Part II - Applications of MultiSeq: Network Analysis of Dynamical Recognition in RNA:Protein Complexes

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Protein: RNA Complexes in Translation Evolutionary Analysis & Dynamics





r-Proteins/r-RNA "Signatures ribosomate Volution"

"Signatures Vibosontal evolution" **PNAS** 2008, **BMC** 2009, **BJ** 2010 "Motion L1 Stalk:tRNA" **JMB** 2010, "Ribosome Biogenesis" **JPC** 2012,3 "Whole cell simulations on GPUs" **IEEE** 2009,**Plos CB** 2011,**PRL**2011, **JCC** 2013, **PNAS** 2013, 2010 **PRL** 2013, **CSB** 2013 **C** 2012 **Nature** 2014

Molecular Dynamics Simulations

MD performed with NAMD2 (1) - System Setup

Simulation Parameters

Minimization: 290,000 steps Production run: 108 ns Forcefields: CHARMM27 (2), AMBER (3) Time step: 1 fs VdW frequency: 2 fs VdW cutoff: 12 Å Switching distance: 10 Å Pair list distance: 14 Å Particle Mesh Ewald Full electrostatic update: 4 fs Ensemble: NPT Langevin temperature: 298.15 K Langevin pressure: 1 atm Periodic boundary conditions

Contents of System(4) GluRS Glu-tRNA^{Glu} EF-Tu GTP Ions: Mg²⁺, K⁺ H₂O: ~27,000 molecules System: ~130,000 atoms

System Perturbations

Deprotonation/protonation of reactants (aa,NTs) Changing rotamer states of residues (5) involved in long-lived salt bridges

- (1) Phillips, J.C. et al. *J. Comput Chem*, (2005); (2) MacKerell, A. et al. *Biopolymers* (2001);
- (3) Case, D. et al. J. Comput. Chem. (2005); (4) Eargle, J. et al. JMB 2010, FEBS Let. 2010;
- (5) Dunbrack Jr. and Cohen. Protein Sci. (1997)

Charging tRNA through allosteric signaling



How to Construct a Network?



Nodes - defined at C_a (protein) and P (nucleotide) atoms

How to Construct a Network?



Edges - connect nodes that are within a contact distance threshold for more than 75% of an MD trajectory



Path length = 10 (*unweighted*)

Information transfer? Weight contacts/links by correlations !

Correlations (C_{ij}) define signaling pathways in GluRS:tRNA



- C_{ii} values calculated over a 16-ns window

Nodes Cluster Together in Modules called Communities



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Communities are subnetworks with many intracommunity edges but few intercommunity edges. (Girvan-Newman Algorithm)



Reaction Mechanism for the Transfer of Glu to tRNA^{Glu}

Perona JJ, Rould MA, Steitz TA Biochemistry 1993

Black A, Eargle J, Sethi A, Luthey-Schulten Z. *JMB* 2010 100s ns MD simulations









community containing amino acid moiety

community containing AMP moiety







Communities Partition the Interaction Network Hierarchically



Communities Partition the Interaction Network Hierarchically



Dynamical Networks, Conservation, and Betweeness



Optimal signal pathways: U13, U35 to A76 Critical(conserved) nodes connecting communities

Betweeness routes - highest density pair optimal paths

Network Viewer in VMD*



* J. Eargle and Z. Luthey-Schulten, Bioinformatics, 2012, 28, 3000–3001

Changing Networks: tRNA Migration from GluRS to EF-Tu

>2-fold increase in k_{cat} for CysRS:tRNA:EF-Tu

Zhang C, Perona J, Kang R, Francklyn C, Hou Y. *JMB* 2006 Hausmann C, Praetorius-Ibba M, Ibba M. *NAR* 2007

8-fold increase

in k_{cat} for

LeuRS:tRNA:EF-Tu

Change in Protein: RNA Contacts as tRNA Migrates Glu-TRNA Glu C_s / GluRS Arg205 C_z







Change in Protein:RNA Contacts During tRNA Migration



FEBS 2012

yellow - T arm community

purple - D arm community

Evolution of Protein/RNA Interfaces: Dynamics of EF-Tu/tRNA Recognition



J. Eargle, et al. JMB 2008

Dynamical Recognition EF-Tu/tRNA (E.coli)



J. Eargle, A. Sethi, A. Black, L. Trabuco & Z. Luthey-Schulten. JMB "Dynamics of Recognition in EF-Tu/tRNA Complex" (2008)



Effects of Modified Bases on Dynamics of AC Stem/Loop

Similar NMR structures for unmodified tRNA-Phe Nikonowicz, *JMB* 2002, *personal communication*

Eargle, et al. JMB (2008))

Flexibility in tRNA structures observed in crystallography and simulations

X-ray 5 tRNAs

MD EF-Tu:tRNA:cys

MD: RMSD and Interarm Angle in tRNA:cys

JMB 2008 & FEBS Let. 2010

Molecular Signatures in Evolution of Translation

Universal Phylogenetic Tree

Dynamical function of ribosomal signatures: idiosyncrasies in ribosomal RNA and/or proteins characteristic of the domains of

life Roberts, ... Woese, Luthey-Schulten (2008) *PNAS*; Chen,... Gruebele, Luthey-Schulten (2010) *BJ* Chen, ...Ha, Woodson, Luthey-Schulten, (2012) *JPCB*; Lai, Chen, Luthey-Schulten (2013) *JPCB* Kim,... Luthey-Schulten Z., Ha, and Woodson (2014) *Nature* "Protein-guided RNA dynamics during early ribosome assembly"

Sequence and **Structure** Signatures in Ribosomal RNA

Signature analysis

- Signatures are sequence and/or structural features that are characteristic of a domain of life^{*}.
- Identify the sequence signatures in the 16S rRNA and proteins using MultiSeq
- MultiSeq has a coloring based on signatures, but use Scripts to collect quantitative data about the signatures: position, composition.
- Signature analysis can be applied to any set of sequences/groupings

90,000 Environmental 165 rRNA Distinct A & B Sequence Signatures

• Analysis of the ribosomal signatures in 90,000 new environmental samples shows that no "gray" area exists: a ribosome is either bacterial or archaeal in nature.

• Split across cluster: 10,000 sequence on each node

Data: "Greengenes", Lawrence Berkeley

Largest signature region in the SSU

5-way helical junction rRNA with S4

Model for Ribosome Assembly – Structural Intermediates

Collaboration with Gruebele, Ha, Woodson labs

VMD/MultiSeq Tutorials

- 1. Evolution of Translation: AARS: tRNA
- 2. Evolution of Translation: EF-Tu:tRNA
- 3. Evolution of Translation: Ribosome
- 4. Dynamical Network Analysis
- 5. Hybrid MD-GO Folding of RNA