Molecular Dynamics to Sample Energy Landscape

Initial coordinates have bad contacts, causing high energies and forces (due to averaging in observation, crystal packing, or due to difference between theoretical and actual forces)

Minimization finds a nearby local minimum.

kТ

Heating and cooling or equilibration at fixed temperature permits biopolymer to escape local minima with

Energy

Conformation

Initial dynamics samples thermally accessible states.

Molecular Dynamics to Sample Energy Landscape



Patience is required to observe Molecular Events



Molecular Dynamics Ensembles

Constant energy, constant volume (NVE)

Constant temperature, constant volume (NVT)

Constant temperature, constant pressure (NPT)

Choose the ensemble that best fits your system and start the simulations

Equilibrium Properties of Proteins



through simulation (red = more flexible)

aligned together to see flexibility

Thermal Motion of Ubiquitin from MD

RMSD values per residue





RMS deviations for the KcsA protein and its selectivity filer indicate that the protein is stable during the simulation with the selectivity filter the most stable part of the system.



Temperature factors for individual residues in the four monomers of the KcsA channel protein indicate that the most flexible parts of the protein are the N and C terminal ends, residues 52-60 and residues 84-90. Residues 74-80 in the selectivity filter have low temperature factors and are very stable during the simulation.

Improving the speed cutoffs, PME, rigid bonds, multiple timesteps

- Nonbonded interactions require order N² computer time!
 - Truncating at $R_{\rm cutoff}$ reduces this to order N $R_{\rm cutoff}{}^3$
 - Particle mesh Ewald (PME) method adds long range electrostatics at order N log N, only minor cost compared to cutoff calculation.
- Can we extend the timestep, and do this work fewer times?
 - Bonds to hydrogen atoms, which require a 1fs timestep, can be held at their equilibrium lengths, allowing 2fs steps.
 - Long range electrostatics forces vary slowly, and may be evaluated less often, such as on every second or third step.

Boundary Conditions?



What happens if you put water under vacuum!? Problems: Density, pressure, boundary effects, ... One solution: reflective boundaries, not quite good.

Periodic Boundary Conditions



Spherical boundary conditions



NAMD: The Program we will Use





NAMD programmer J. Phillips Ph. D. UIUC Physics

Simulation of large biomolecular systems

2002 Gordon Bell Award for parallel scalability.

Runs at NSF centers, on clusters, and on desktop.

Available for **FREE** as precompiled binaries; includes source code.

10,000 registered users.



NAMD TUTORIAL



NAMD Developer: James Phillips

Timothy Isgro James Phillips Marcos Sotomayor Elizabeth Villa

Files needed:

structure mypsf.psf coordinates mypdb.pdb

Define temperature

set temperature 310
;# target temperature used several times below

Starting simulation with random velocities

starting from scratch
temperature \$temperature
;# initialize velocities randomly

Continuing a simulation with positions and velocities from previous run

continuing a run
set inputname
binCoordinates
binVelocities
extendedSystem
firsttimestep
numsteps

myinput \$inputname.coor \$inputname.vel \$inputname.xsc 50000 100000

;# only need to edit this in one place!
;# coordinates from last run (binary)
;# velocities from last run (binary)
;# cell dimensions from last run
;# last step of previous run
;# run stops when this step is reached

Organizing output

outputName		myoutput									
;#	base r	name f	or	outp	out	fro	m thi	is r	run		
restartfree	1	50	00	;	;# {	500	step	s =	eve	ry 1	.ps
dcdfreq		50	00								
xstFreq		50	00								
outputEnergies	5	100		;# :	100	ste	eps =	ev	ery	0.2	ps
outputTiming		1000									
;#	shows	time	per	ste	ep a	and	time	to	comj	plet	ion





Underlying Langevin equation for all atoms

$$m_i \frac{d^2 x_i(t)}{dt^2} = F_{i,\text{ff}} - \gamma m_i \frac{d x_i(t)}{dt} + R_i(t)$$

Using periodic boundary conditions avoids surface effects; permits particle mesh Ewald (PME) electrostatics; permits pressure control

<pre># Periodic Boundary cellBasisVector1</pre>	condit	tions 0	0	•# vector to the next image
cellBasisVector1 cellBasisVector2	0.	44.8	0.	,# vector to the next image
cellBasisVector3	0.	0	51.3	
cellOrigin	0.	0.	0.	;# the *center* of the cell
wrapWater	on			;# wrap water to central cell
wrapAll	on			;# wrap other molecules too
wrapNearest	off			;# use for non-rectangular cells

particle mesh Ewald electrostatics (avoids cut-off of long-range Coulomb forces)

#PME (for full-system periodic electrostatics)
PME yes
PMEGridSizeX 32 ;# 2^5, close to 31.2
PMEGridSizeY 45 ;# 3^2 * 5, close to 44.8
PMEGridSizeZ 54 ;# 2 * 3^3, close to 51.3

Fix atoms

fixedAtoms	on	
fixedAtomsFile	myfixedatoms.pdb	;# flags are in this file
fixedAtomsCol	В	;# set beta non-zero to fix an atom

Energy-minimize structure (T=0), reset temperature T, run:

minimize 1000 ;# lower potential energy for 1000 steps
reinitvels \$temperature ;# since minimization zeros velocities
run 50000 ;# 100ps

The NAMD Output File / 1

Preamble

Info: NAMD 2.5b2ss03 for Linux-i686-Clustermatic Info: Info: Please visit http://www.ks.uiuc.edu/Research/namd/ Info: and send feedback or bug reports to namd@ks.uiuc.edu Info: Info: Please cite Kale et al., J. Comp. Phys. 151:283-312 (1999) Info: in all publications reporting results obtained with NAMD. Info: Info: Built Fri May 30 13:09:06 CDT 2003 by jim on umbriel Info: Sending usage information to NAMD developers via UDP. Info: Sent data is: 1 NAMD 2.5b2ss03 Linux-i686-Clustermatic 47 umbriel jim Info: Running on 47 processors.

The NAMD Output File / 2

Energies

ETITLE:	TS	BOND	ANGLE	DIHED	IMPRP
	ELECT	VDW	BOUNDARY	MISC	KINETIC
	TOTAL	TEMP	TOTAL2	TOTAL3	TEMPAVG
F	RESSURE	GPRESSURE	VOLUME	PRESSAVG	GPRESSAVG
ENERGY:	1000	0.0000	0.0000	0.0000	0.0000
-970)22.1848	9595.3175	0.0000	0.0000	14319.5268
-731	07.3405	300.2464	-73076.6148	-73084.1411	297.7598
-6	626.5205	-636.6638	240716.1374	-616.5673	-616.6619

The NAMD Output File / 1

Writing out trajectories

OPENING COORDINATE DCD FILE WRITING COORDINATES TO DCD FILE AT STEP 1000

Performance information

Info: Benchmark time: 47 CPUs 0.0475851 s/step 0.275377 days/ns 13540 kB memory

TIMING: 1000 CPU: 18.35, 0.01831/step Wall: 50.1581, 0.0499508/step, 6.92374 hours remaining, 14244 kB of memory in use.

Warnings

Warning: Pairlistdist is too small for 1 patches during timestep 17. Warning: Pairlists partially disabled; reduced performance likely. Warning: 20 pairlist warnings since previous energy output.

The NAMD Experience / 1

You will first simulate ubiquitin in a water sphere and water box:

Generating a Protein Structure File (PSF)

- Go to 1-1-build directory
- Open VMD, choose extension TkCon
- Make from 1UBQ.pdb a structure without hydrogens, ubqp.pdb
- Create psf file for ubqp.pdb: ubq.pdb and ubq.psf
- Check if files exist

Solvate the protein in a water sphere (from VMD)



Solvate the protein in a water box (from VMD)



The NAMD Experience / 2

- **RMSD** value for equilibration
- Atomic RMSD values of equilibrated protein
- Velocity distribution
- Temperature distribution
- Specific heat
- Diffusion of whole protein
- Heat diffusion
- Temperature Echoes

NAMD Tutorial (Part 2)

2 Analysis

- > 2.1 Equilibrium
 - > 2.1.1 RMSD for individual residues
 - 2.1.2 Maxwell-Boltzmann Distribution
 - > 2.1.3 Energies
 - > 2.1.4 Temperature distribution
 - > 2.1.5 Specific Heat

Maxwell Distribution of Atomic Velocities



 $\sigma = x, y, z$



