewandte Chemie*  
10.1002/anie.201305765

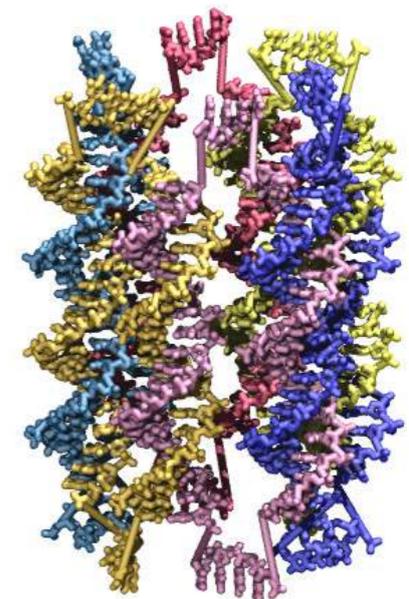
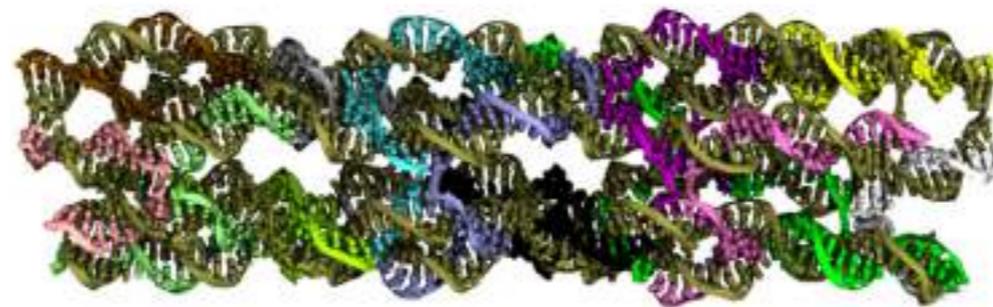
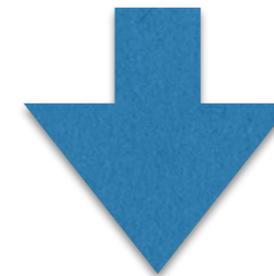
# MD simulation of DNA channel conductivity



## caDNAno or NanoEngineer-1



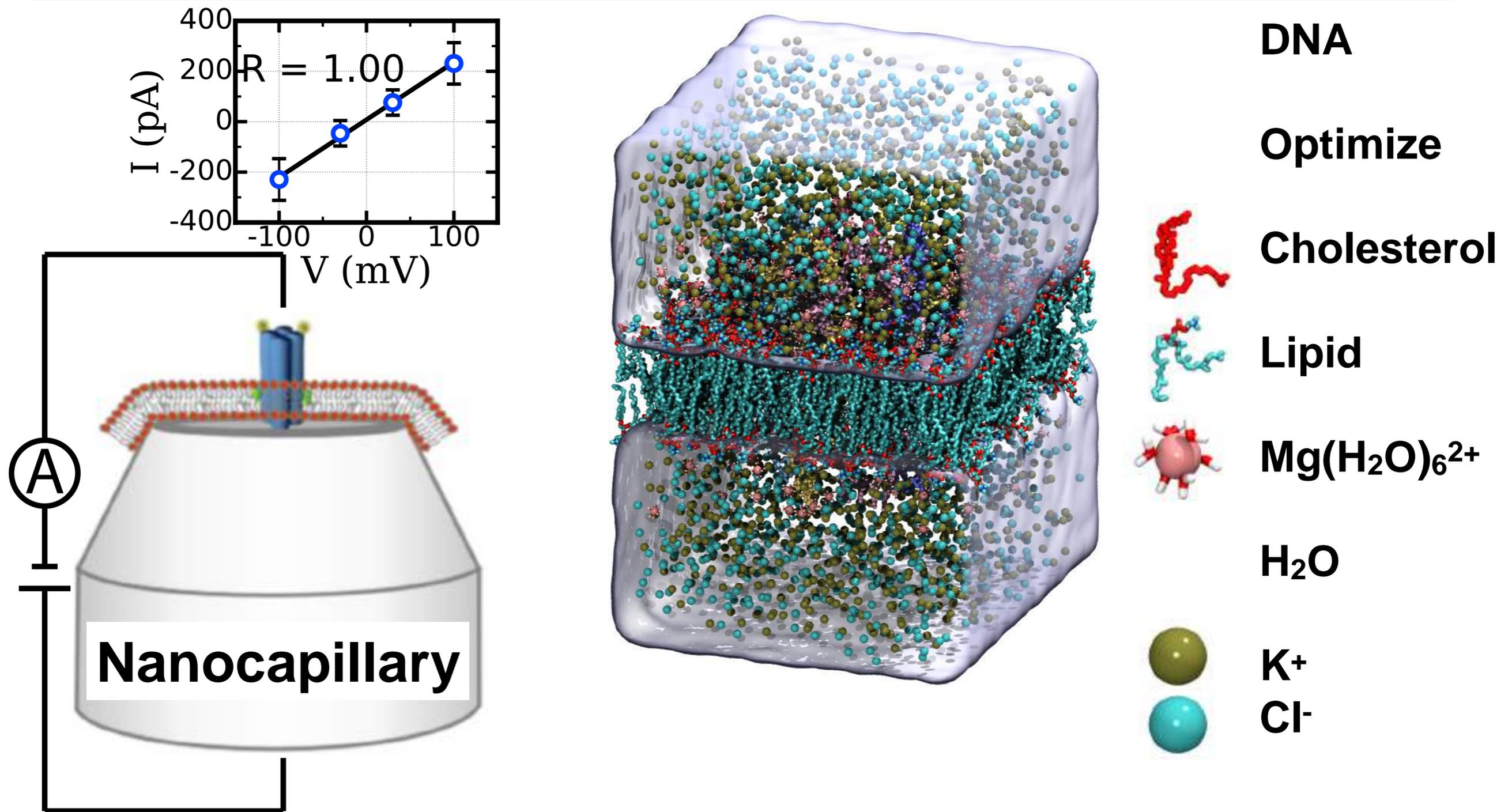
all-atom  
coordinates



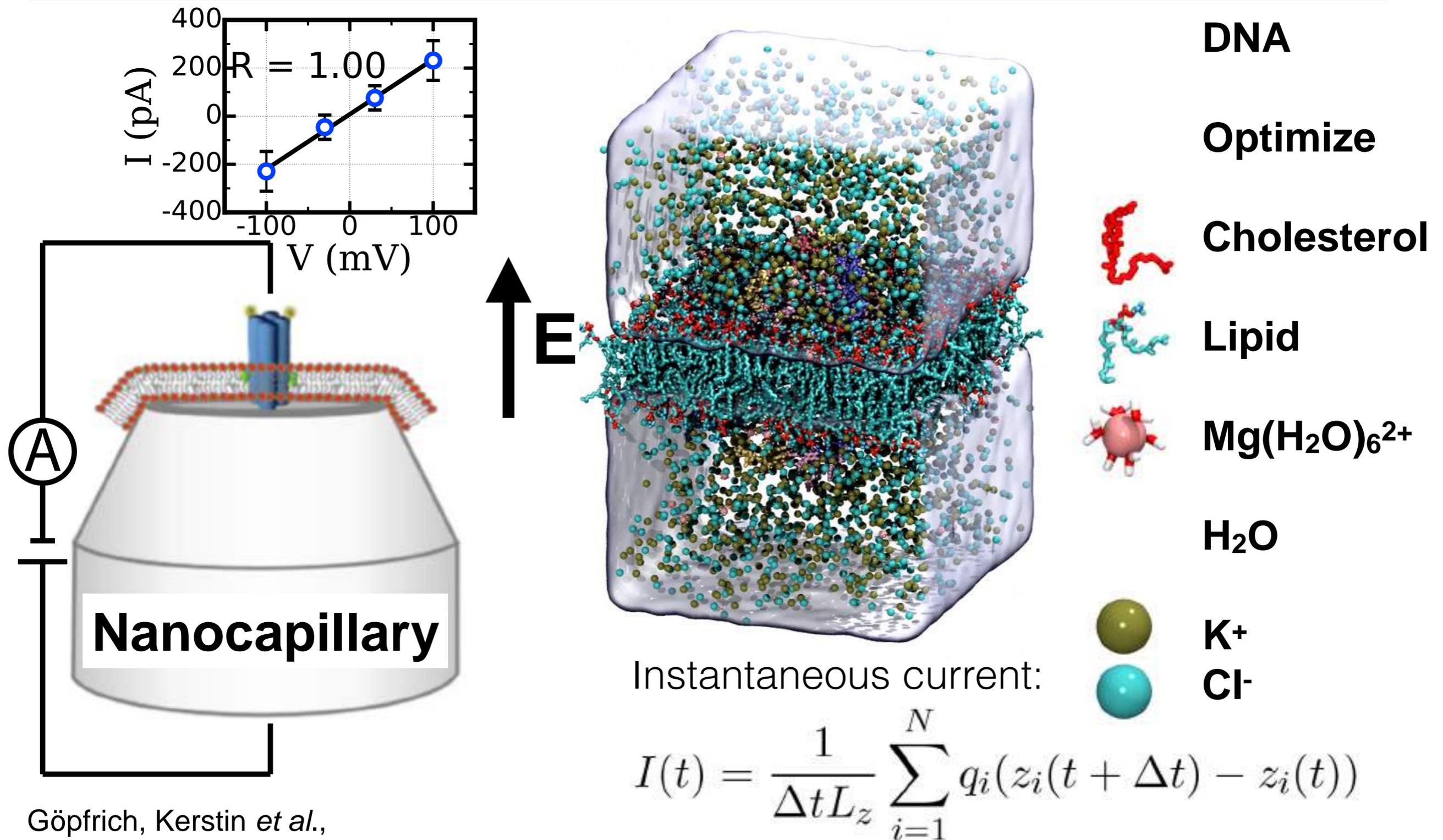
Göpfrich, Kerstin *et al.*,  
*Nano Lett.*, 2015, 15(5), 3134–3138.

(Right) design from Burns, Jonathan R. *et al.*,  
*Nat. Nanotechnol.*, 2016, 11, 152–156.

# MD simulation of DNA channel conductivity

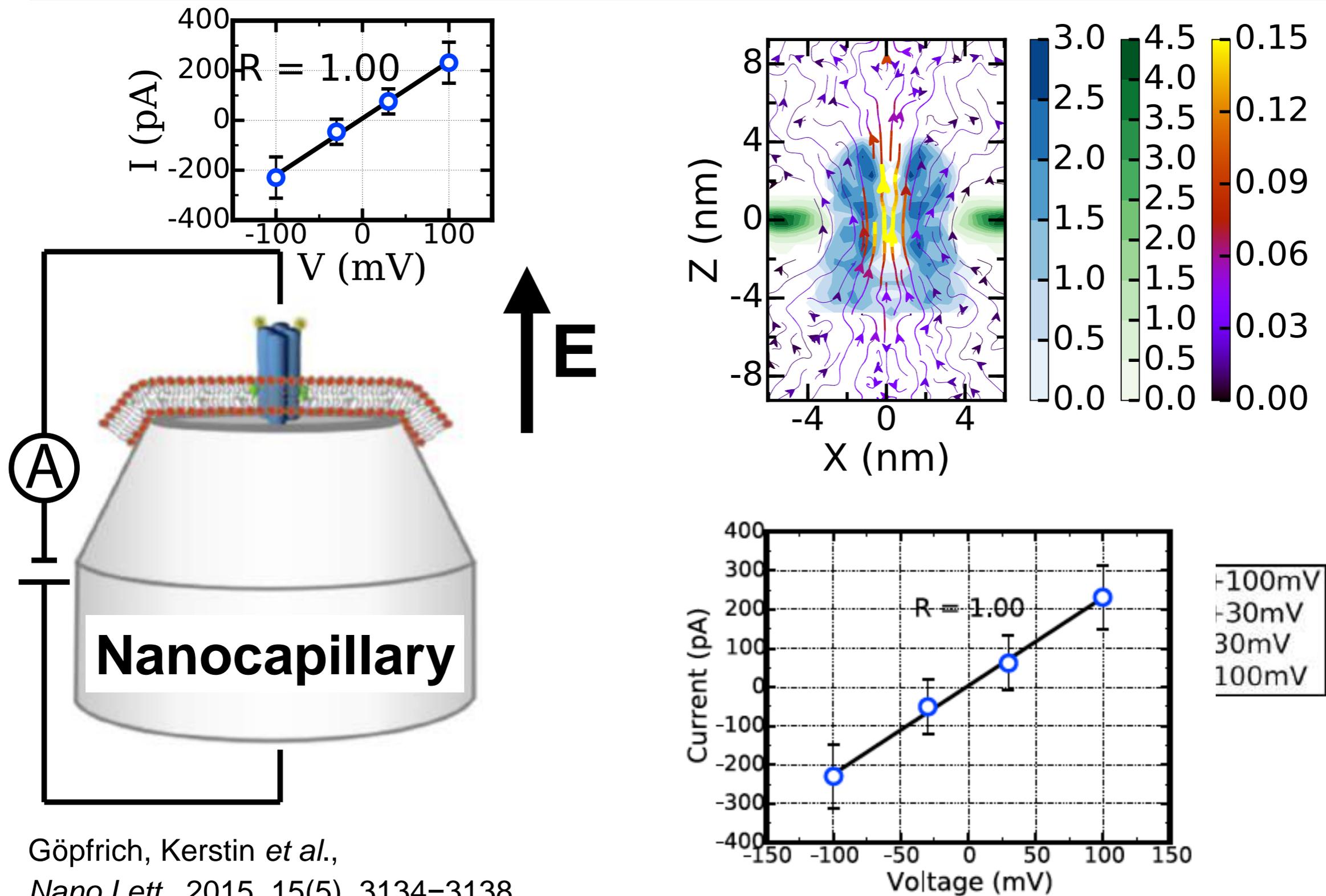


# MD simulation of DNA channel conductivity



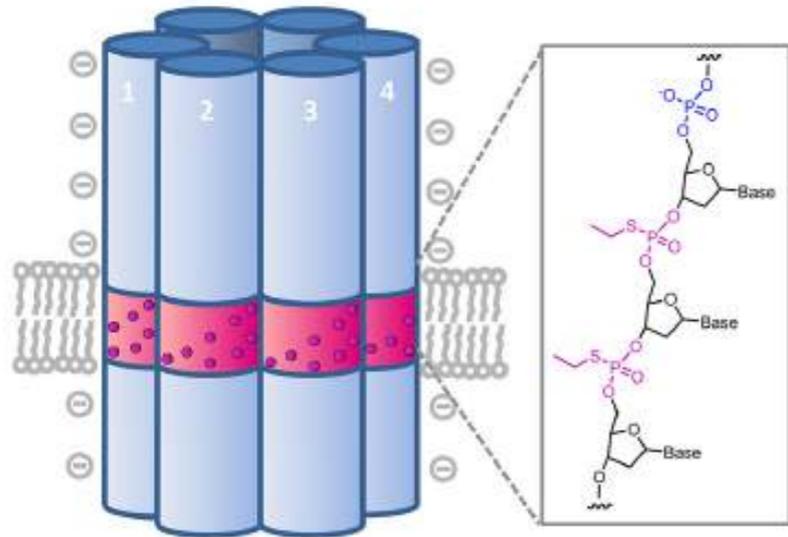
Göpfrich, Kerstin *et al.*,  
*Nano Lett.*, 2015, 15(5), 3134–3138.

# MD simulation of DNA channel conductivity



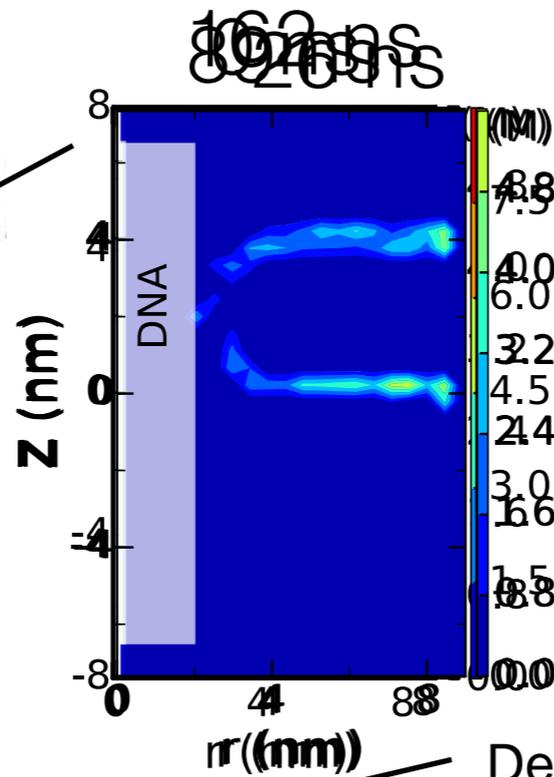
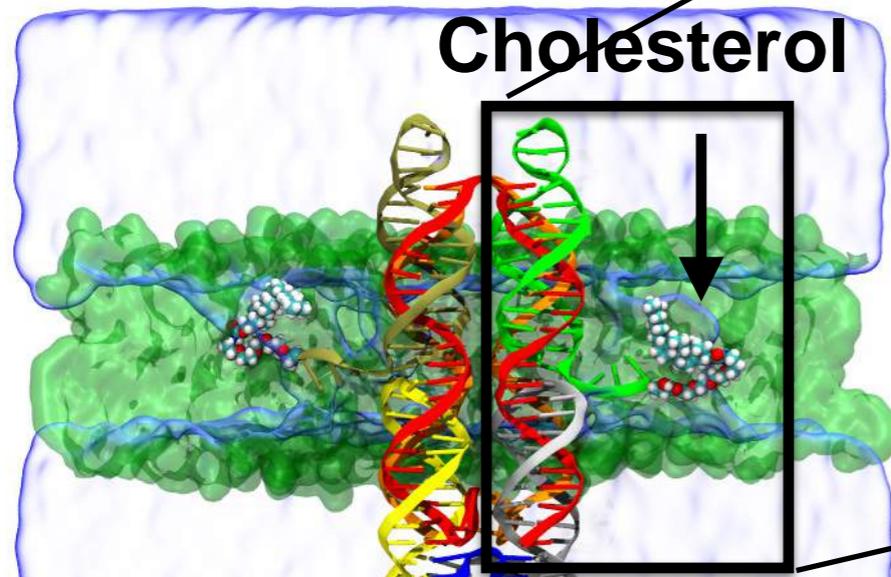
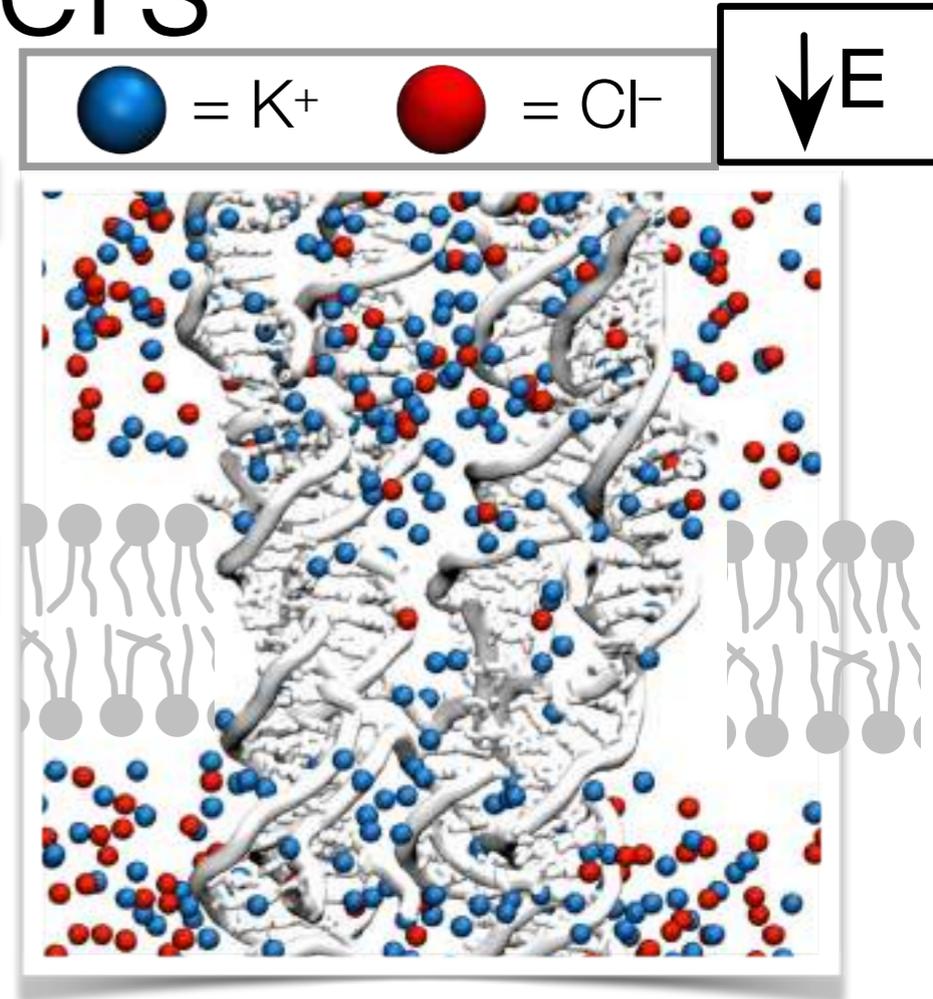
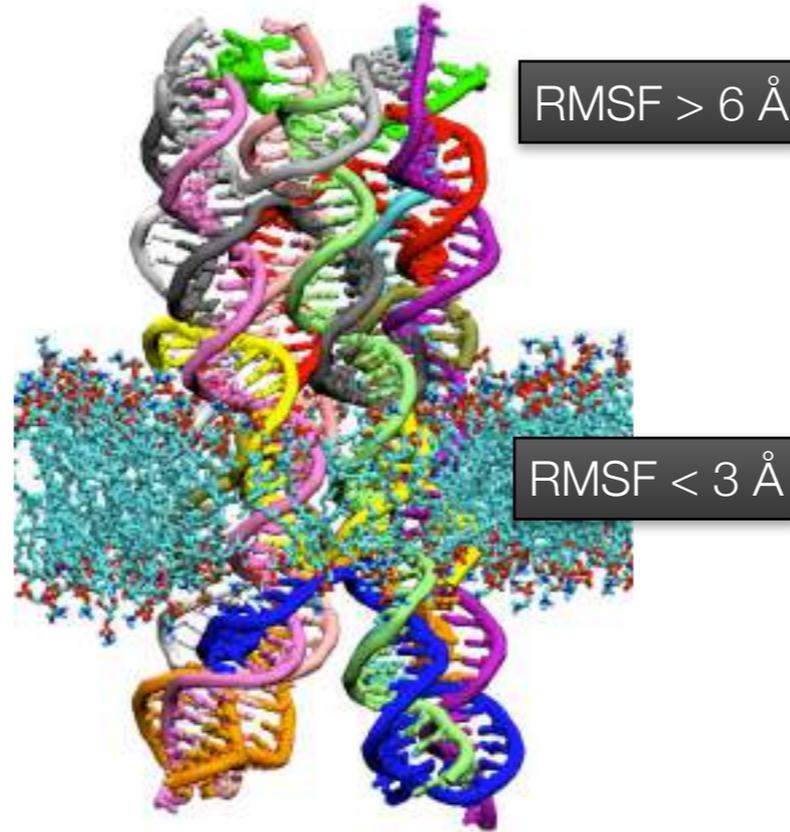
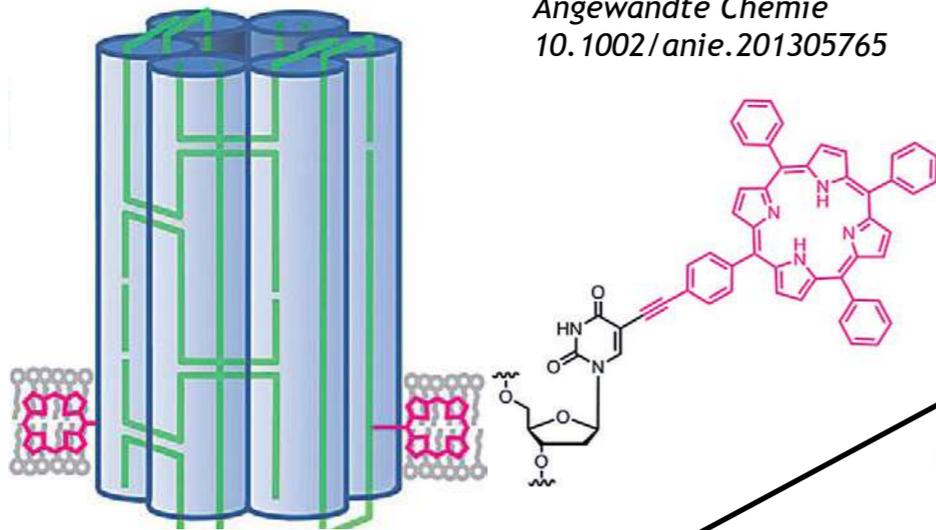
Göpfrich, Kerstin *et al.*,  
*Nano Lett.*, 2015, 15(5), 3134–3138.

# Chemistry matters

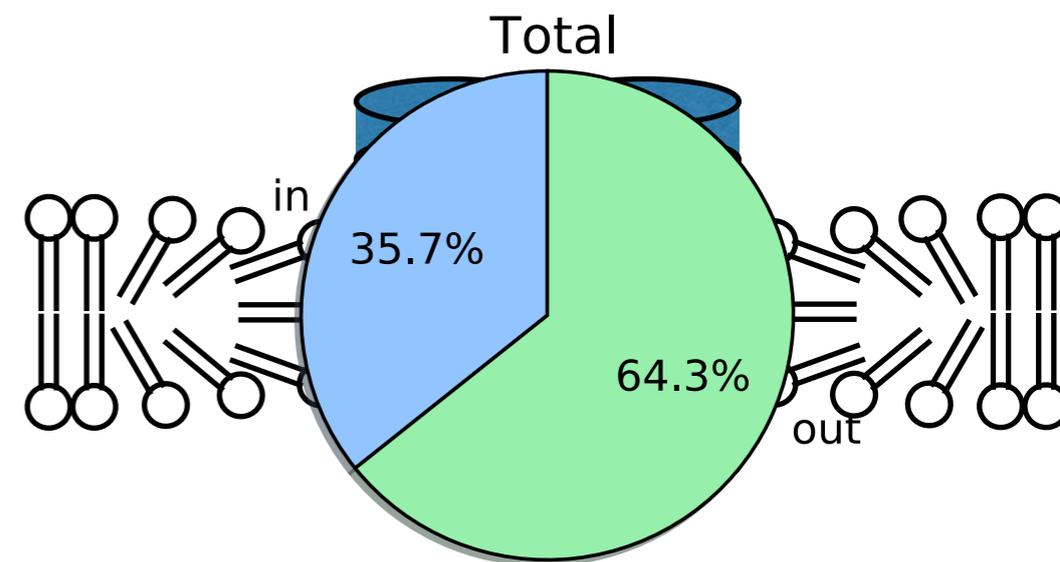


*Nano Letters*, 13: 2351

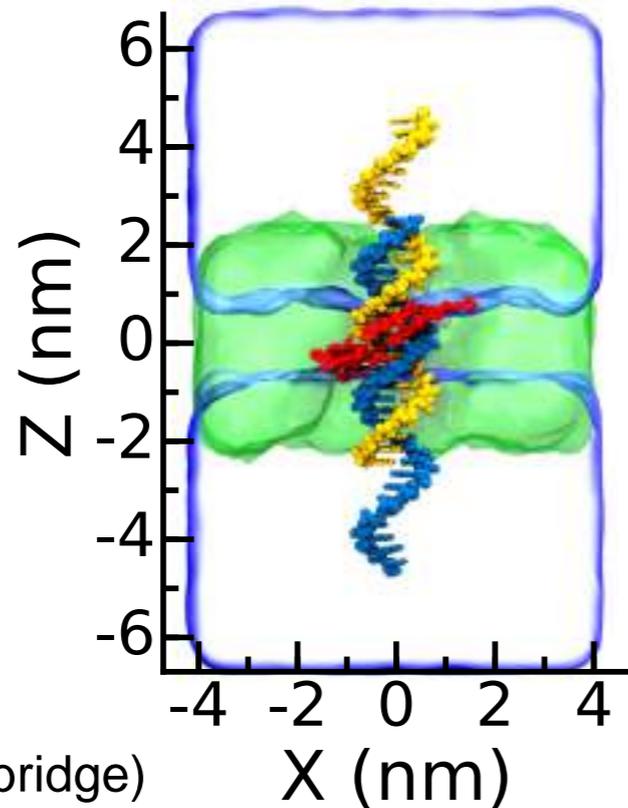
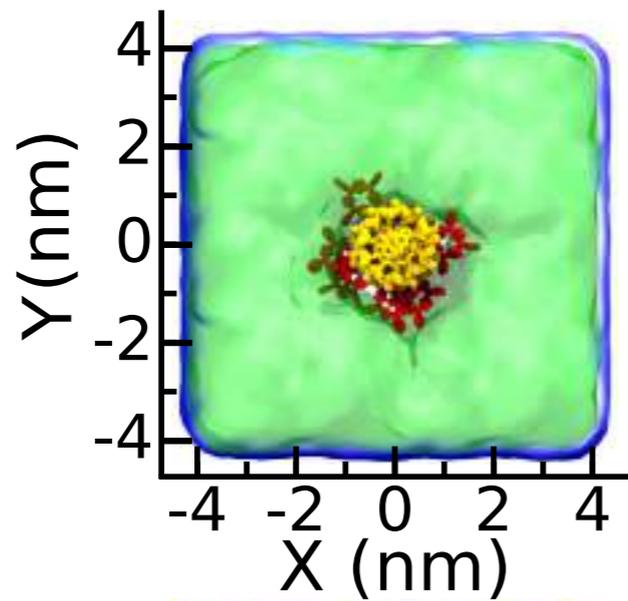
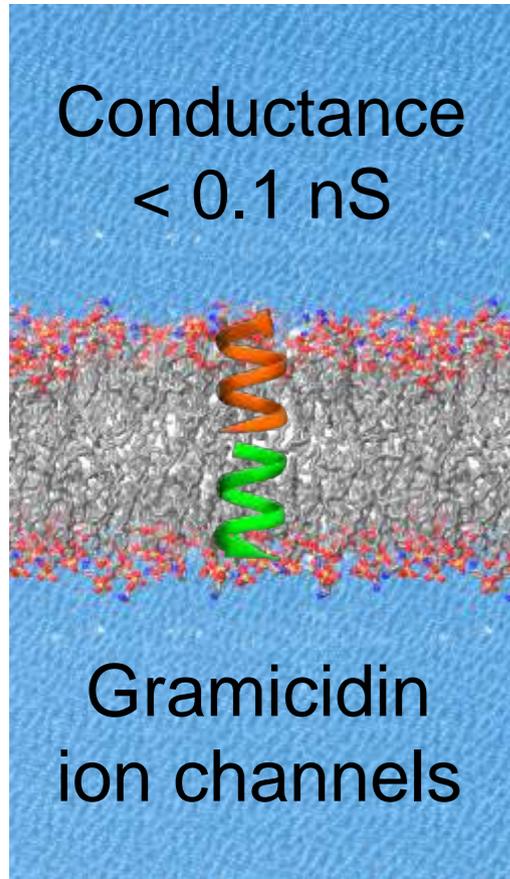
*Angewandte Chemie*  
10.1002/anie.201305765



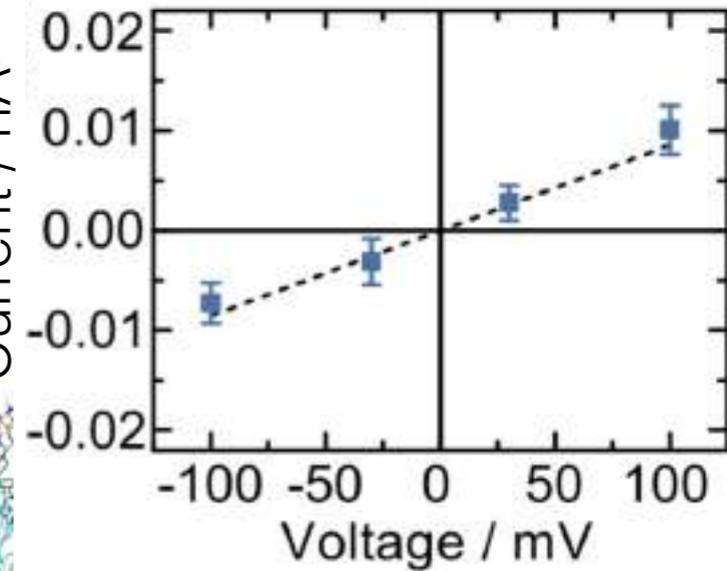
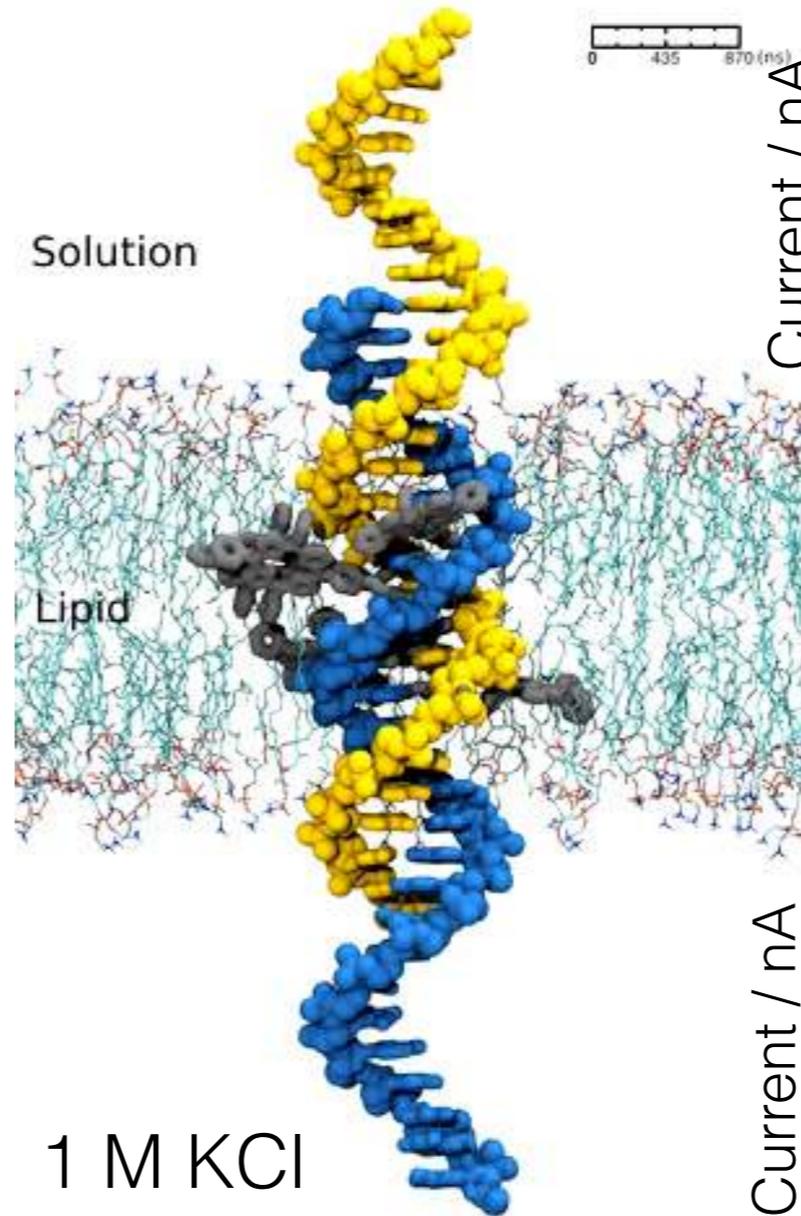
Density of lipid head groups



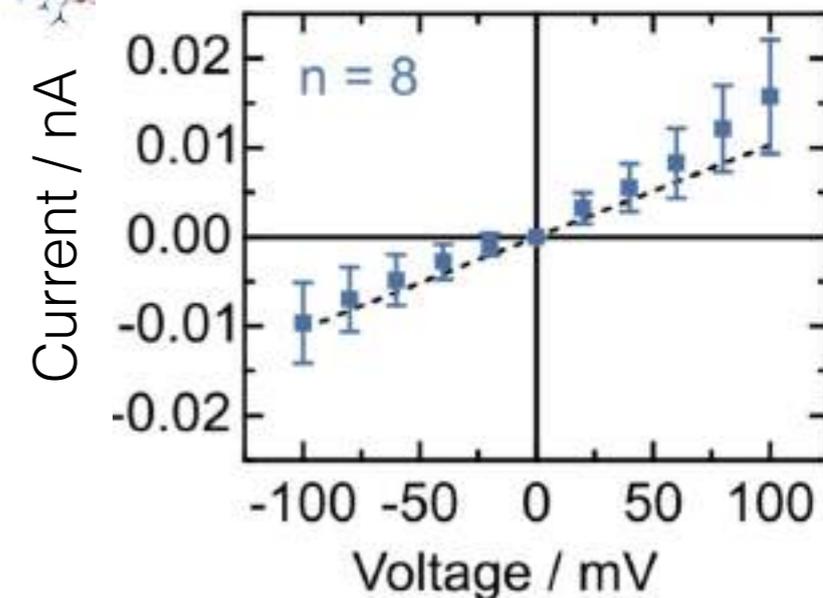
# Small conductance DNA channel



1 helix  
140,000 atoms



Conductance:  $\sim 0.1 \text{ nS}$



Experiment:

Ulrich Keyser (Cambridge)

Eugen Stulz (Southampton)

Mathias Winterhalter (Jacobs U)

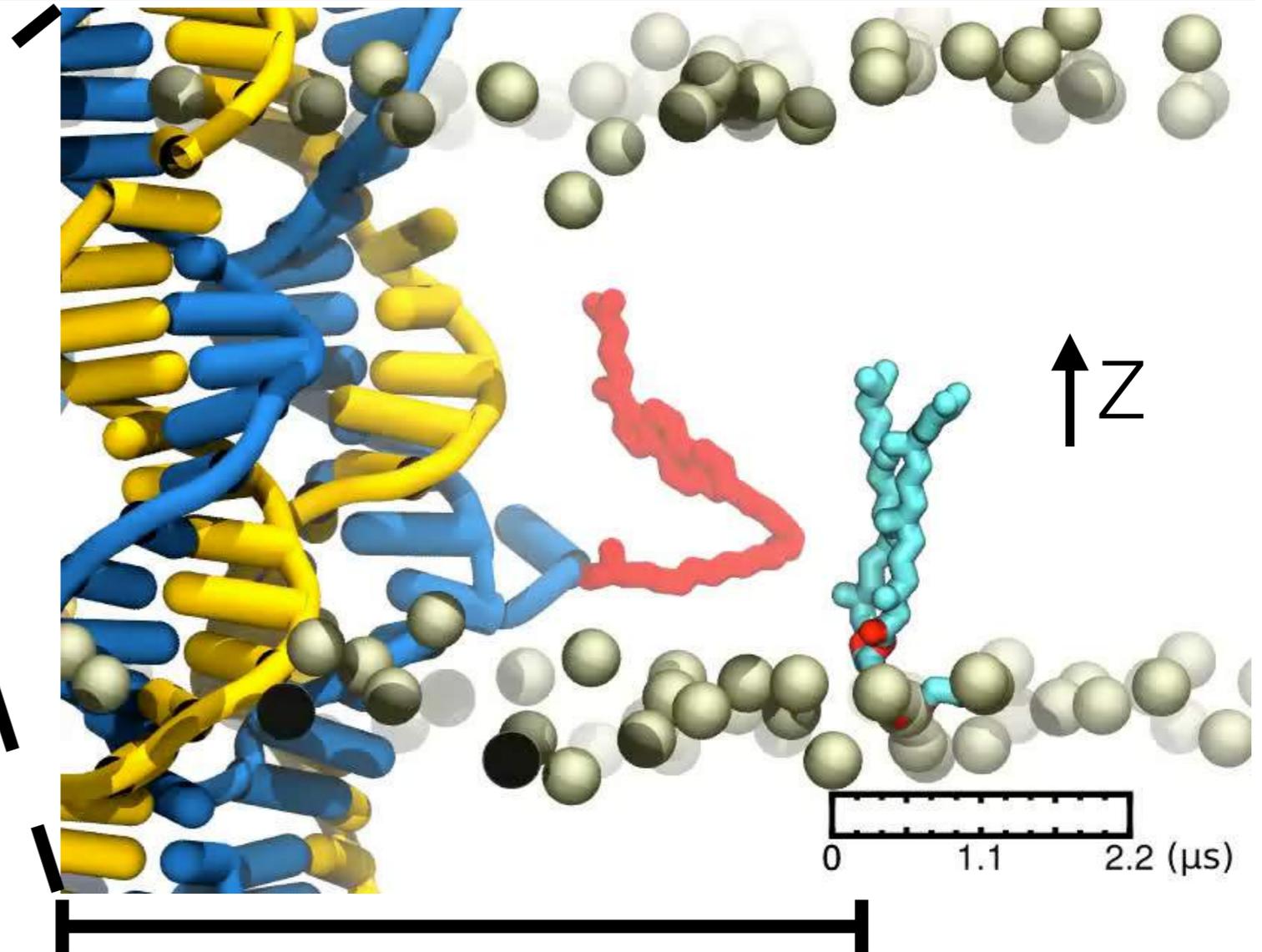
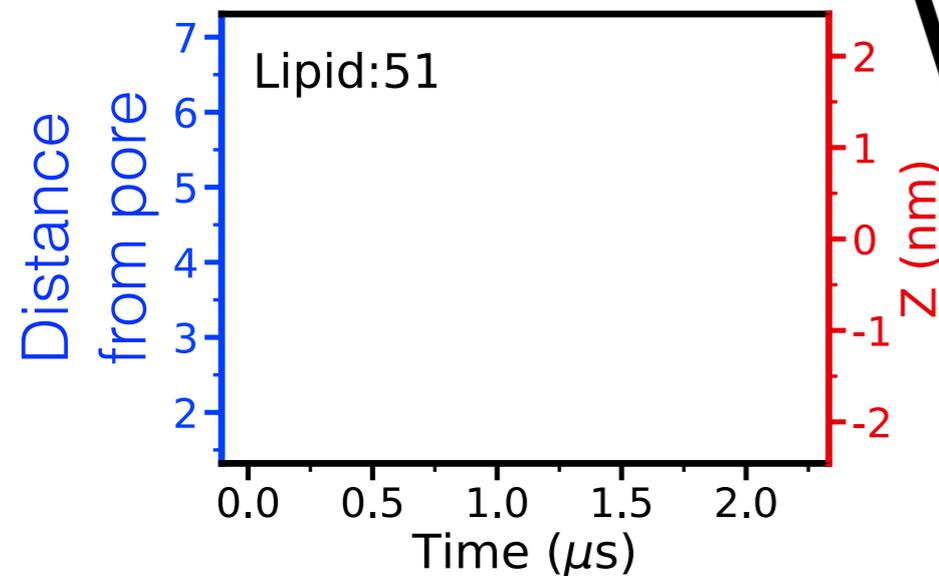
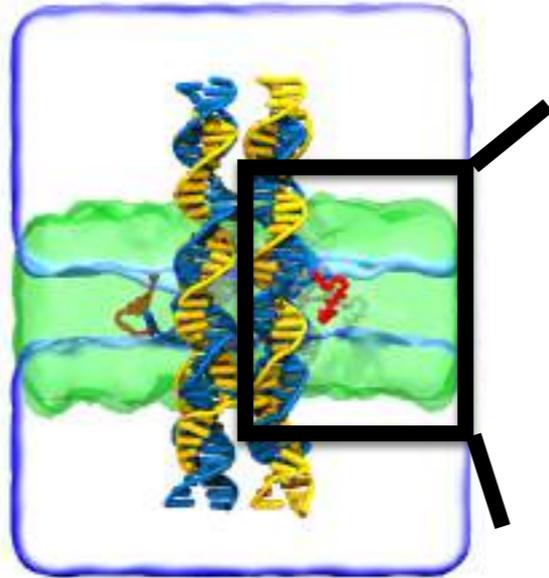
Goepfrich, et al., Nano Lett 16: 4665 (2016)



Chen Yu Li

# All-atom MD simulation of lipid-DNA interface

No applied electric field

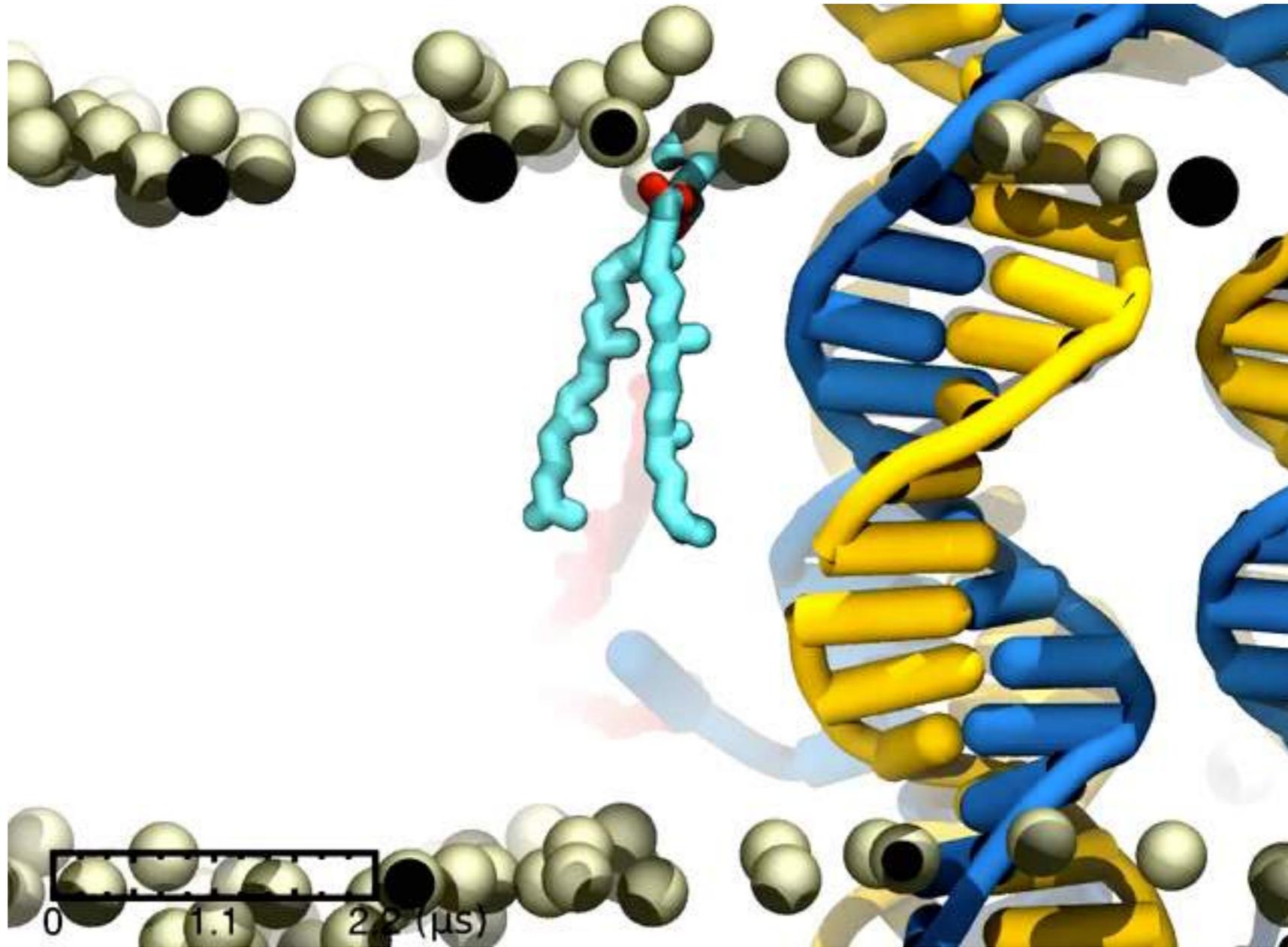


Distance from pore

**Lipid molecules can translocate to the other leaflet through the toroidal pore made by DNA**

Nature Communications 9:2426 (2018)

Lipid translocation through toroidal pores is very common and very fast



# Lipid translocation in cells is catalyzed by enzyme

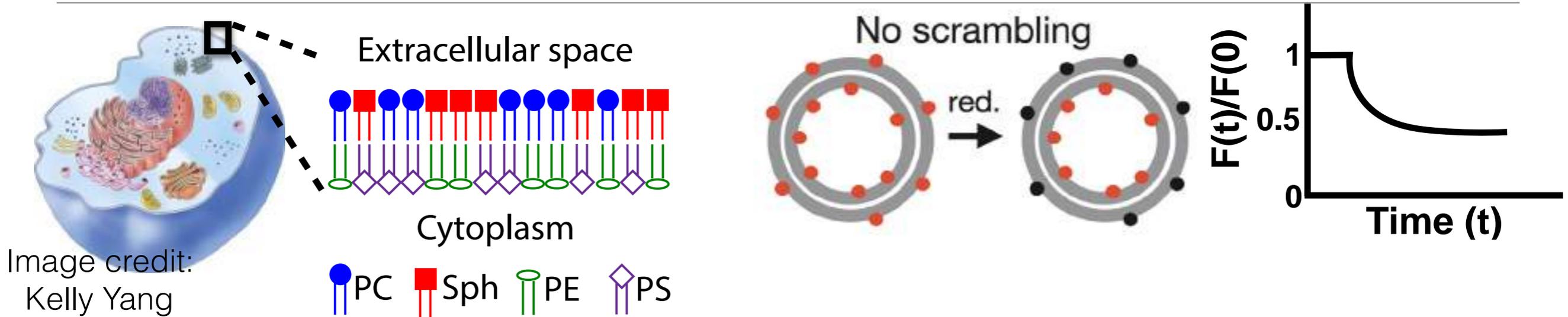
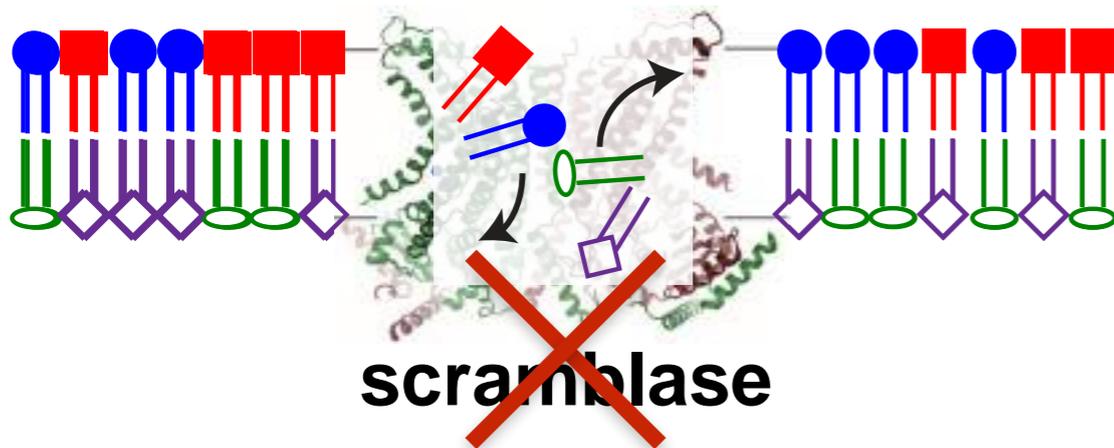


Image credit:  
Kelly Yang

**Lipid molecules are asymmetrically distributed in the cell membrane.**

**apoptosis or thrombin formation:**



**Deficiency in lipid scrambling could result in autoimmune response or Scott syndrome.**

$$\frac{F(t)}{F(0)} = x_{in}e^{-k_s t} + (1-x_{in})e^{-k_q t}$$

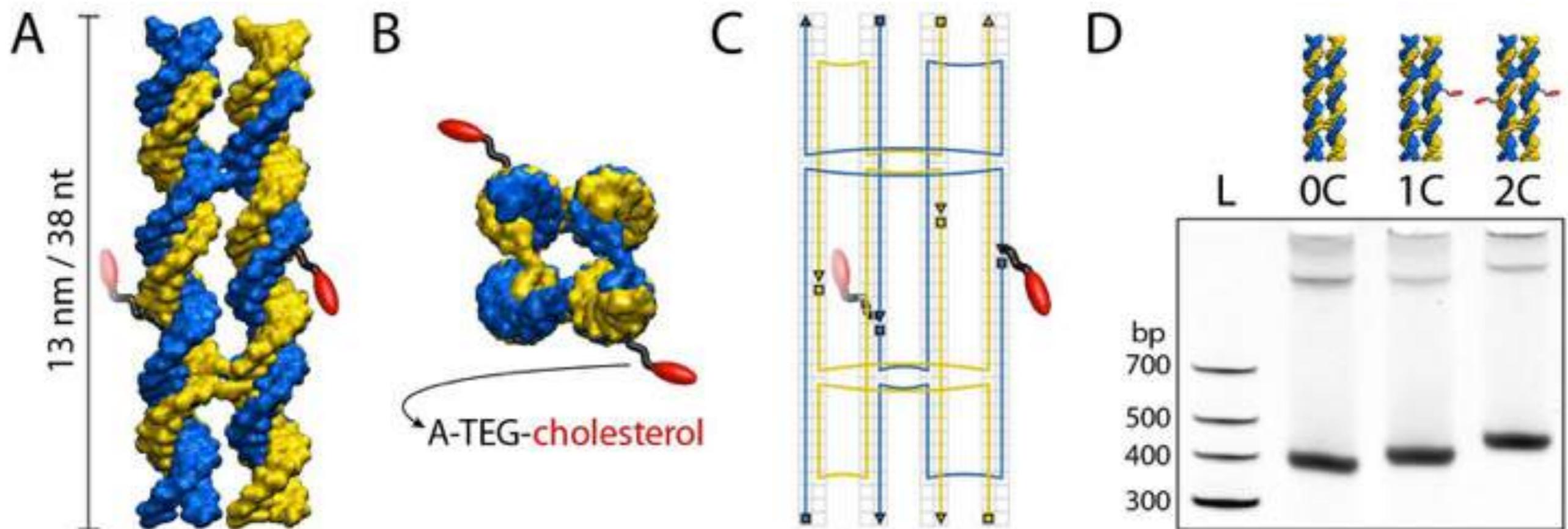
$x_{in}$ : ratio of labeled lipid in the inner leaflet

$k_q$  and  $k_s$ : rate constant for quenching and scrambling

# Experimental verification



Alex Ohmann

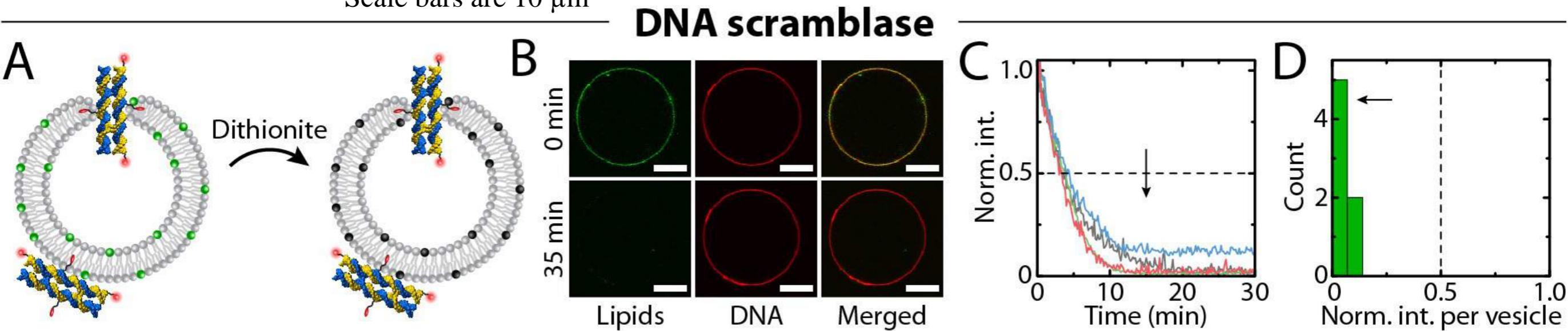


# Experimental verification



Alex Ohmann

Scale bars are 10  $\mu\text{m}$

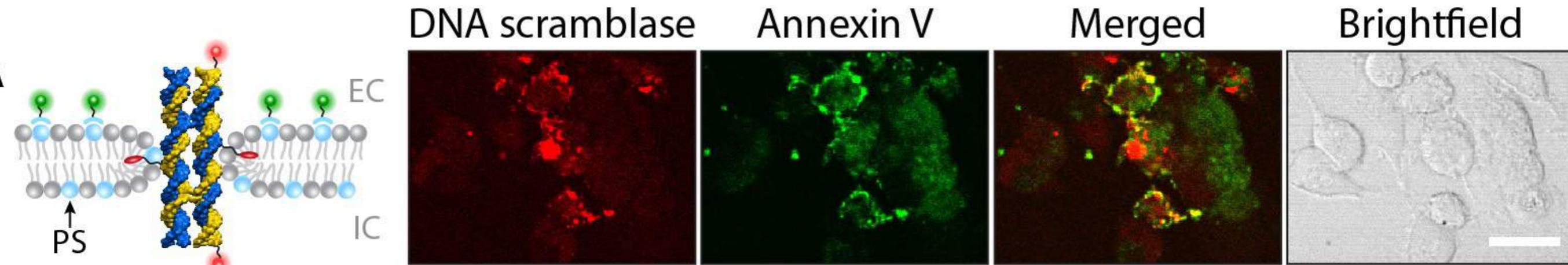


# Works in human cells

Human cells contain PS lipids at the inner membrane

Annexin V binds specifically to PS lipids

Scale bar is 20  $\mu\text{m}$



Breast cancer cells from the cell line MDA-MB-231

**Positive control:** apoptosis-inducing microbial alkaloid staurosporine

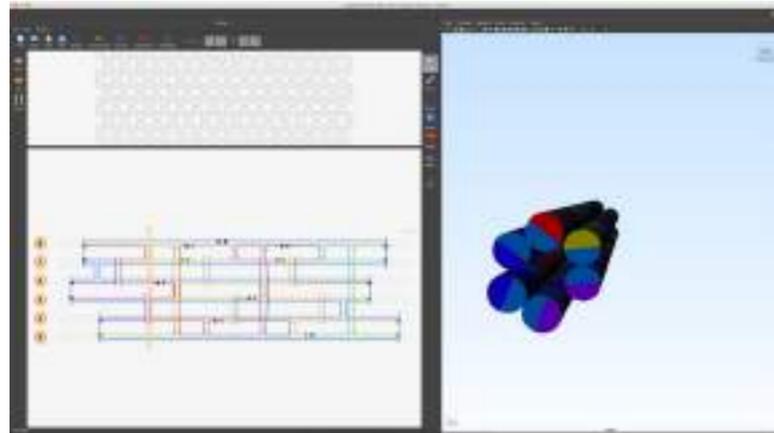
**Negative control:** DNA folding buffer

# Tutorials overview

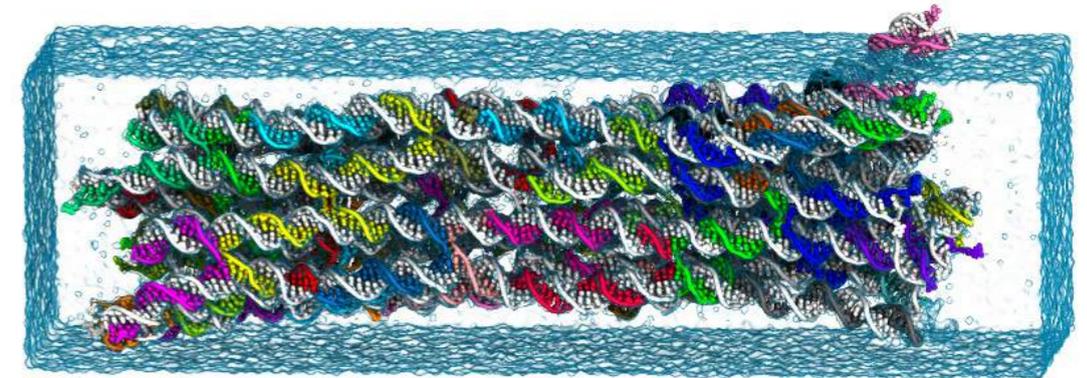
## Design

## Simulations

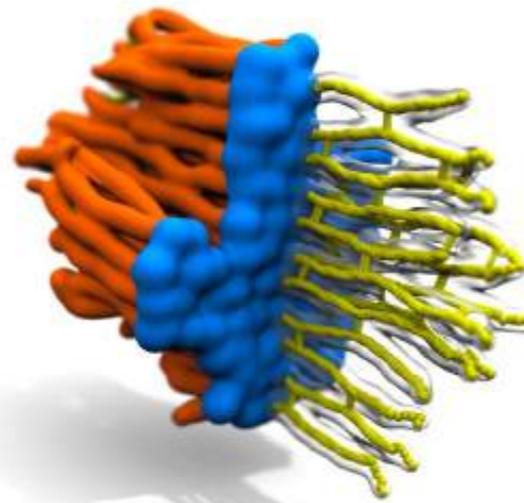
**Cadnano (today)**



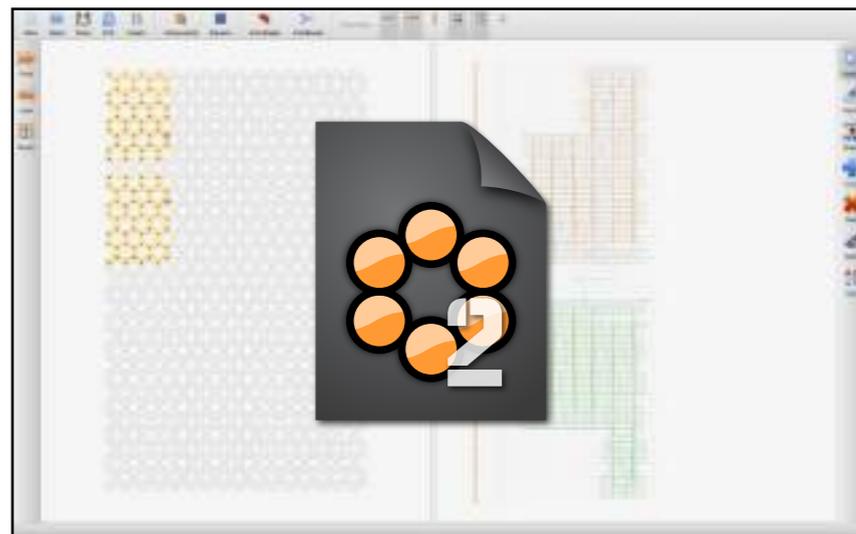
**All-atom with NAMD (today)**



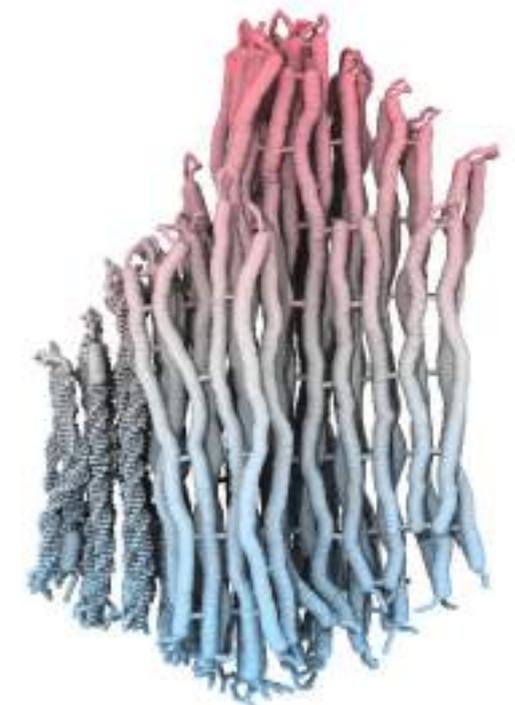
**Images  
and movies  
with VMD**



**Cadnano toolkit (tomorrow)**



**Coarse-grained  
with ARBD  
(tomorrow)**



# Acknowledgements



Jejoong Yoo



Chen-Yu Li



TeraGrid™

