Part II - Applications of MultiSeq Evolution of Translation: Dynamics of Recognition in RNA:Protein Complexes

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Biology

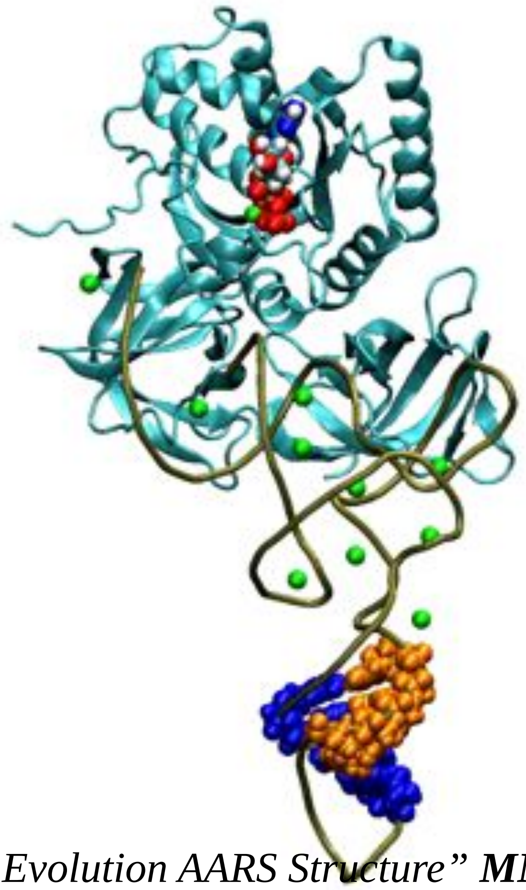
Workshop 2012 – UIUC Beckman

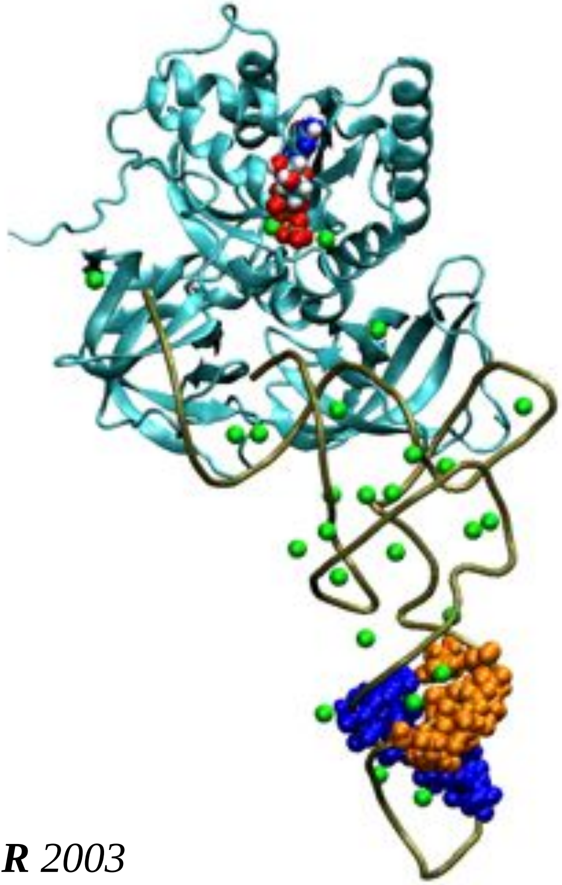
NIH Center Macromolecular Modeling and Bioinformatics

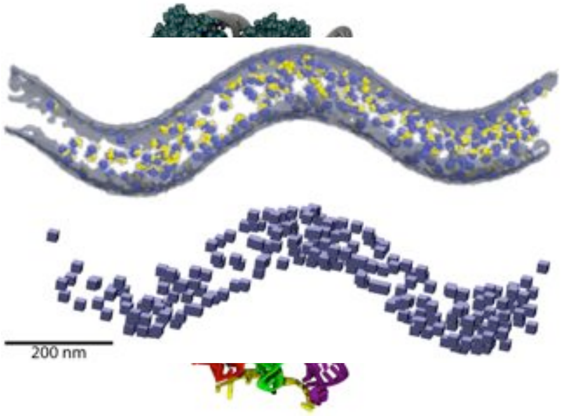
ILLINOIS
UNIVERSITY OF ILLINOIS AT URBANA-CHAMPAIGN

Protein:RNA Complexes in Translation

Evolution, Dynamics, Analysis







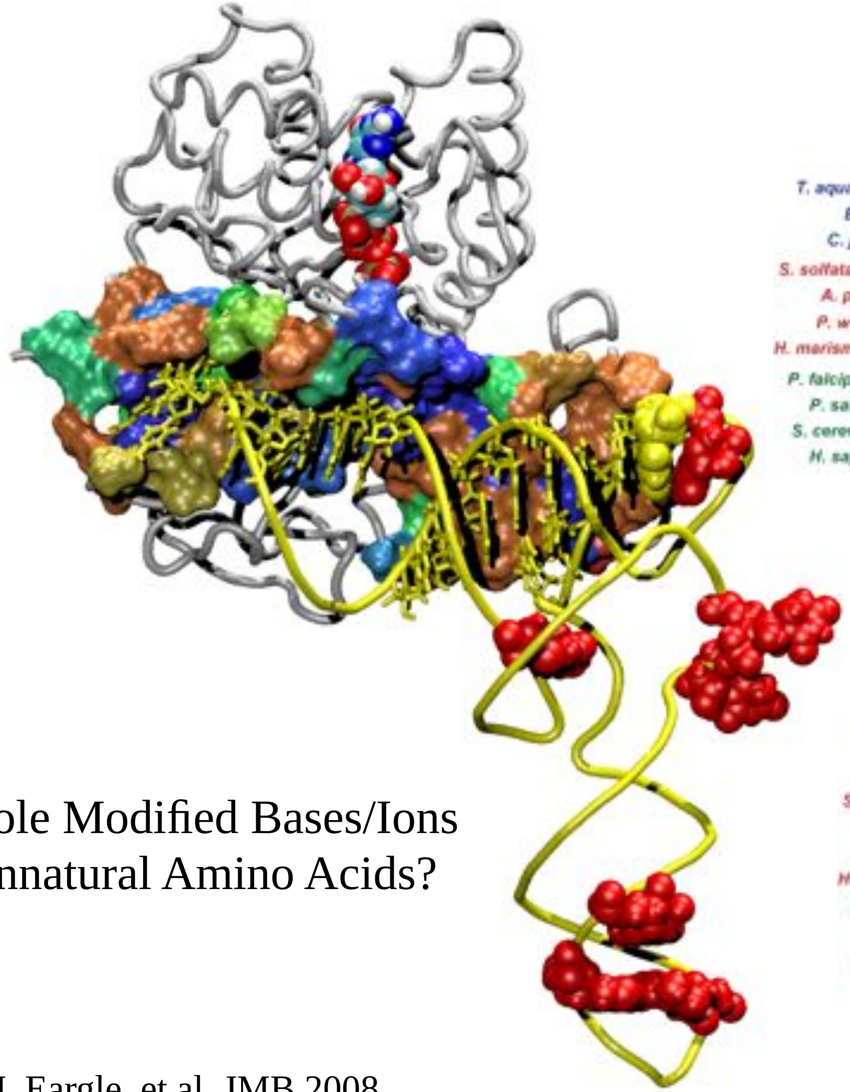
Proteins/RNA
Polyribosomes
Ribosome

“Evolution AARS Structure” ***MMBR*** *2003*
“Evol. Profiles Class I&II AARS” ***JMB*** *2005*
“Evolution SepRS/CysRS” ***PNAS*** *2005*
“Dynamic Signaling Network” ***PNAS*** 2009
“Exit Strategy Charged tRNA” ***JMB*** 2010
“Mistranslation in Mycoplasma” ***PNAS*** 2011

“Dynamical Recognition Novel Amino Acids” ***JMB*** 2008
“tRNA Dynamics” ***FEBS*** *2010*

“Signatures ribosomal evolution”
PNAS *2008*, ***BMC*** *2009*, ***BJ*** *2010*
“Motion L1 Stalk:tRNA” ***JMB 2010***
“Whole cell simulations on GPUs“
IEEE *2009*,***Plos CB*** *2011*,***PRL****2011*

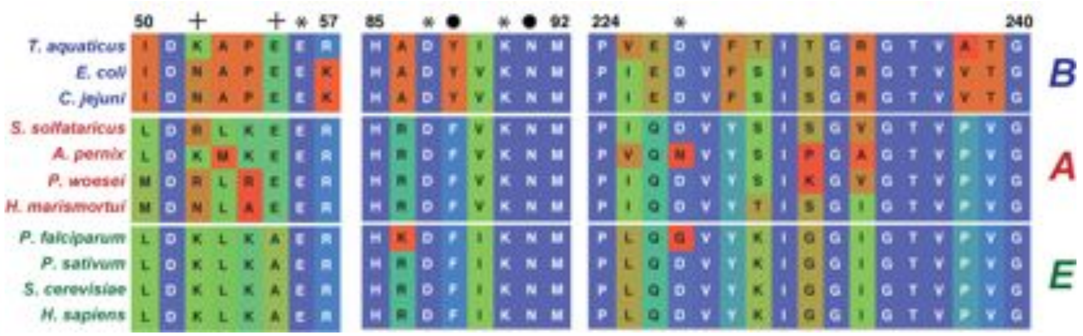
Evolution of Protein/RNA Interfaces: Dynamics of EF-Tu/tRNA Recognition



Role Modified Bases/Ions
Unnatural Amino Acids?

J. Eargle, et al. JMB 2008

Switch I Switch II Domain II



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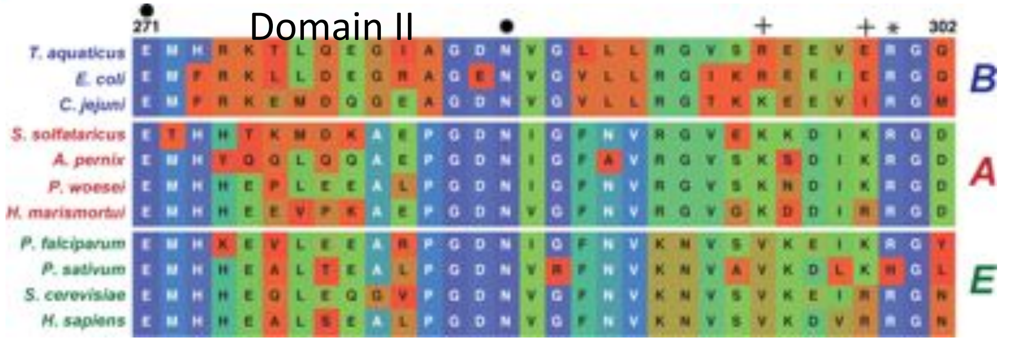
T. aquaticus	I	D	K	A	P	E	E	R	H	A	D	Y	I	K	N	M	P	V	E	D	V	F	T	I	T	G	R	G	T	V	A	T	G	B
E. coli	I	D	N	A	P	E	E	K	H	A	D	Y	V	K	N	M	P	I	E	D	V	F	S	I	S	G	R	G	T	V	V	T	G	
C. jejuni	I	D	N	A	P	E	E	K	H	A	D	Y	V	K	N	M	P	I	E	D	V	F	S	I	S	G	R	G	T	V	V	T	G	
S. solfataricus	L	D	R	L	K	E	E	R	H	R	D	F	V	K	N	M	P	I	Q	D	V	Y	S	I	S	G	V	G	T	V	P	V	G	A
A. pernix	L	D	K	M	K	E	E	R	H	R	D	F	V	K	N	M	P	V	Q	N	V	Y	S	I	P	G	A	G	T	V	P	V	G	
P. woesei	M	D	R	L	R	E	E	R	H	R	D	F	V	K	N	M	P	I	Q	D	V	Y	S	I	K	G	V	G	T	V	P	V	G	
H. marismortui	M	D	N	L	A	E	E	R	H	R	D	F	V	K	N	M	P	I	Q	D	V	Y	T	I	S	G	I	G	T	V	P	V	G	
P. falciparum	L	D	K	L	K	A	E	R	H	K	D	F	I	K	N	M	P	L	Q	G	V	Y	K	I	G	G	I	G	T	V	P	V	G	E
P. sativum	L	D	K	L	K	A	E	R	H	R	D	F	I	K	N	M	P	L	Q	D	V	Y	K	I	G	G	I	G	T	V	P	V	G	
S. cerevisiae	L	D	K	L	K	A	E	R	H	R	D	F	I	K	N	M	P	L	Q	D	V	Y	K	I	G	G	I	G	T	V	P	V	G	
H. sapiens	L	D	K	L	K	A	E	R	H	R	D	F	I	K	N	M	P	L	Q	D	V	Y	K	I	G	G	I	G	T	V	P	V	G	

Domain II



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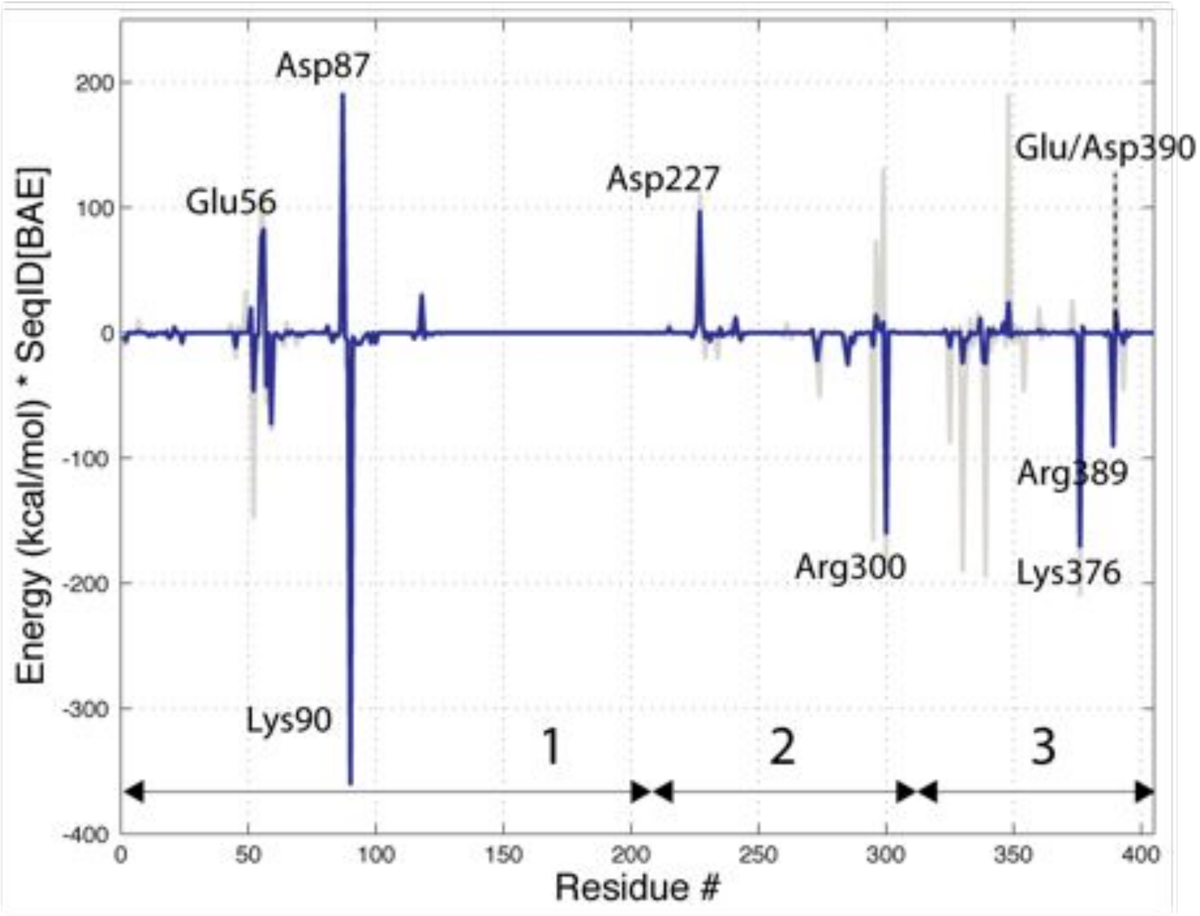
T. aquaticus	E	M	H	R	K	T	L	Q	E	G	I	A	G	D	N	V	G	L	L	L	R	G	V	S	R	E	E	V	E	R	G	Q	B
E. coli	E	M	F	R	K	L	L	D	E	G	R	A	G	E	N	V	G	V	L	L	R	G	I	K	R	E	E	I	E	R	G	Q	
C. jejuni	E	M	F	R	K	E	M	D	Q	G	E	A	G	D	N	V	G	V	L	L	R	G	T	K	K	E	E	V	I	R	G	M	
S. solfataricus	E	T	H	H	T	K	M	D	K	A	E	P	G	D	N	I	G	F	N	V	R	G	V	E	K	K	D	I	K	R	G	D	A
A. pernix	E	M	H	Y	Q	Q	L	Q	Q	A	E	P	G	D	N	I	G	F	A	V	R	G	V	S	K	S	D	I	K	R	G	D	
P. woesei	E	M	H	H	E	P	L	E	E	A	L	P	G	D	N	I	G	F	N	V	R	G	V	S	K	N	D	I	K	R	G	D	
H. marismortui	E	M	H	H	E	E	V	P	K	A	E	P	G	D	N	V	G	F	N	V	R	G	V	G	K	D	D	I	R	R	G	D	
P. falciparum	E	M	H	K	E	V	L	E	E	A	R	P	G	D	N	I	G	F	N	V	K	N	V	S	V	K	E	I	K	R	G	Y	E
P. sativum	E	M	H	H	E	A	L	T	E	A	L	P	G	D	N	V	R	F	N	V	K	N	V	A	V	K	D	L	K	H	G	L	
S. cerevisiae	E	M	H	H	E	Q	L	E	Q	G	V	P	G	D	N	V	G	F	N	V	K	N	V	S	V	K	E	I	R	R	G	N	
H. sapiens	E	M	H	H	E	A	L	S	E	A	L	P	G	D	N	V	G	F	N	V	K	N	V	S	V	K	D	V	R	R	G	N	

Dynamical Recognition EF-Tu/tRNA (E.coli)

Combining MD with Evolutionary Analysis (MultiSeq)

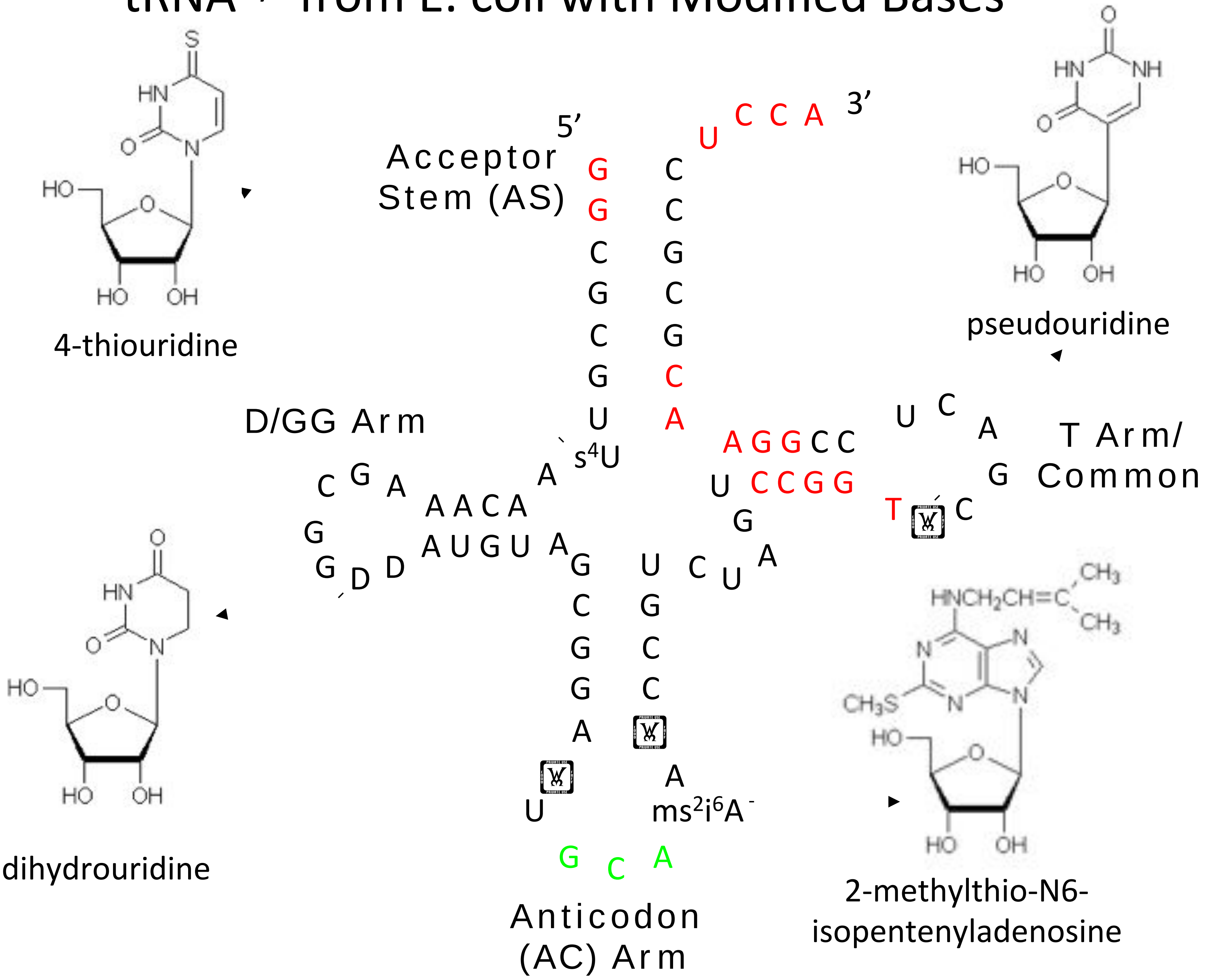
Non-bonded Interaction Energy



BAE

J. Eargle, A. Sethi, A. Black, L. Trabuco & Z. Luthey-Schulten. JMB “Dynamics of Recognition in EF-Tu/tRNA Complex” (2008)

tRNA^{Cys} from E. coli with Modified Bases



Acceptor Stem (AS)

D/GG Arm

T Arm/ Common

Anticodon (AC) Arm

4-thiouridine

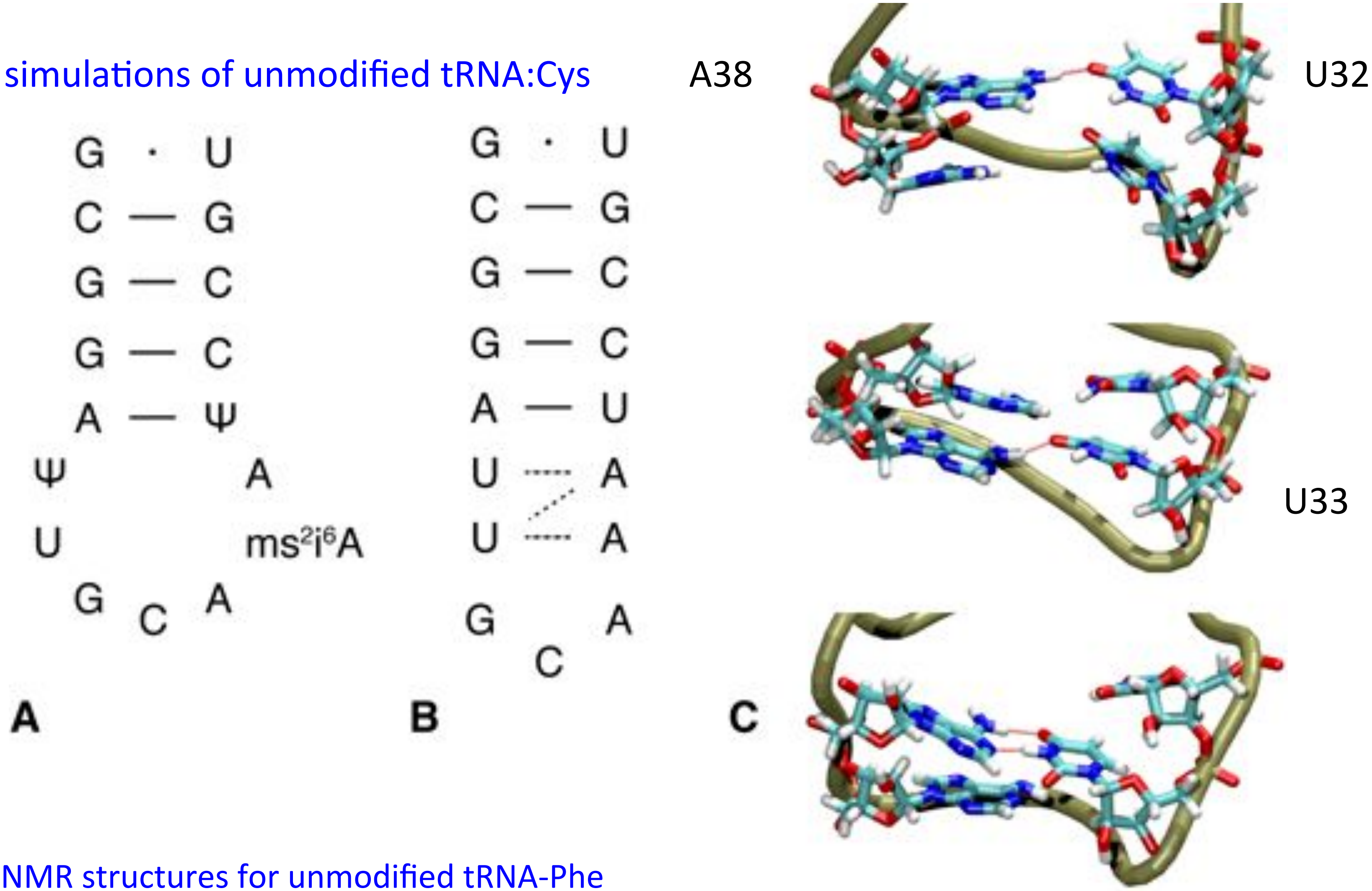
pseudouridine

dihydrouridine

2-methylthio-N6-isopentenyladenosine

Effects of Modified Bases on Dynamics of AC Stem/Loop

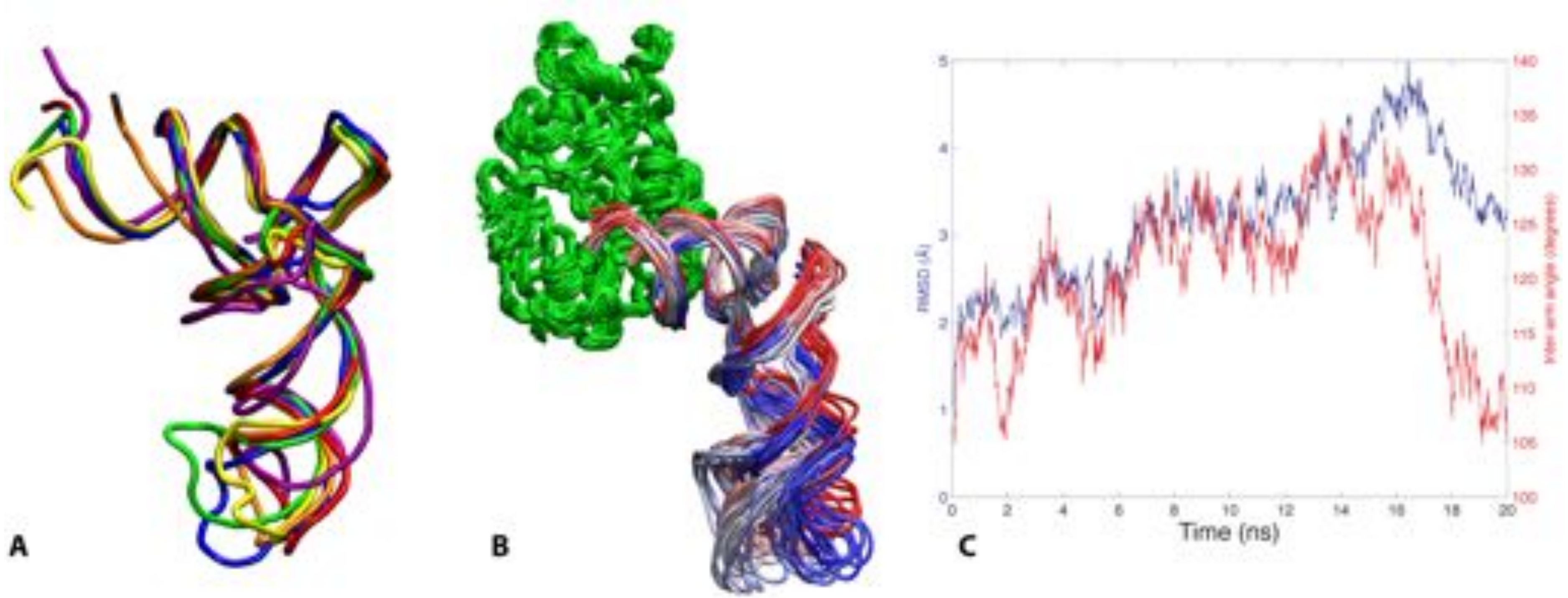
MD simulations of unmodified tRNA:Cys



Similar NMR structures for unmodified tRNA-Phe
Nikonowicz, *JMB* 2002, *personal communication*

Eargle, et al. *JMB* (2008))

Flexibility in tRNA structures observed in crystallography and simulations



X-ray 5 tRNAs

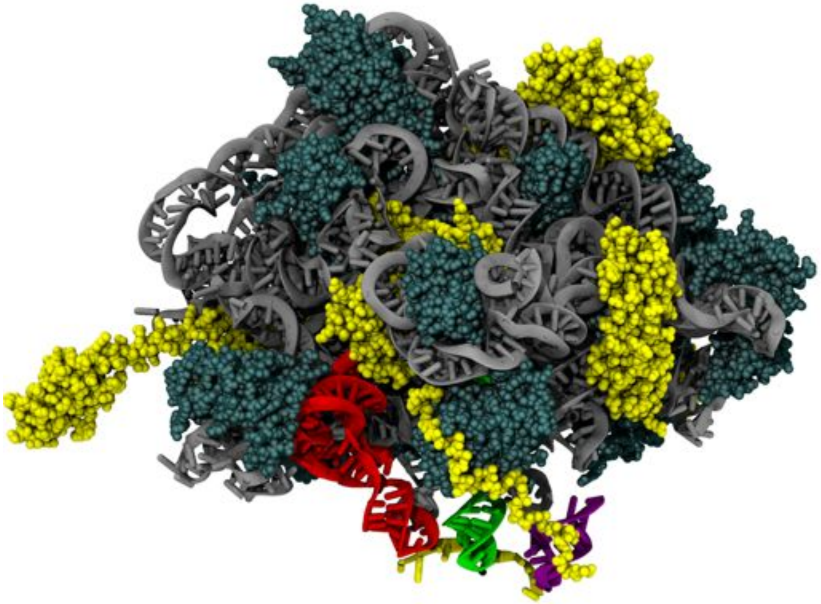
MD EF-Tu:tRNA:cys

MD: RMSD and Interarm
Angle in tRNA:cys

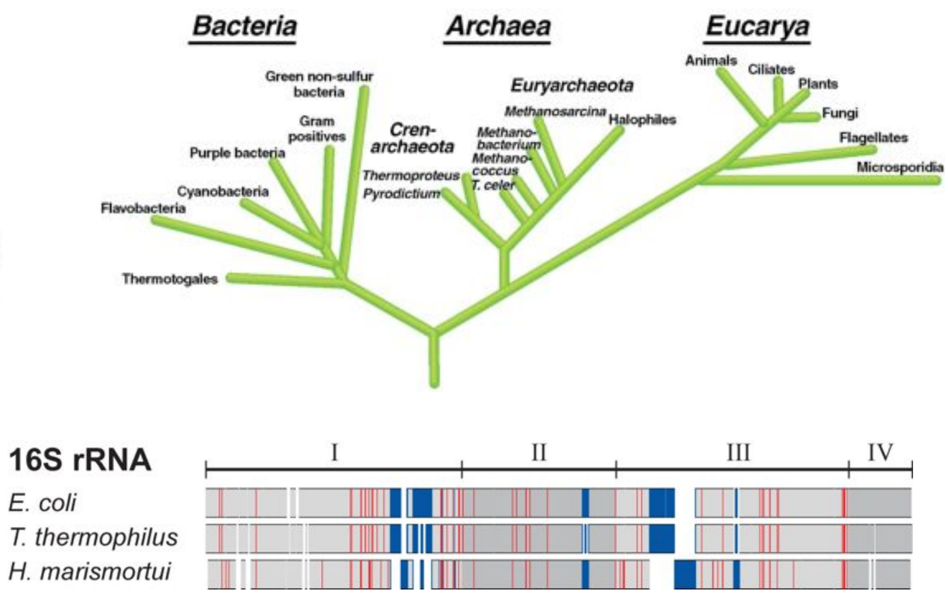
JMB 2008 & FEBS Let. 2010

Molecular Signatures in Ribosome Evolution

Archaeal - LSU



Universal Phylogenetic Tree



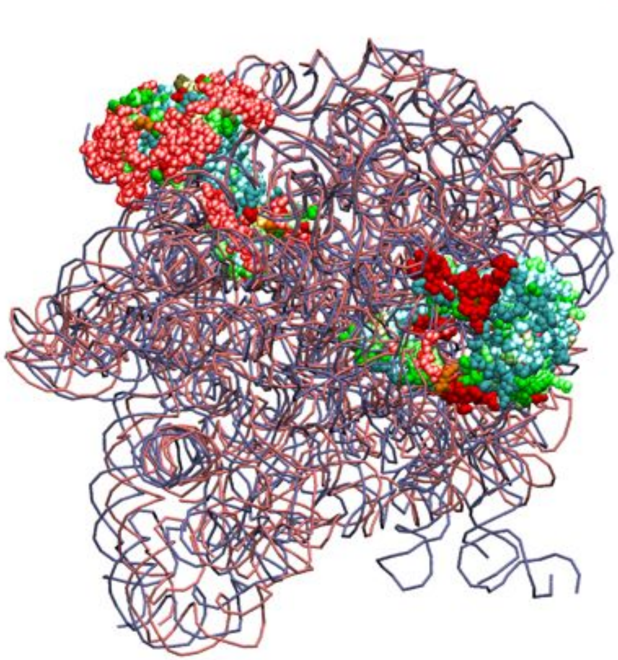
Dynamical function of ribosomal signatures: idiosyncrasies in ribosomal RNA and/or proteins characteristic of the domains of life

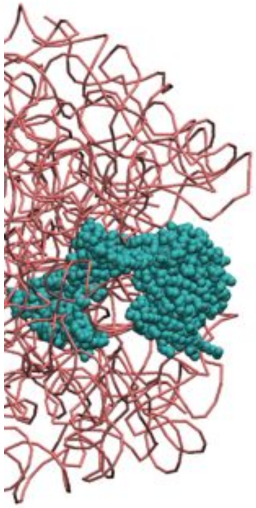
Roberts, Sethi, Montoya, Woese, Luthey-Schulten (2008) ***PNAS*** 105:13953
Chen, Eargle, Sarkar, Gruebele, Luthey-Schulten (2010) ***Biophysical Journal***

Structural Overlaps with STAMP

Comparing ribosomal large subunit with r-proteins L2 and L3
180,000 atoms in 4 rRNAs and 58 proteins

E. col





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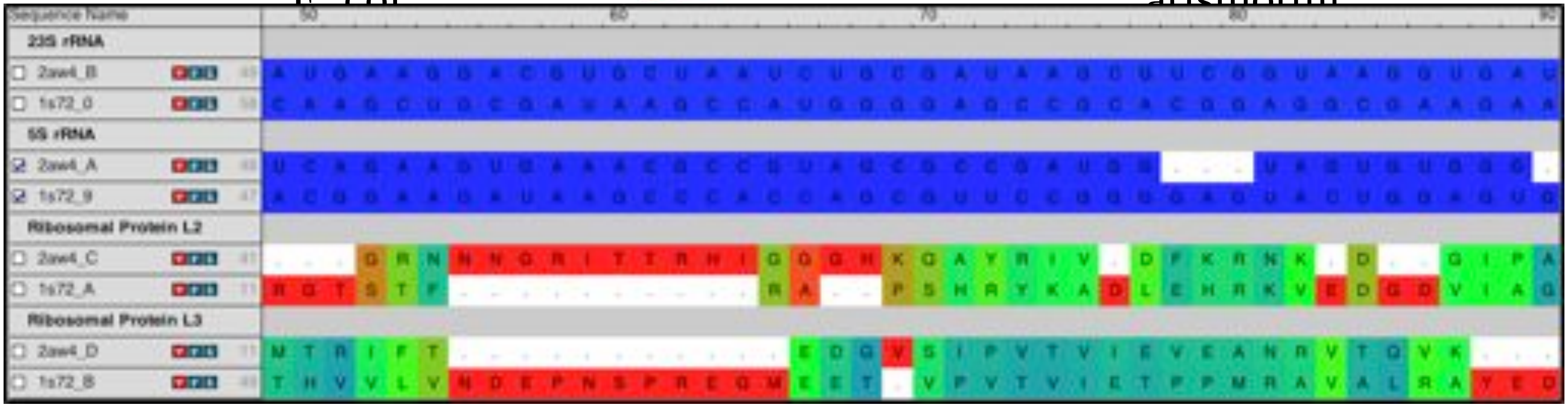


Sequence Name 50 60 70 80 90

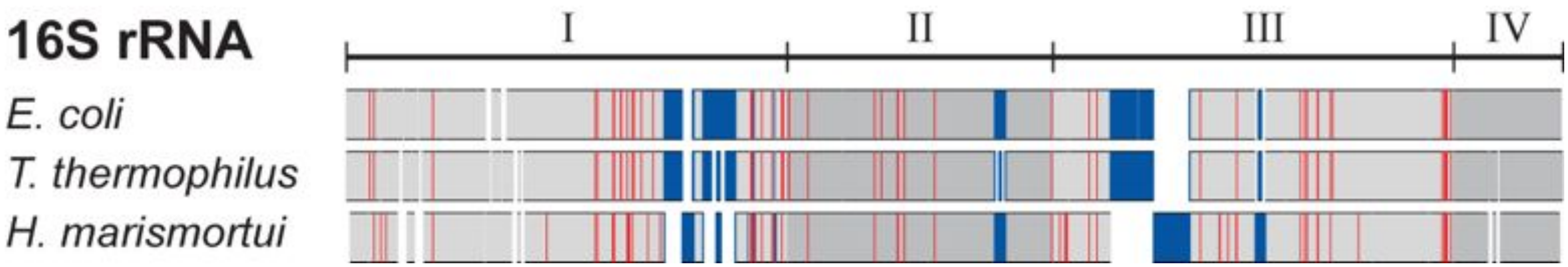
23S rRNA
☐ 2aw4_B
☐ 1s72_0

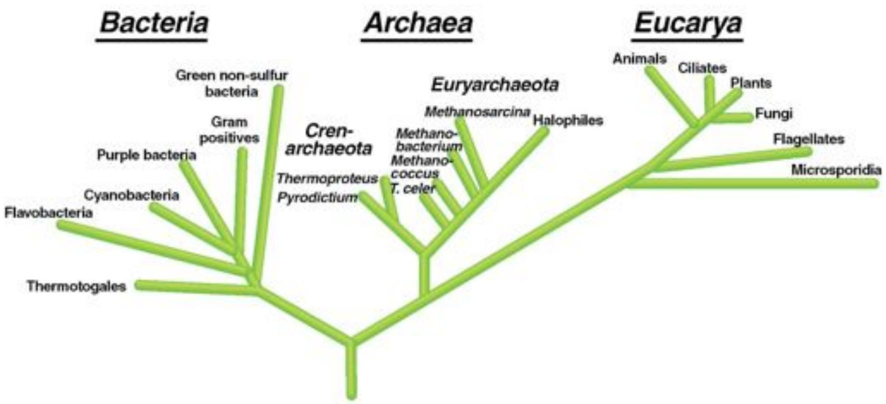
5S rRNA
☑ 2aw4_A
☑ 1s72_9

Ribosomal Protein L2
☐ 2aw4_C - - - G R N N N G R I T T R H I G G G H K G A Y R I V - D F K R N K - D - - G I P A
☐ 1s72_A R G T S T F - - - - - - - - - - R A - - P S H R Y K A D L E H R K V E D G D V I A G

Ribosomal Protein L3
☐ 2aw4_D M T R I F T - - - - - - - - - - - E D G V S I P V T V I E V E A N R V T Q V K - - -
☐ 1s72_B T H V V L V N D E P N S P R E G M E E T - V P V T V I E T P P M R A V A L R A Y E D

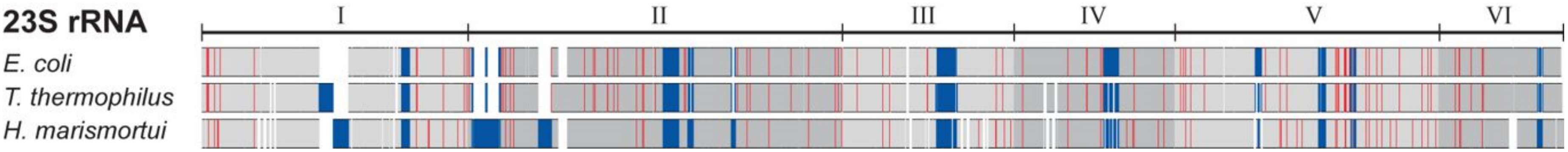
Sequence and Structure Signatures in Ribosomal RNA





Ribosomal Signatures: Idiosyncrasies in rRNA and/or rproteins characteristic of the domains of life

69 (119) & **6 (14)** in 16S (23S)



Roberts, Montoya, Sethi, Woese, ZLS, *PNAS* 2008

Signature analysis

- Signatures are sequence and/or structural features that are characteristic of a domain of life*.
- Identify the sequence signatures in the 16S rRNA and proteins using MultiSeq
- MultiSeq has a coloring based on signatures, but use Scripts to collect quantitative data about the signatures: position, composition.
- Signature analysis can be applied to any set of sequences/groupings

* E. Roberts, A. Sethi, J. Montoya, C. Woese, Z Luthey-Schulten (2008) *PNAS*

Signature analysis script

```
proc get_taxa_signatures {args} {

    initializeMultiSeqEnvironment

    # Load the alignment.
    set sequenceIDs [::SeqData::Fasta::loadSequences $alignmentFilename]

    # Find the reference species.
    set refSequenceIDs {}
    foreach species $referenceSpecies {
        set found 0
        foreach sequenceID $sequenceIDs {
            if {[string toupper [::SeqData::getScientificName $sequenceID]] == [string toupper $species]} {
                lappend refSequenceIDs $sequenceID
                set found 1
                break
            }
        }
    }

   # Group the sequence ids.
    array set groupData {names {}}
    foreach sequenceID $sequenceIDs {

        set group ""
        if {$rankName == "domain"} {
            set group [::SeqData::getDomainOfLife $sequenceID]
        } else {
            set group [::SeqData::getLineageRank $sequenceID $rankName]
        }
        if {![info exists groupData($group)]} {
            lappend groupData(names) $group
            set groupData($group) {}
        }
        lappend groupData($group) $sequenceID
    }
```

Signature analysis script (cont)

```
    # Calculate the signatures for the groups.
    set signatures [::SeqEdit::Metric::Signatures::calculateSignatures $sequenceGroups $groupConsensusCutoff \
                 $otherGroupMaxCutoff $otherGroupMaxGapFraction $maxConservedBlockDistance $minConservedBlockSize]

    # Print the signatures.
    puts ""
    puts "Ordered Signatures"
    printSignatureHeader $groups $refSequenceIDs
    for {set position 0} {$position < [llength [lindex $signatures 0]]} {incr position} {

        # Get the signature for each of the groups.
        set isSignature 0
        for {set groupIndex 0} {$groupIndex < [llength $groups]} {incr groupIndex} {
            set groupSignature [lindex [lindex $signatures $groupIndex] $position]
            if {[string length $groupSignature] == 1} {
                set isSignature 1
            }
        }

        if {$isSignature} {
            printSignatureLine $position $groups $refSequenceIDs $signatures $sequenceGroups
        }
    }
}
```

Signature analysis script (cont)

```
proc getCompositionString {sequenceIDs position} {

    # Go through each group to count the elements.
    array set counts {values {}}
    foreach sequenceID $sequenceIDs {
        set element [::SeqData::getElement $sequenceID $position]
         if {![info exists counts($element)]} {
             lappend counts(values) $element
             set counts($element) 1
         } else {
             incr counts($element)
         }
    }

    set ret ""
    foreach element $counts(values) {
        set percentage [expr round(100.0*double($counts($element))/double([llength $sequenceIDs]))]
        if {$percentage > 5} {
            if {$ret != ""} {
                append ret "/"
            }
            append ret "${element}($percentage\%)"
        }
    }

    return $ret
}
```

Signature script output

```
Ordered Signatures
  Pos  E._coli T._therm H._maris   Archaea                        Bacteria
  --- -------- -------- -------- ------------------------------ ------------------------------
   35       33       33       27   U(28%)/C(72%)                  *A(96%)
   42       39       39       33   *U(100%)                       *G(95%)
   51       47       47       42   *G(97%)                        C(85%)/U(7%)/G(8%)
   59       53       53       49   *C(100%)                       A(83%)/G(14%)
  222      113      106       87   *C(100%)                       *G(95%)
  661      248      243      227   *G(97%)                        C(75%)/G(9%)/U(11%)
  749      314      309      293   *G(100%)                       *C(97%)
  753      317      312      296   *G(97%)                        *U(91%)/G(7%)
  777      338      333      317   *G(97%)                        A(89%)/N(9%)
  779      339      334      318   *G(97%)                        *C(98%)
  780      340      335      319   *C(94%)                        *U(92%)/A(6%)
  790      349      344      328   G(86%)/N(11%)                  *A(92%)
  791      350      345      329   C(67%)/U(19%)/N(11%)           *G(98%)
  800      358      353      337   *G(100%)                       *U(91%)/C(7%)
  802      359      354      338   *C(97%)                        G(59%)/U(9%)/C(9%)/A(20%)
  804      361      356      340   *C(94%)                        G(84%)/A(14%)
  809      365      360      344   *A(92%)/U(8%)                  *U(97%)
  811      367      362      346   C(83%)/G(14%)                  *U(97%)
  823      377      372      356   C(89%)/U(11%)                  *G(92%)
  833      386      381      365   *G(94%)                        C(89%)/U(6%)
  840      393      388      372   G(83%)/C(14%)                  *A(97%)
  851      403      398      382   *A(100%)                       C(89%)
  920      447      441      416   C(31%)/U(67%)                  *G(92%)
  990      487      471      424   G(89%)/A(8%)                   *A(91%)
 1083      508      491      446   *A(97%)                        *U(92%)
 1085      510      493      448   *G(97%)                        *A(97%)
 1087      512      495      450   *C(92%)/U(8%)                  *U(97%)
 1089      514      497      452   *G(92%)/N(6%)                  C(89%)/U(8%)
 1101      523      506      461   *C(94%)/A(6%)                  *A(96%)
 1119      537      520      475   *C(100%)                       *G(91%)/A(8%)
 1121      539      522      477   *G(92%)/A(6%)                  A(89%)/G(10%)
 1134      549      532      487   *U(94%)                        *C(99%)
```

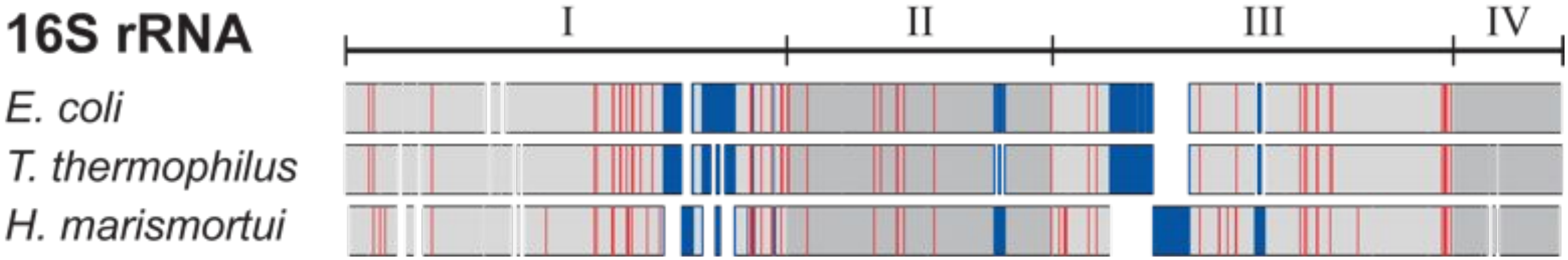
Draw signature figure

- Want to create a graphical representation of the signatures.
- Can use the TCL graphics object to draw figures.

```
proc draw_one_d_alignment {args} {

    # Load the alignment.
    set alignment [::SeqData::Fasta::loadSequences $alignmentFilename]
    set numberSequences [llength $alignment]
    set numberPositions [::SeqData::getSeqLength [lindex $alignment 0]]
    puts "Loaded $numberSequences sequences of length $numberPositions"

    # Create the drawing canvas.
    set canvasWidth [expr $draw(xBorder)+($numberPositions*$draw(positionWidth))+$draw(xBorder)]
    set canvasHeight [expr $draw(yBorder)+($numberSequences*$draw(sequenceHeight))+(($numberSequences-1)*\
                                                                              $draw(sequenceSpacing))+$draw(yBorder)]
    set g [canvas .drawing -width $canvasWidth -height $canvasHeight]

    # Draw the alignment.
    for {set i 0} {$i < $numberSequences} {incr i} {
        set sequenceID [lindex $alignment $i]
        set y1 [expr $draw(yBorder)+($i*($draw(sequenceHeight)+$draw(sequenceSpacing)))]
        set y2 [expr $y1+$draw(sequenceHeight)]
        drawSequencePositions $g $draw(xBorder) $y1 $y2 $sequenceID 0 [expr $numberPositions-1] \
                                    $draw(positionWidth) $draw(sequenceColor) $draw(borderColor) $draw(borderWidth)
    }

    # Save the canvas.
    $g postscript -x 0 -y 0 -width $canvasWidth -height $canvasHeight -file $outputFilename

    cleanupMultiSeqEnvironment
}
```

Draw signature figure (cont)

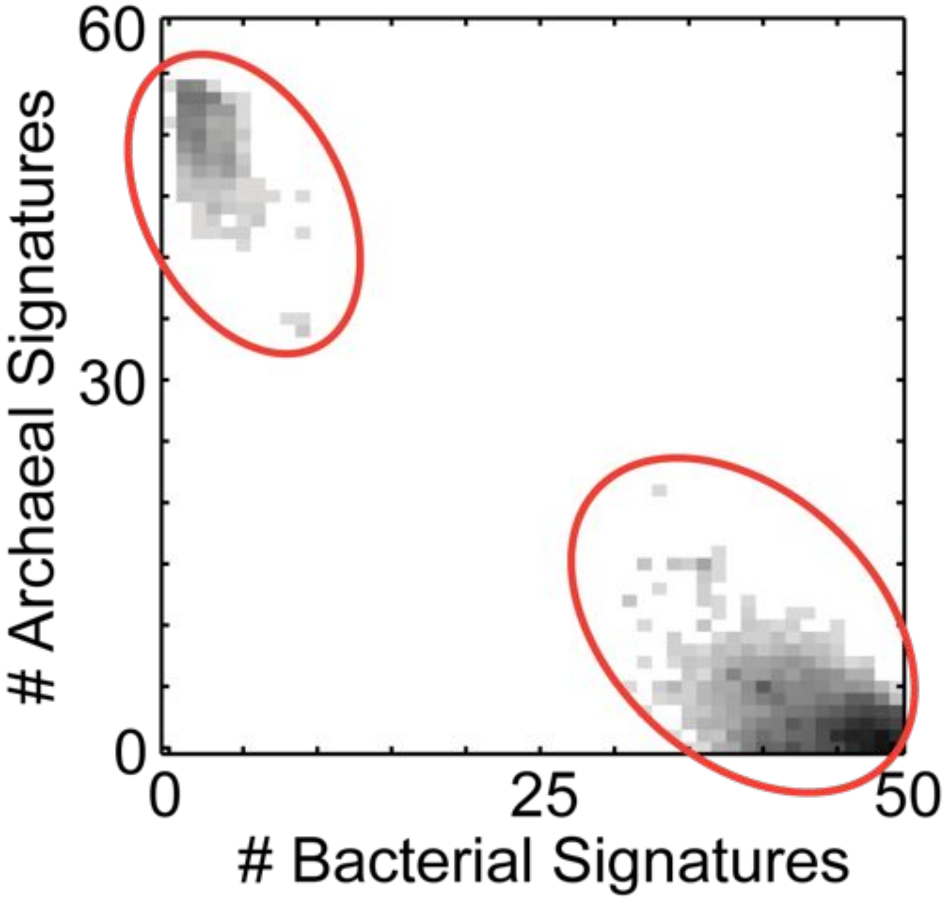
```
proc drawSequencePositions {g x y1 y2 sequenceID firstPosition lastPosition positionWidth sequenceColor borderColor borderWidth}
{
    set startPosition ""
    for {set j $firstPosition} {$j <= $lastPosition} {incr j} {
        set element [::SeqData::getElement $sequenceID $j]

        # If this is the beginning of a sequence block, record that one started.
        if {$element != "-" && $startPosition == ""} {
            set startPosition $j
        }

        # If this is the end of a sequence block, draw the block.
        if {($element == "-" || $j == $lastPosition) && $startPosition != ""} {
            if {$element == "-"} {
                set endPosition [expr $j-1]
            } else {
                set endPosition [expr $j]
            }
            set x1 [expr $x+($startPosition*$positionWidth)]
            set x2 [expr $x+(($endPosition+1)*$positionWidth)]
            $g create rectangle $x1 $y1 $x2 $y2 -fill $sequenceColor -outline $sequenceColor -width $borderWidth
            $g create line $x1 $y1 $x2 $y1 -fill $borderColor -width $borderWidth
            $g create line $x1 $y2 $x2 $y2 -fill $borderColor -width $borderWidth
            set startPosition ""
        }
    }
}
```



90,000 Environmental 16S rRNA
Distinct A & B Sequence Signatures

- Analysis of the ribosomal signatures in 90,000 new environmental samples shows that no "gray" area exists: a ribosome is either bacterial or archaeal in nature.

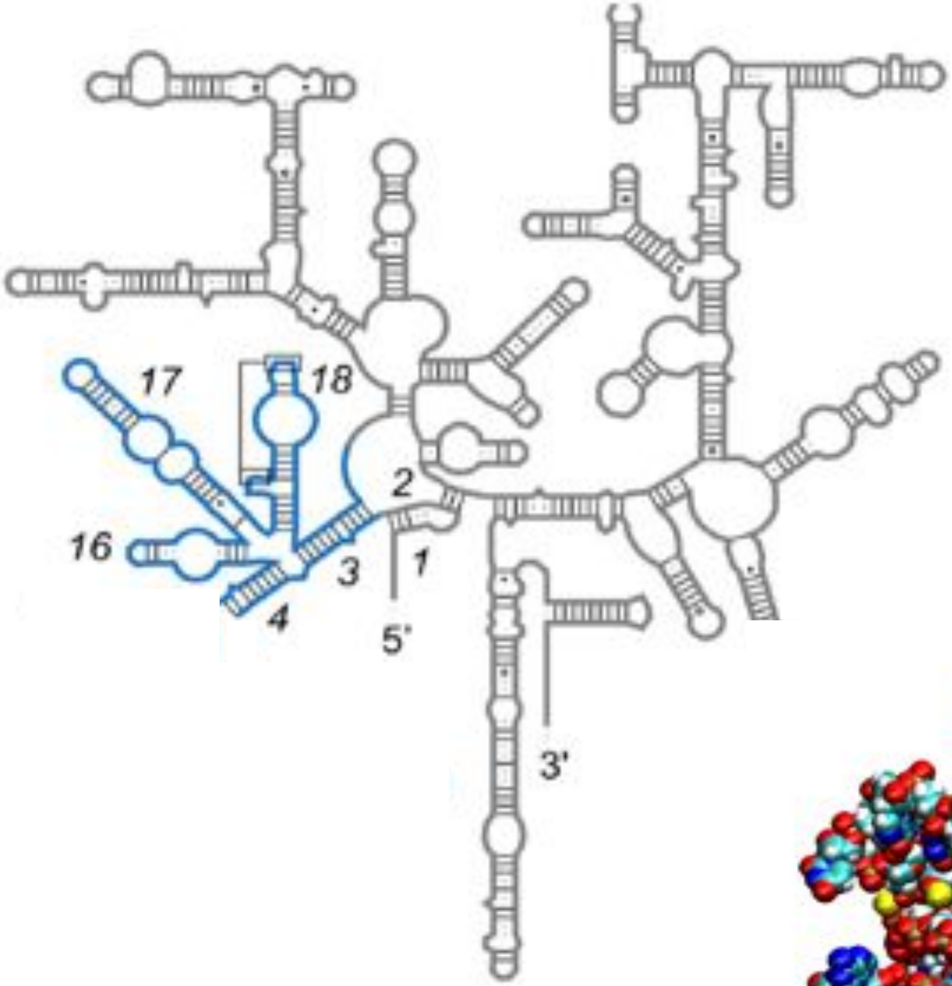
- Split across cluster: 10,000 sequence on each node

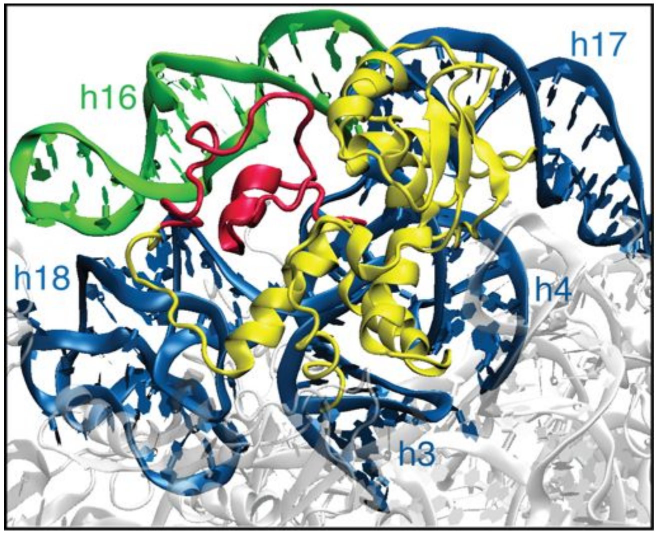


Data: "Greengenes", Lawrence Berkeley

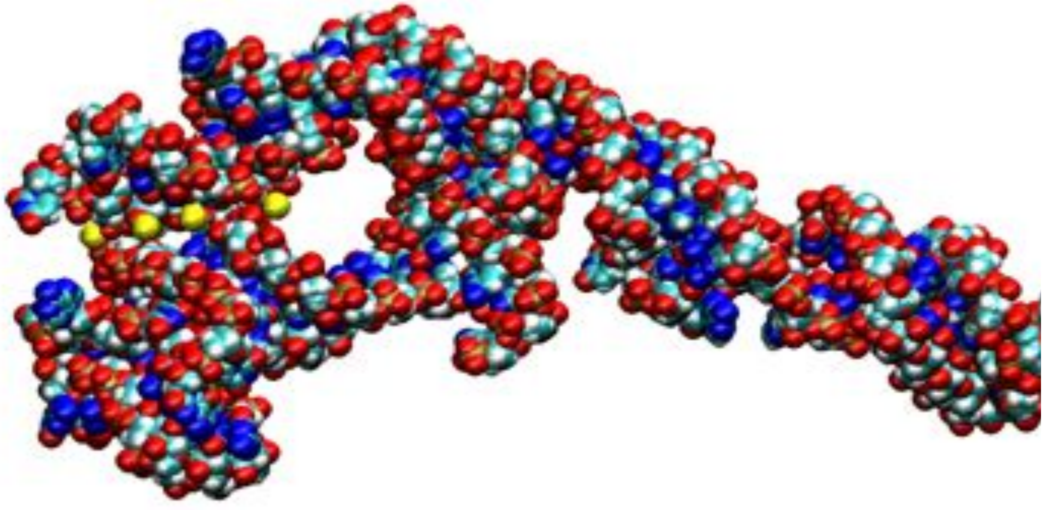
Roberts, et al. *PNAS* 2008

Largest signature region in the SSU

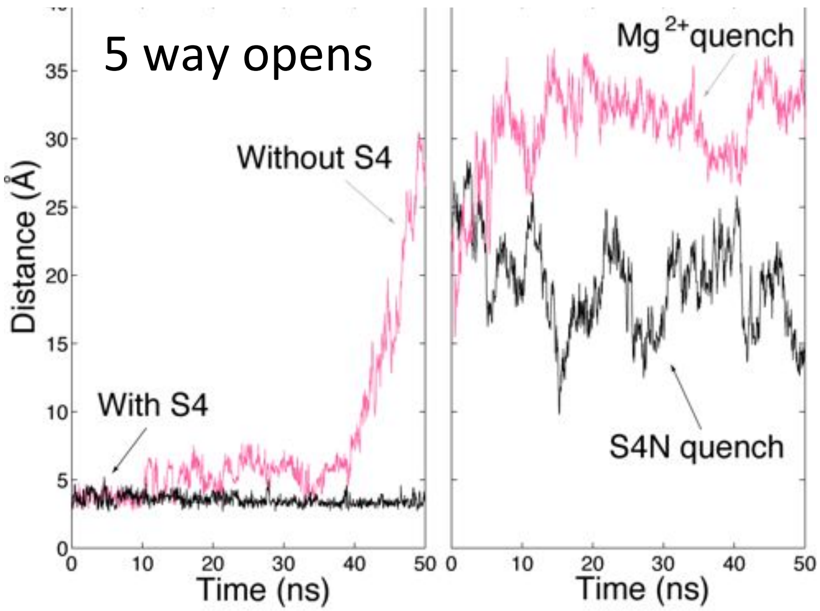


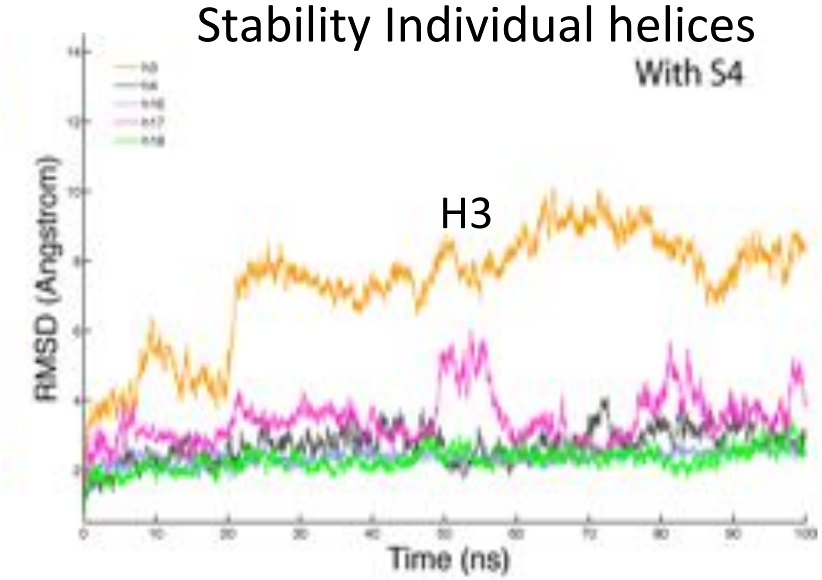


5-way helical junction rRNA with S4

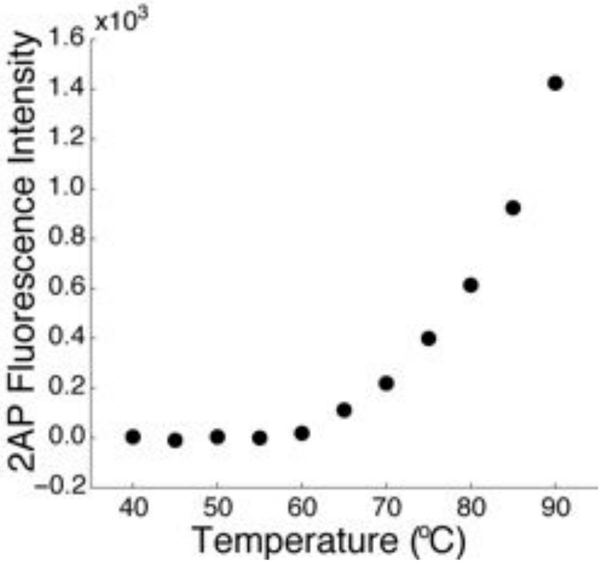


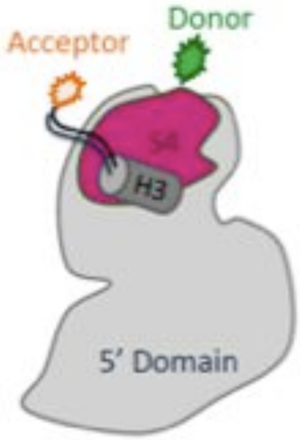
Stability of 5-way junction helices with/without S4




Stability Individual helices

h16 scaled fluorescence



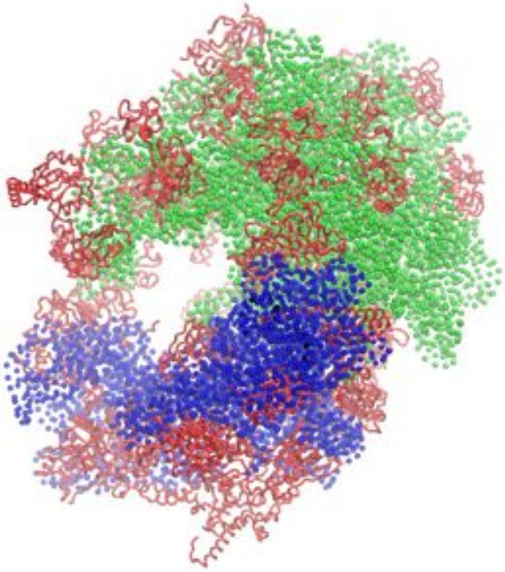


Model for Assembly ?

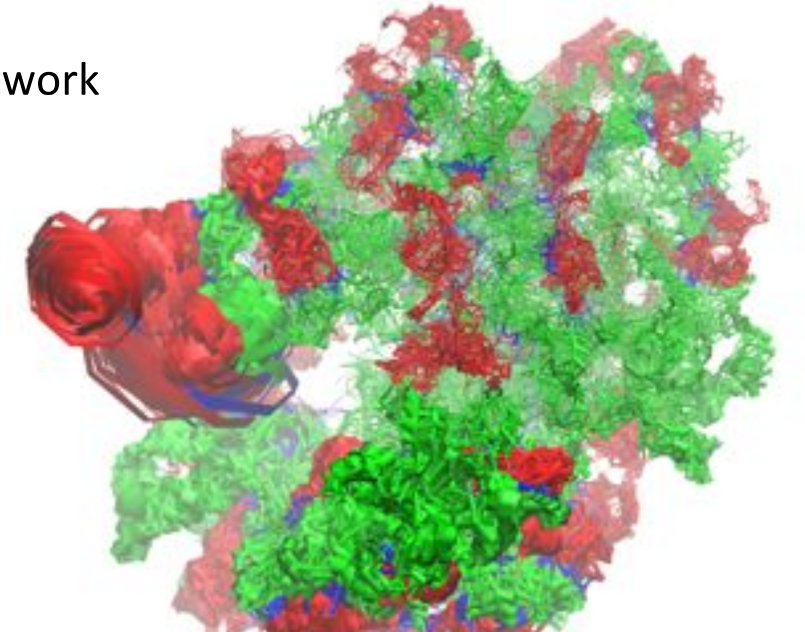
Center for the Physics of Living Cells

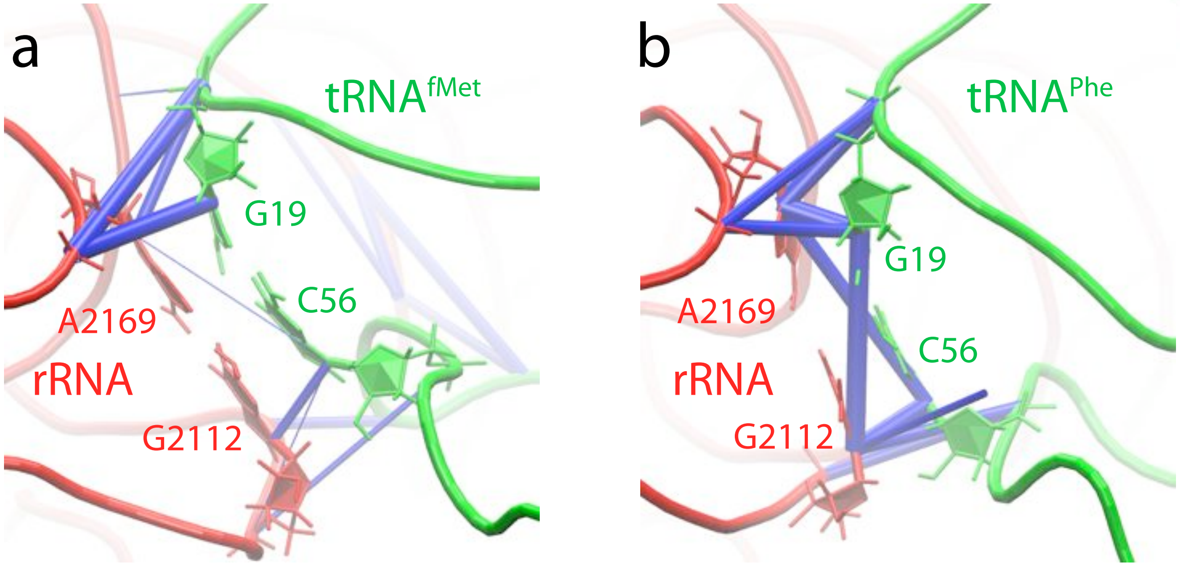
Collaboration with Gruebele, Ha, Woodson labs

Signaling Networks in LSU &SSU



MD 140ns network





P/E site tRNA:rRNA network

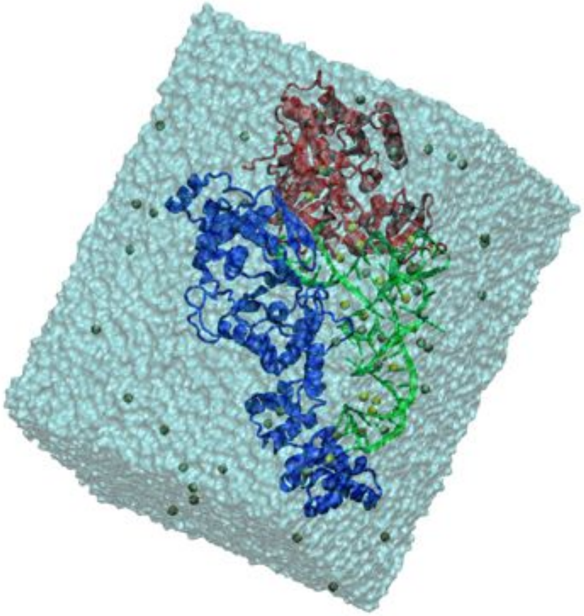
L. Trabuco, E. Schreiner, J. Eargle et al. JMB 2010

VMD/MultiSeq Tutorials

1. Evolution of Translation: AARS:tRNA
2. Evolution of Translation: EF-Tu:tRNA
3. Evolution of Translation: Ribosome
4. Dynamical Network Analysis - new

Molecular Dynamics Simulations

MD performed with NAMD2 (1) - System Setup



Simulation Parameters

Minimization: 290,000 steps
Production run: 108 ns
Forcefields: CHARMM27 (2), AMBER (3)
Time step: 1 fs
VdW frequency: 2 fs
VdW cutoff: 12 Å
Switching distance: 10 Å
Pair list distance: 14 Å
Particle Mesh Ewald
Full electrostatic update: 4 fs
Ensemble: NPT
Langevin temperature: 298.15 K
Langevin pressure: 1 atm
Periodic boundary conditions

Contents of System(4)

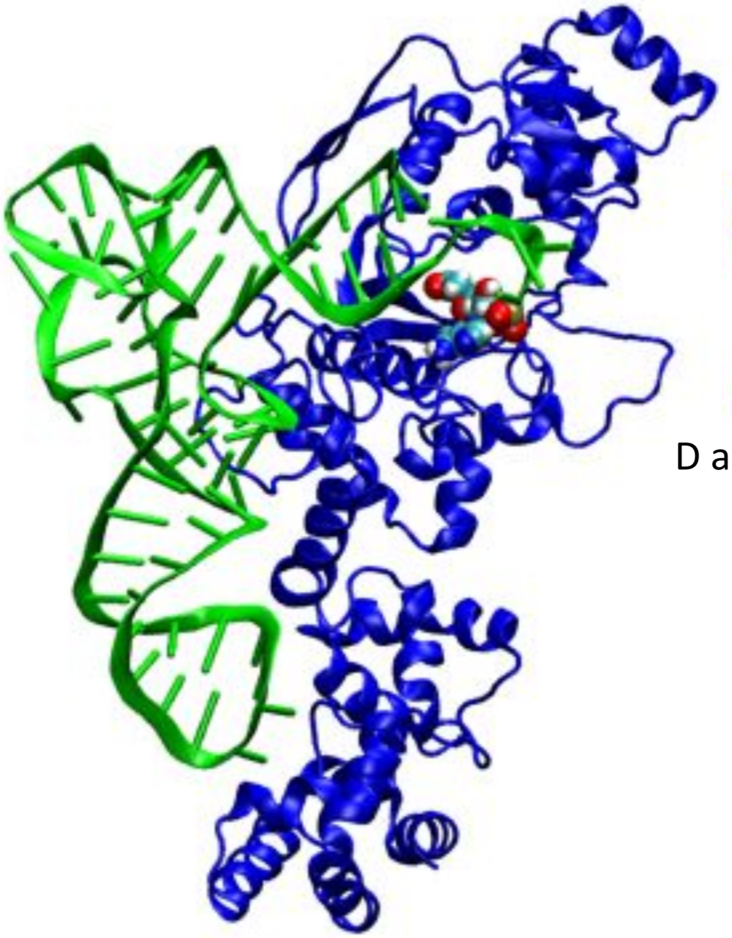
GluRS
Glu-tRNA^{Glu}
EF-Tu
GTP
Ions: Mg^{2+}, K^+
H_2O: ~27,000 molecules
System: ~130,000 atoms

System Perturbations

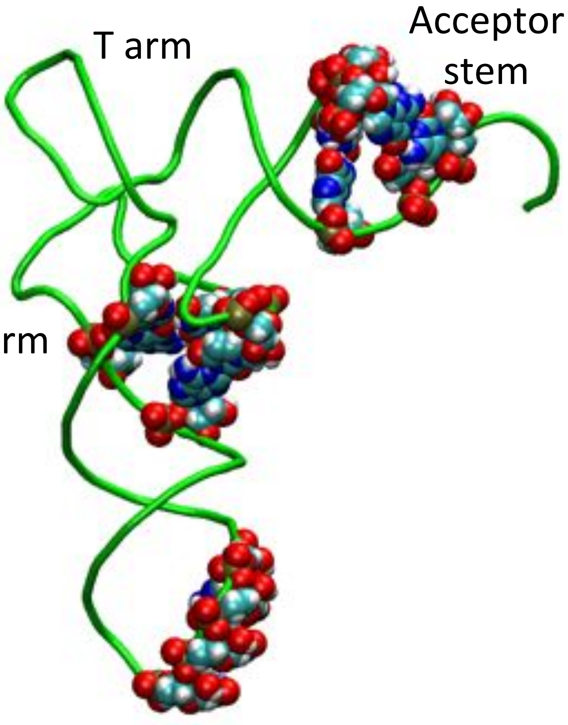
Deprotonation/protonation of reactants (aa,NTs)
Changing rotamer states of residues (5)
involved in long-lived salt bridges

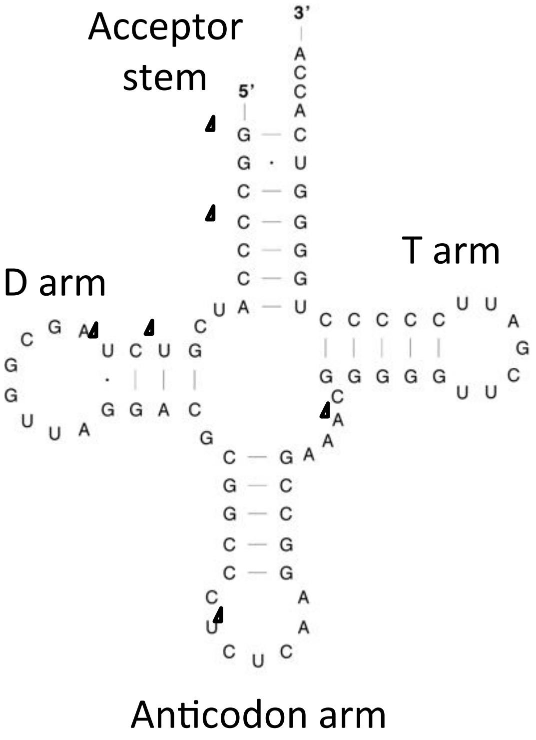
(1) Phillips, J.C. et al. *J. Comput Chem*, (2005); (2) MacKerell, A. et al. *Biopolymers* (2001);
(3) Case, D. et al. *J. Comput. Chem.* (2005); (4) Eargle, J. et al. JMB 2010, FEBS Let. 2010;
(5) Dunbrack Jr. and Cohen. *Protein Sci.* (1997)

Charging tRNA through allosteric signaling



Identity Elements

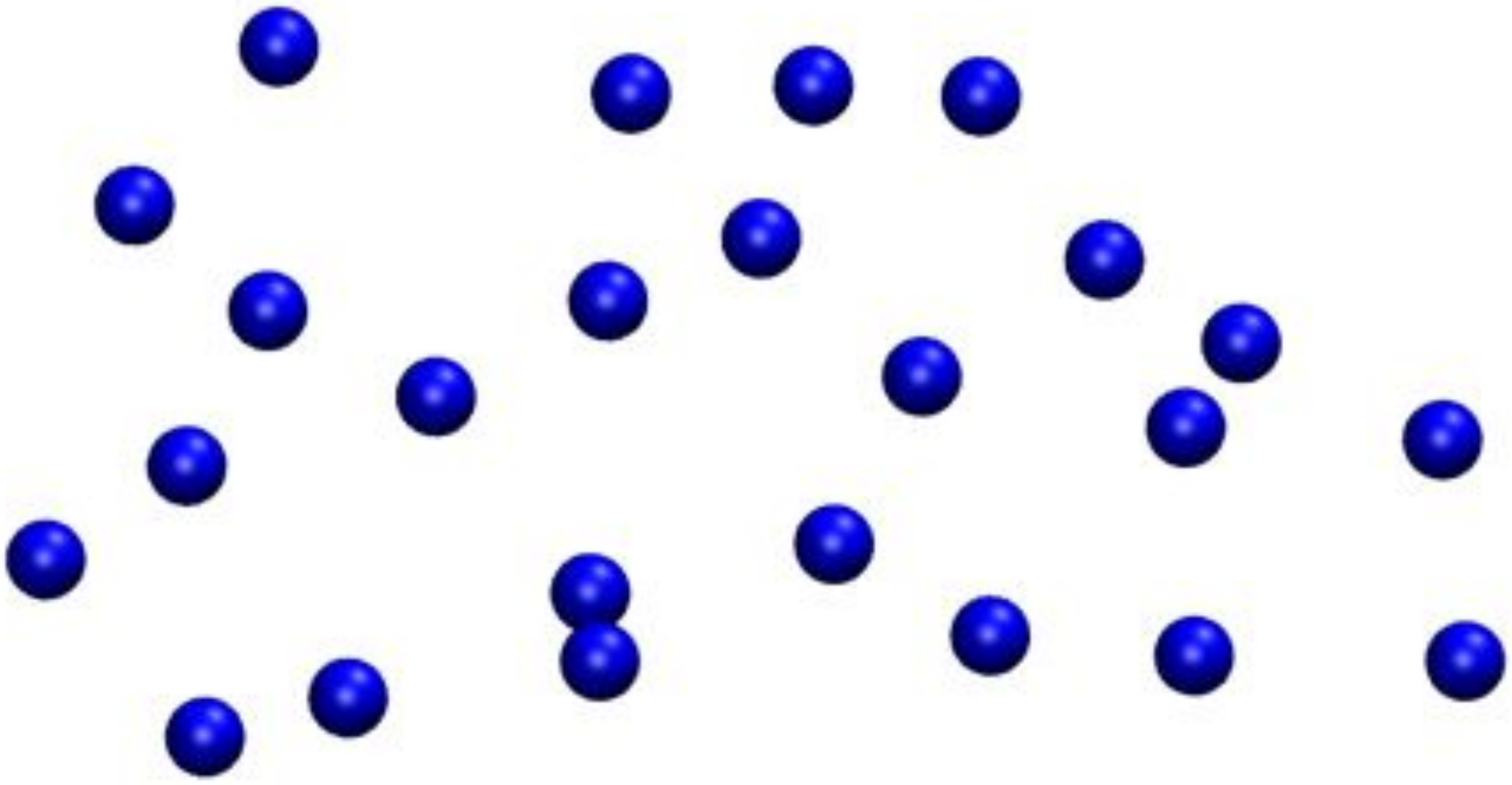




Overall Charging Reaction

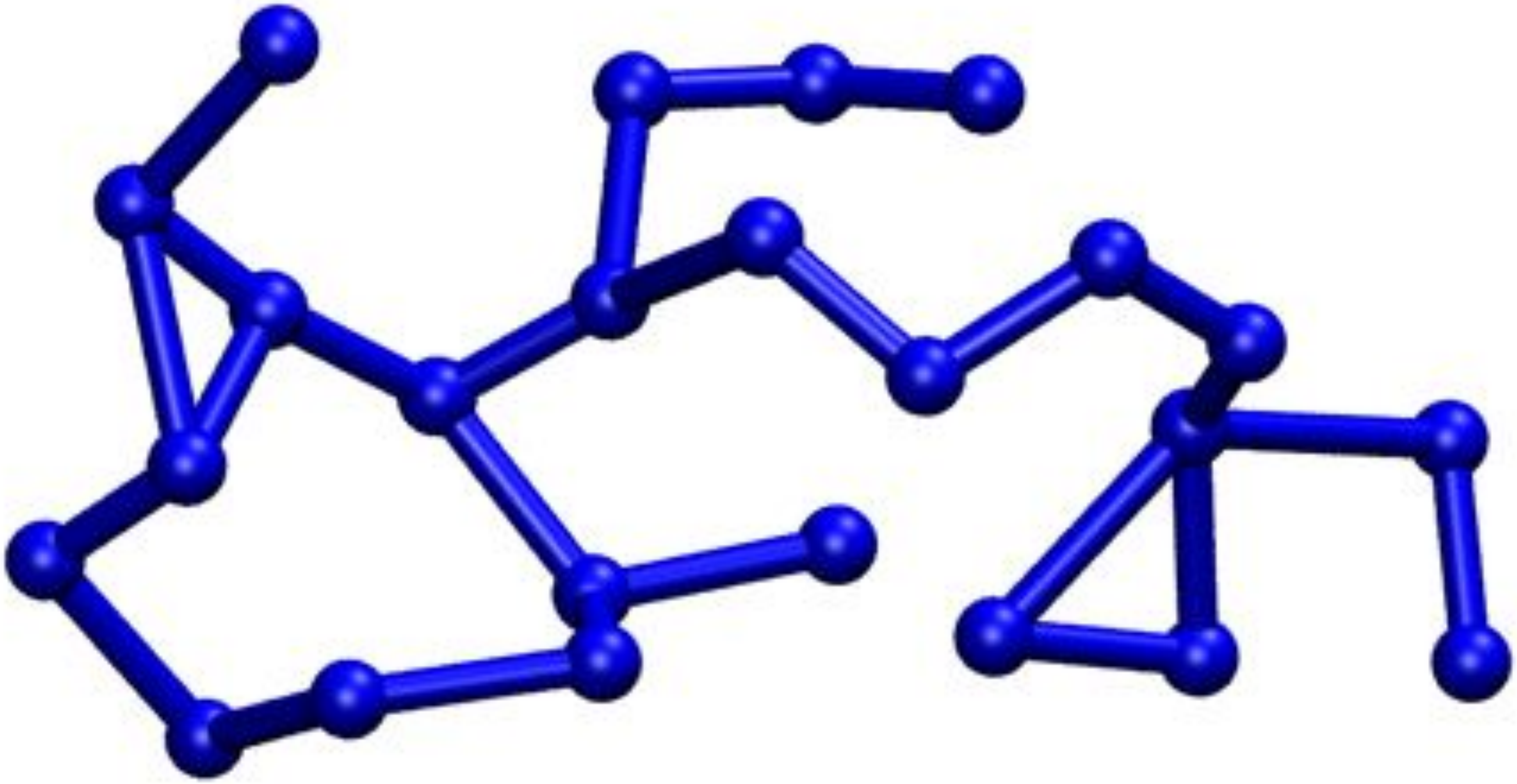
$$\text{ATP} + \text{Glu} + \text{tRNA} \xrightarrow{\text{GluRS}} \text{Glu-tRNA} + \text{AMP} + \text{PP}_i$$

How to Construct a Network?



Nodes - defined at C_a (protein) and P (nucleotide) atoms

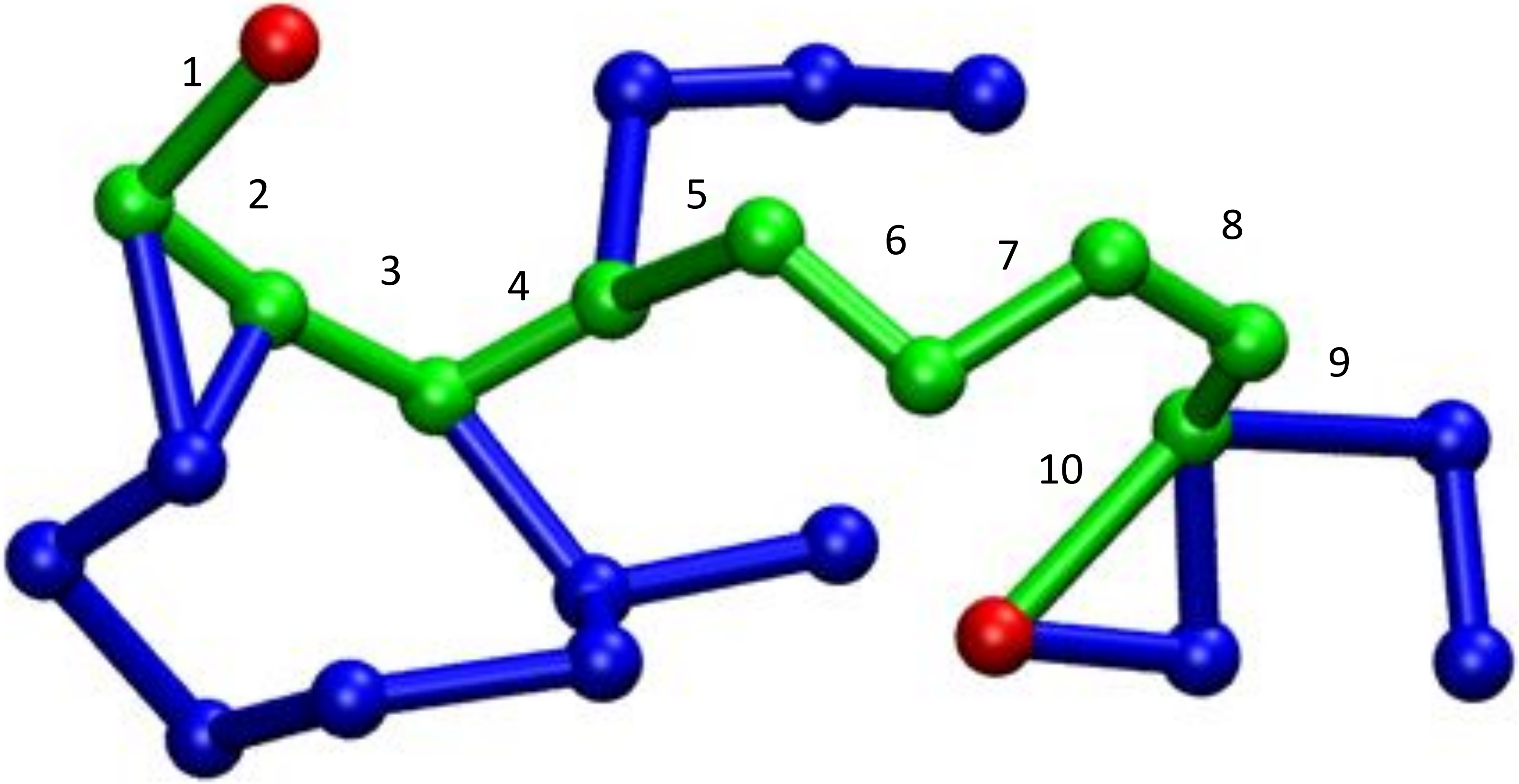
How to Construct a Network?



Edges - connect nodes that are within a contact distance threshold for more than 75% of an MD trajectory

Communication between Identity Elements and Site of Chemistry

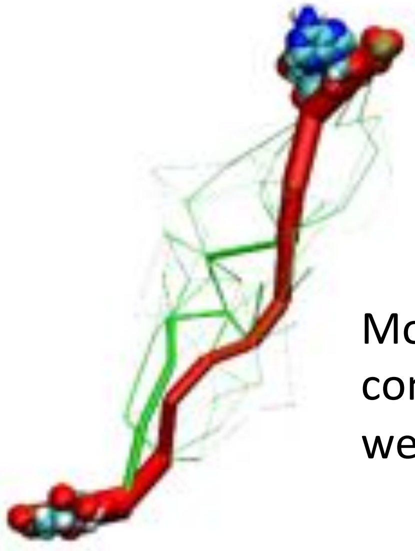
Different Paths can have Different Lengths



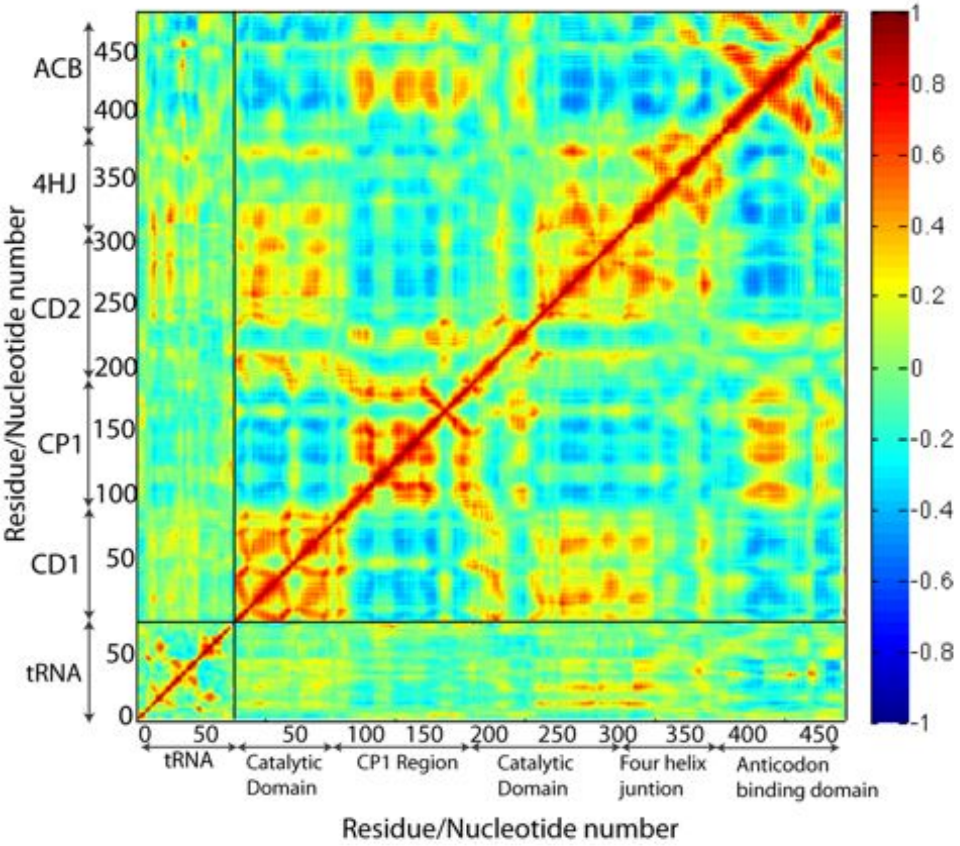
Path length = 10 (*unweighted*)

Information transfer? ***Weight contacts/links by correlations !***

Correlations (C_{ij}) define signaling pathways in GluRS:tRNA

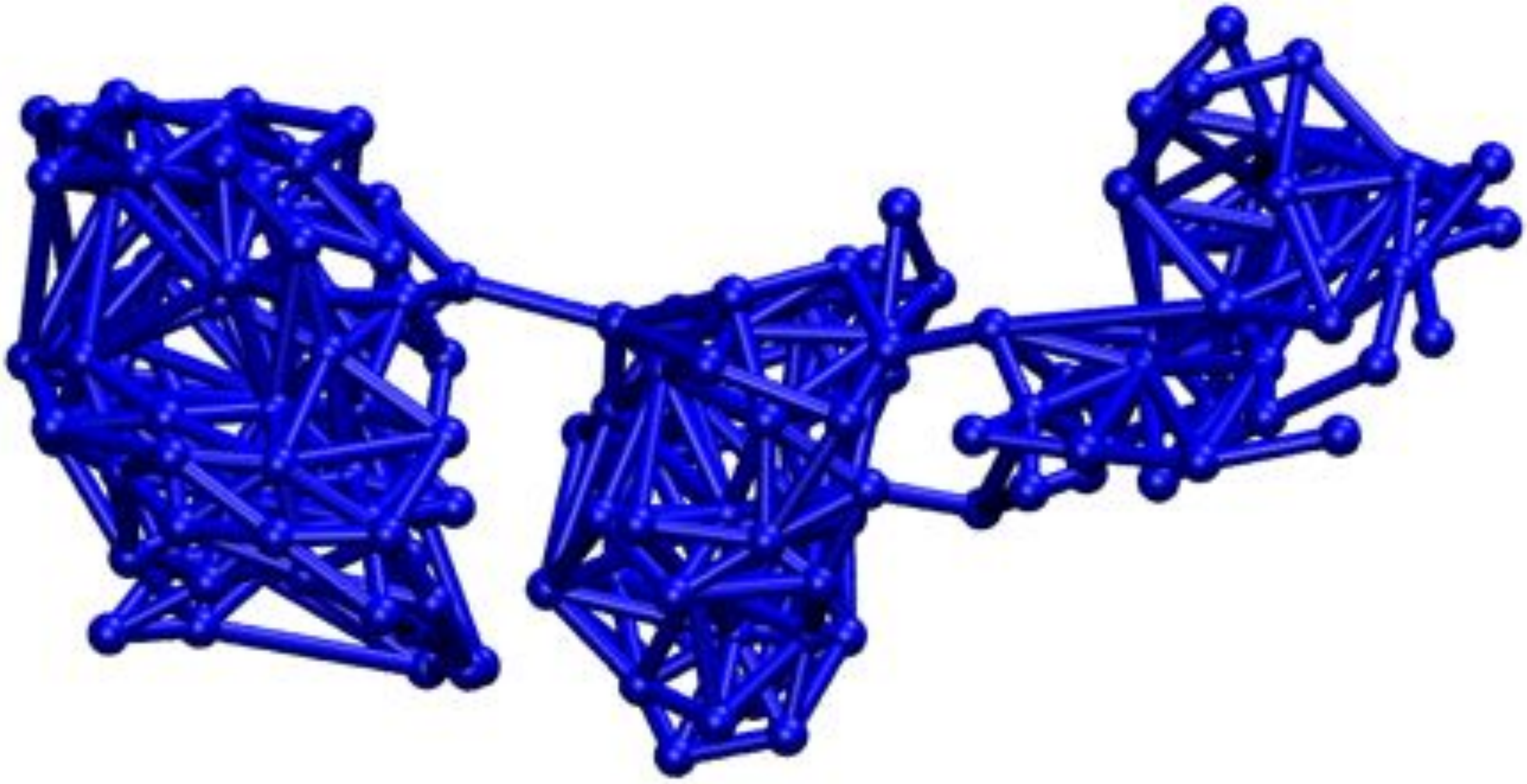


Modular stru
communitie
weighted ne

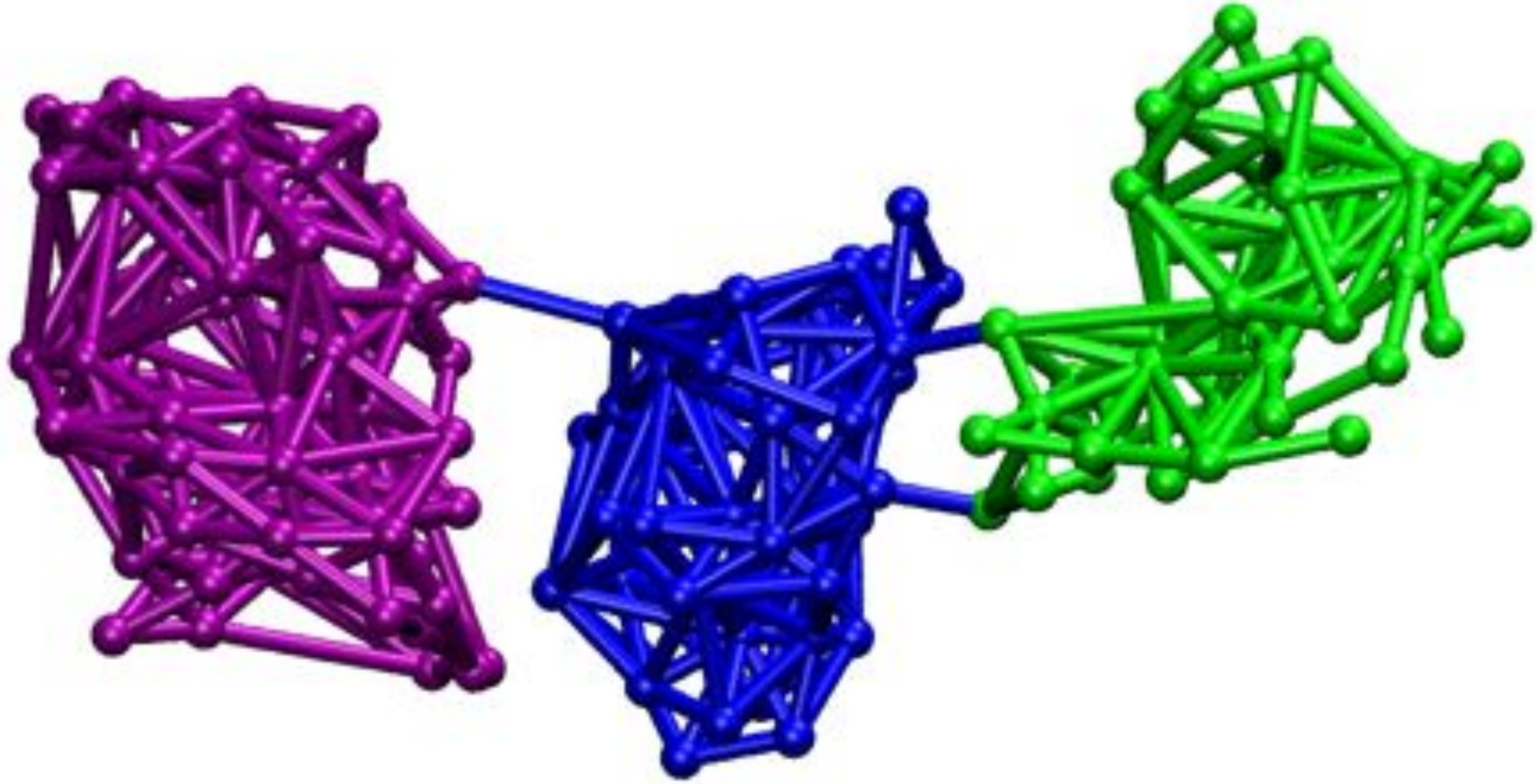


- MD simulations run with NAMD
- NTP ensemble
- Neutralized with Mg^{2+} and K^+
- C_{ij} values calculated over a 16-ns window

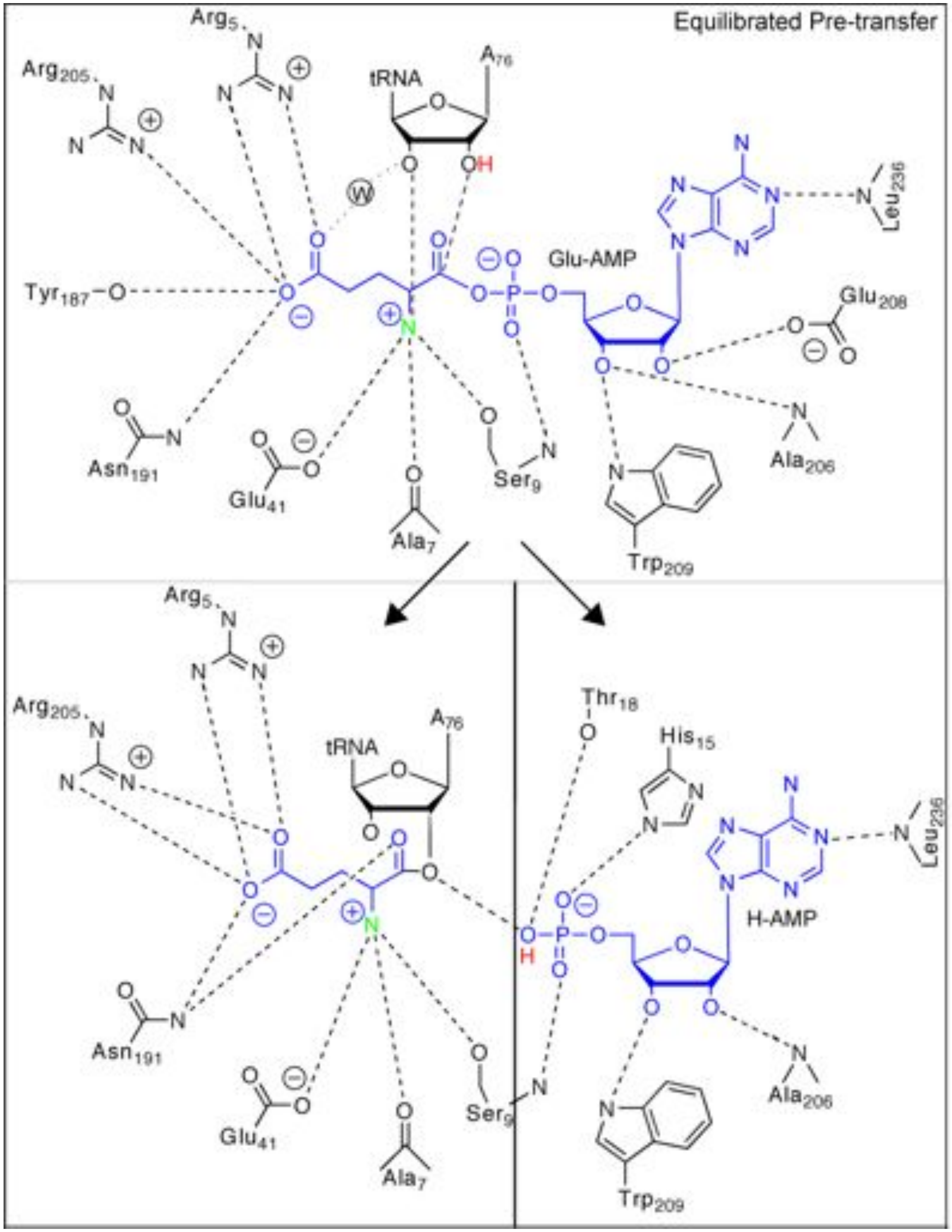
Nodes Cluster Together in Modules called Communities



Nodes Cluster Together in Modules called Communities



Communities are subnetworks with many intracommunity edges but few intercommunity edges.



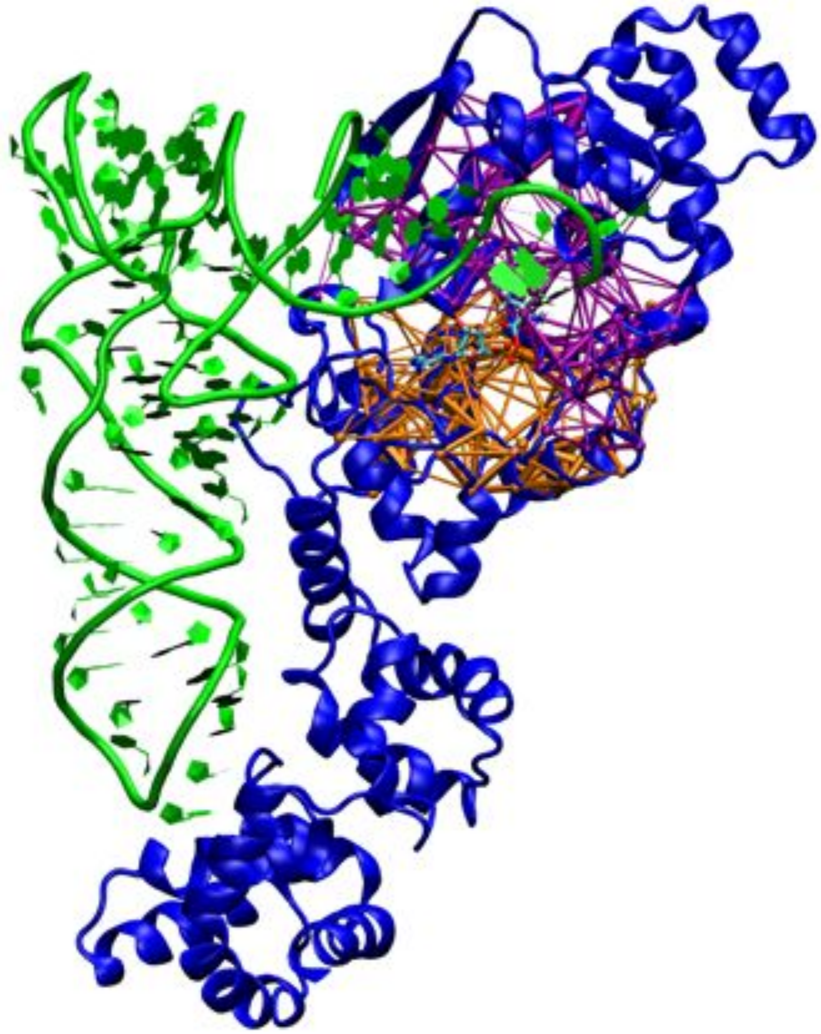
Reaction Mechanism for the Transfer of Glu to tRNA$^{\mathrm{Glu}}$

Perona JJ, Rould MA, Steitz TA
Biochemistry 1993

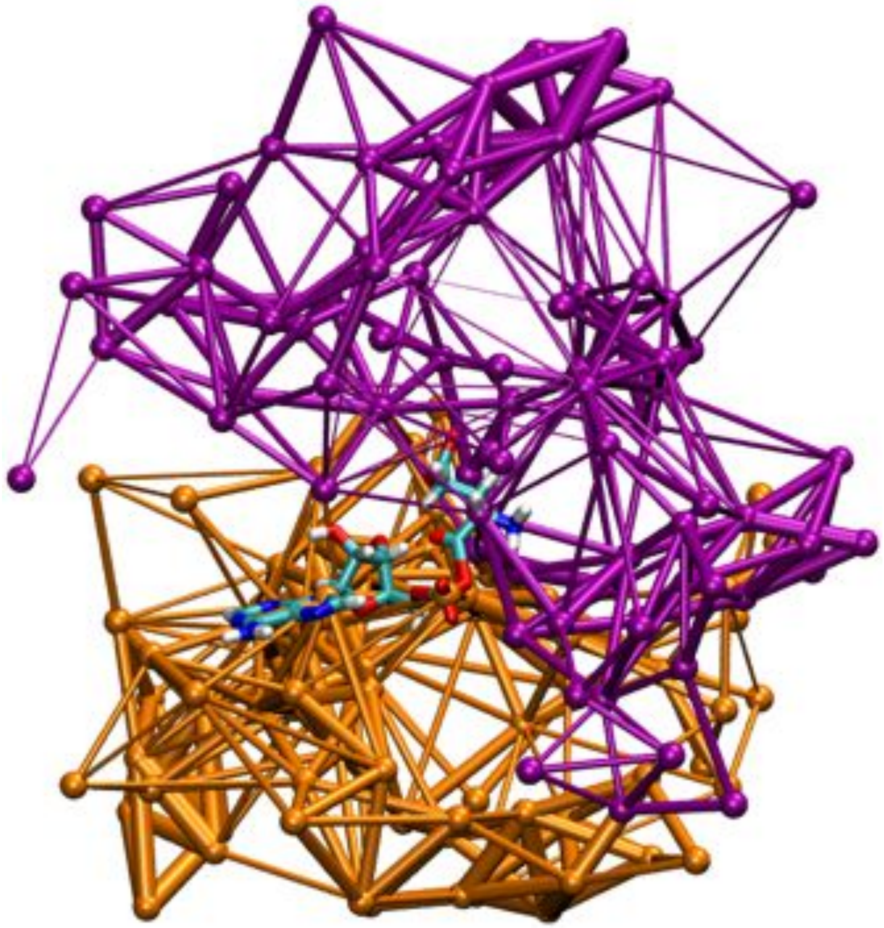
Black A, Eargle J, Sethi A,
Luthey-Schulten Z. *JMB* 2010
100s ns MD simulations

Community Structure Reveals Function in GluRS

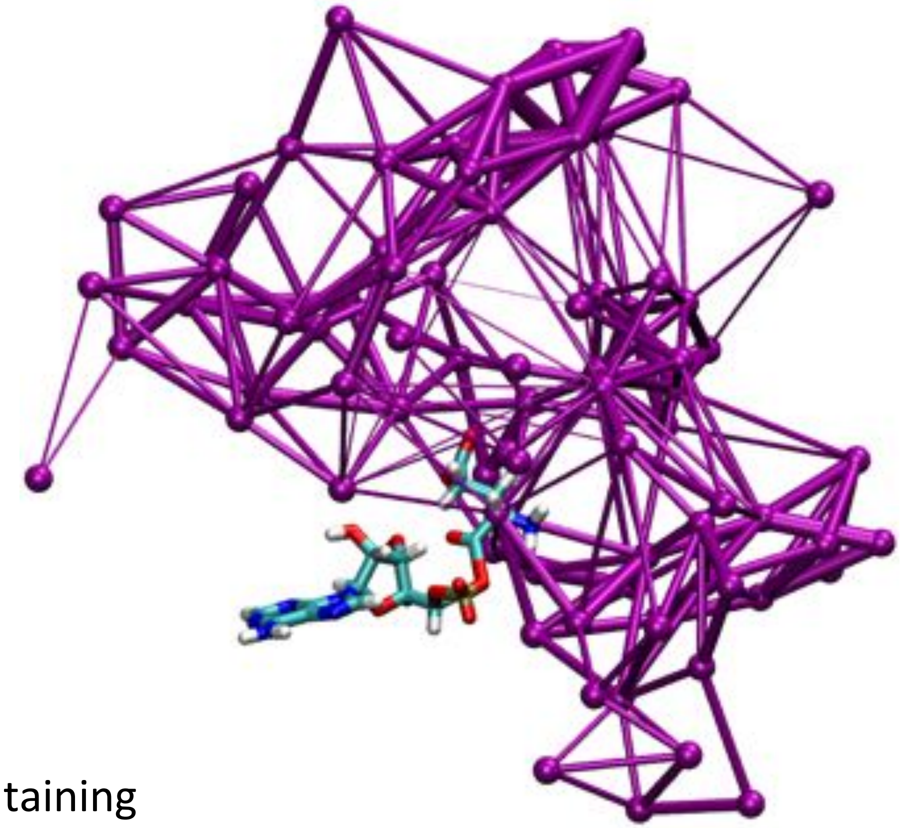
Community Structure Reveals Function in GluRS



Community Structure Reveals Function in GluRS



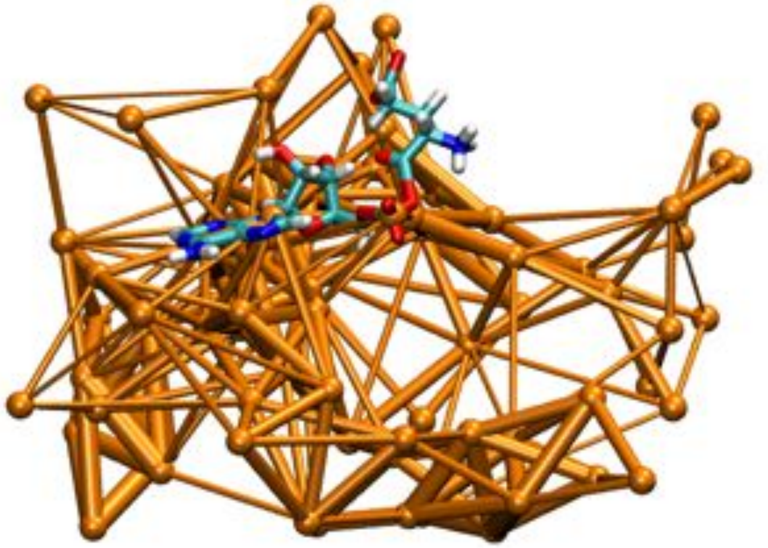
Community Structure Reveals Function in GluRS



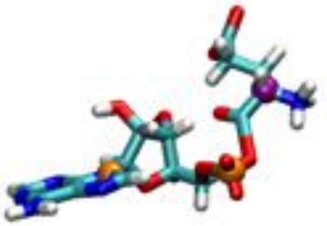
community containing
amino acid moiety

Community Structure Reveals Function in GluRS

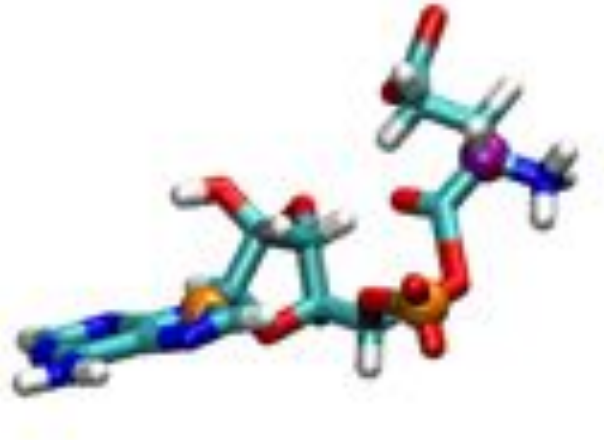
community containing
AMP moiety



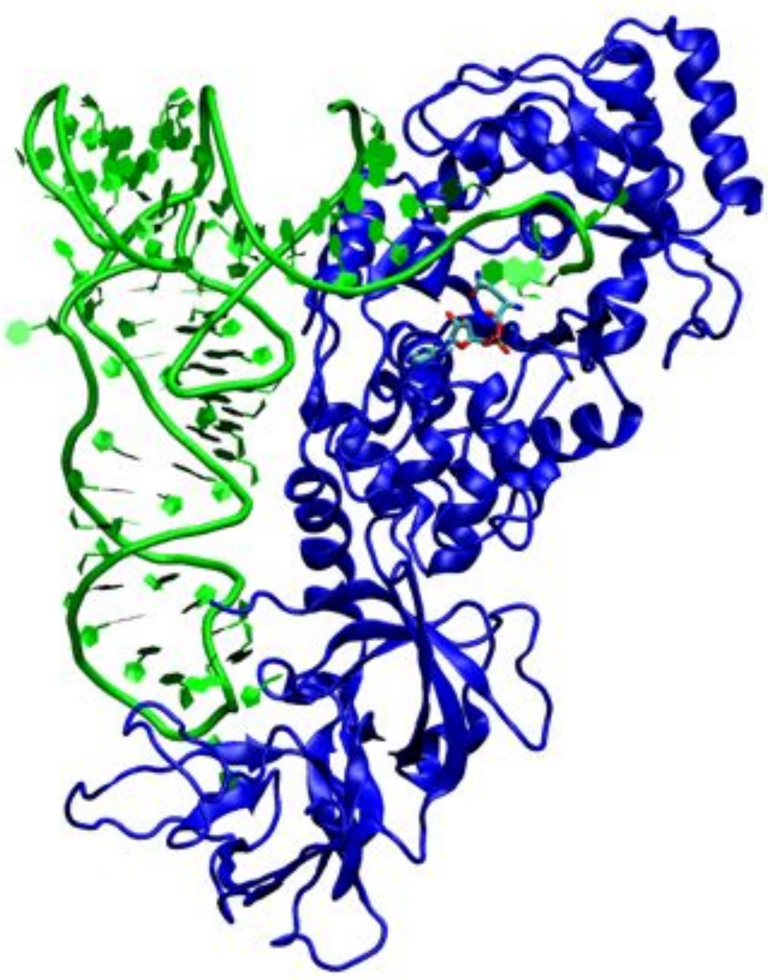
Community Structure Reveals Function in GluRS



Community Structure Reveals Function in GluRS



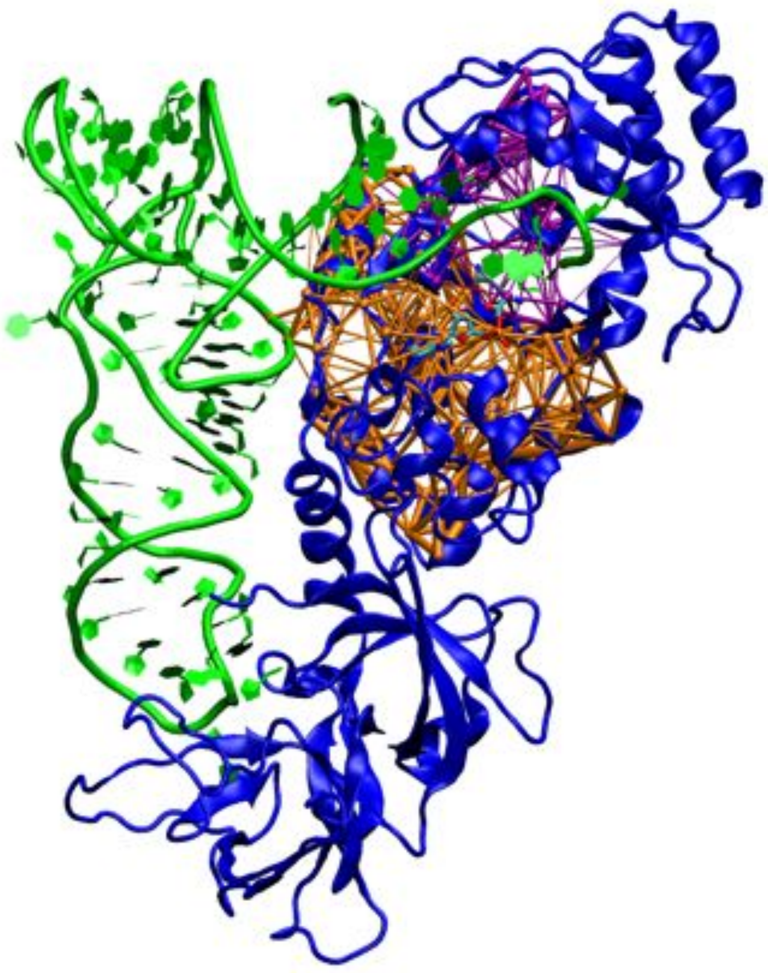
Similar Ligand Separation in GlnRS and LeuRS



GlnRS

LeuRS

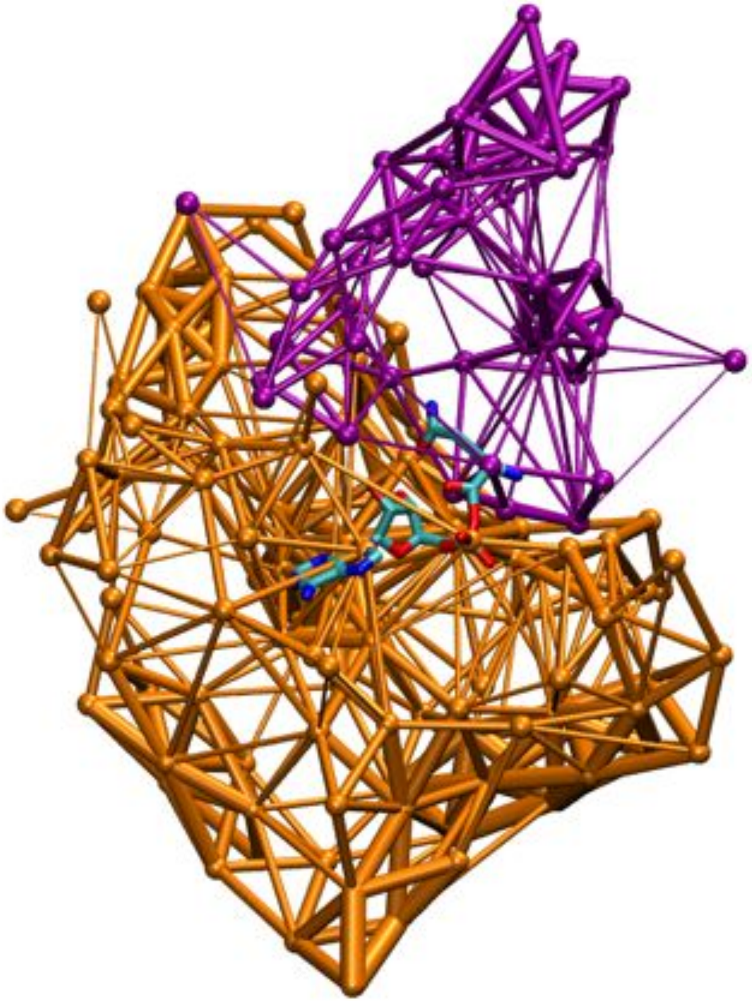
Similar Ligand Separation in GlnRS and LeuRS



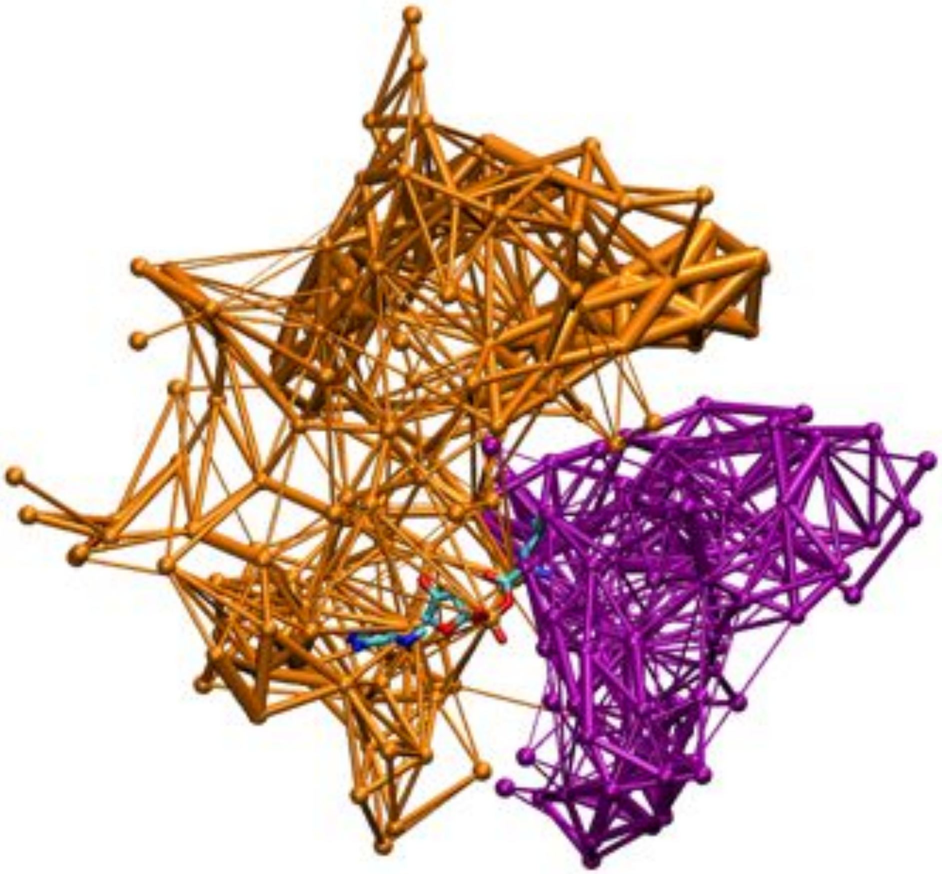
GlnRS

LeuRS

Similar Ligand Separation in GlnRS and LeuRS

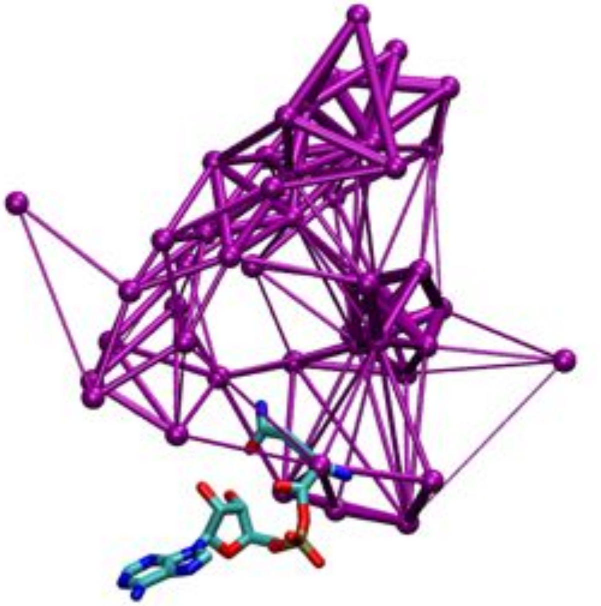


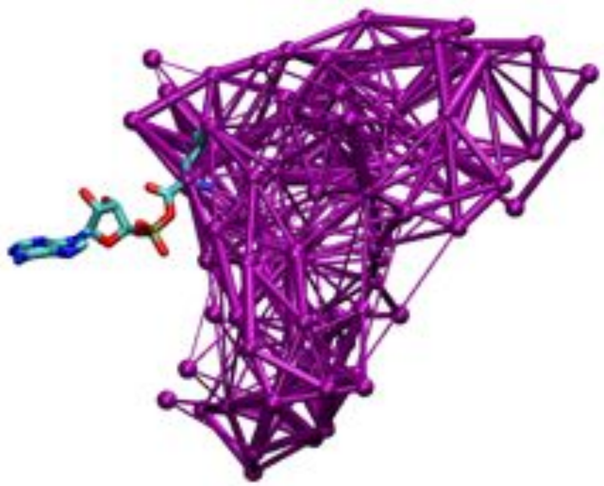
GlnRS



LeuRS

Similar Ligand Separation in GlnRS and LeuRS



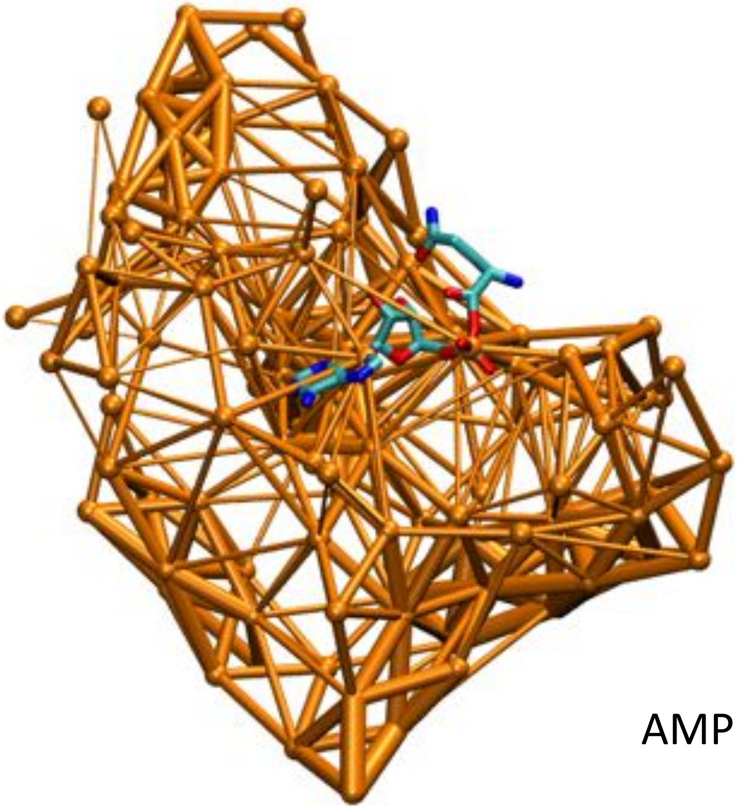


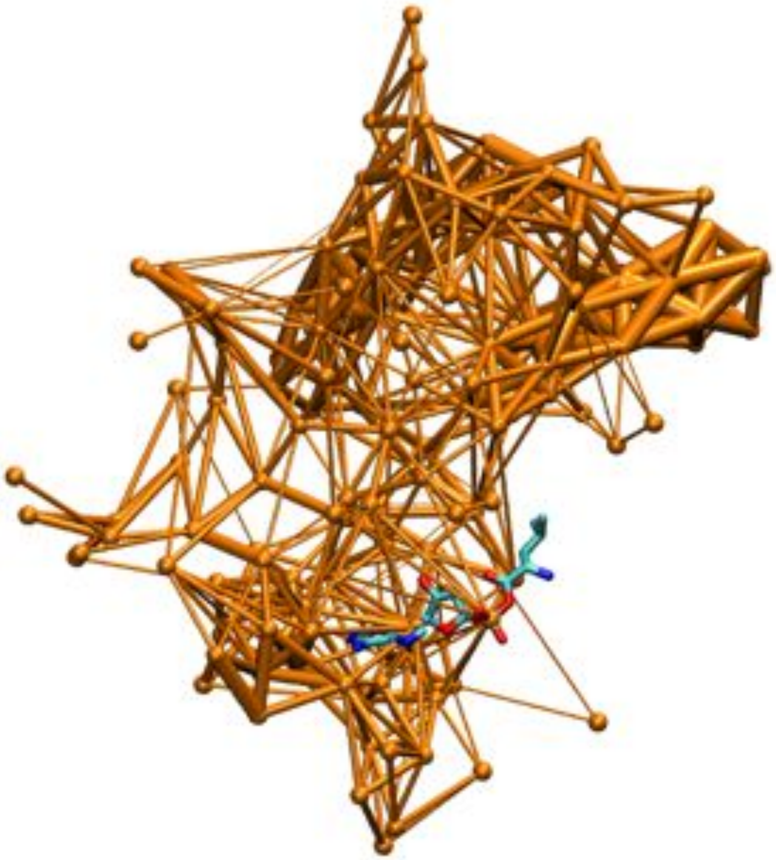
Amino acid community

GlnRS

LeuRS

Similar Ligand Separation in GlnRS and LeuRS



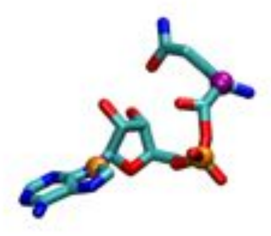


AMP community

GlnRS

LeuRS

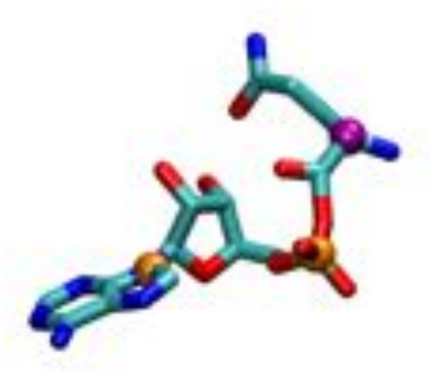
Similar Ligand Separation in GlnRS and LeuRS



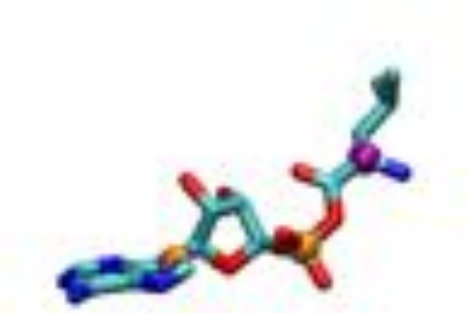
GlnRS

LeuRS

Similar Ligand Separation in GlnRS and LeuRS

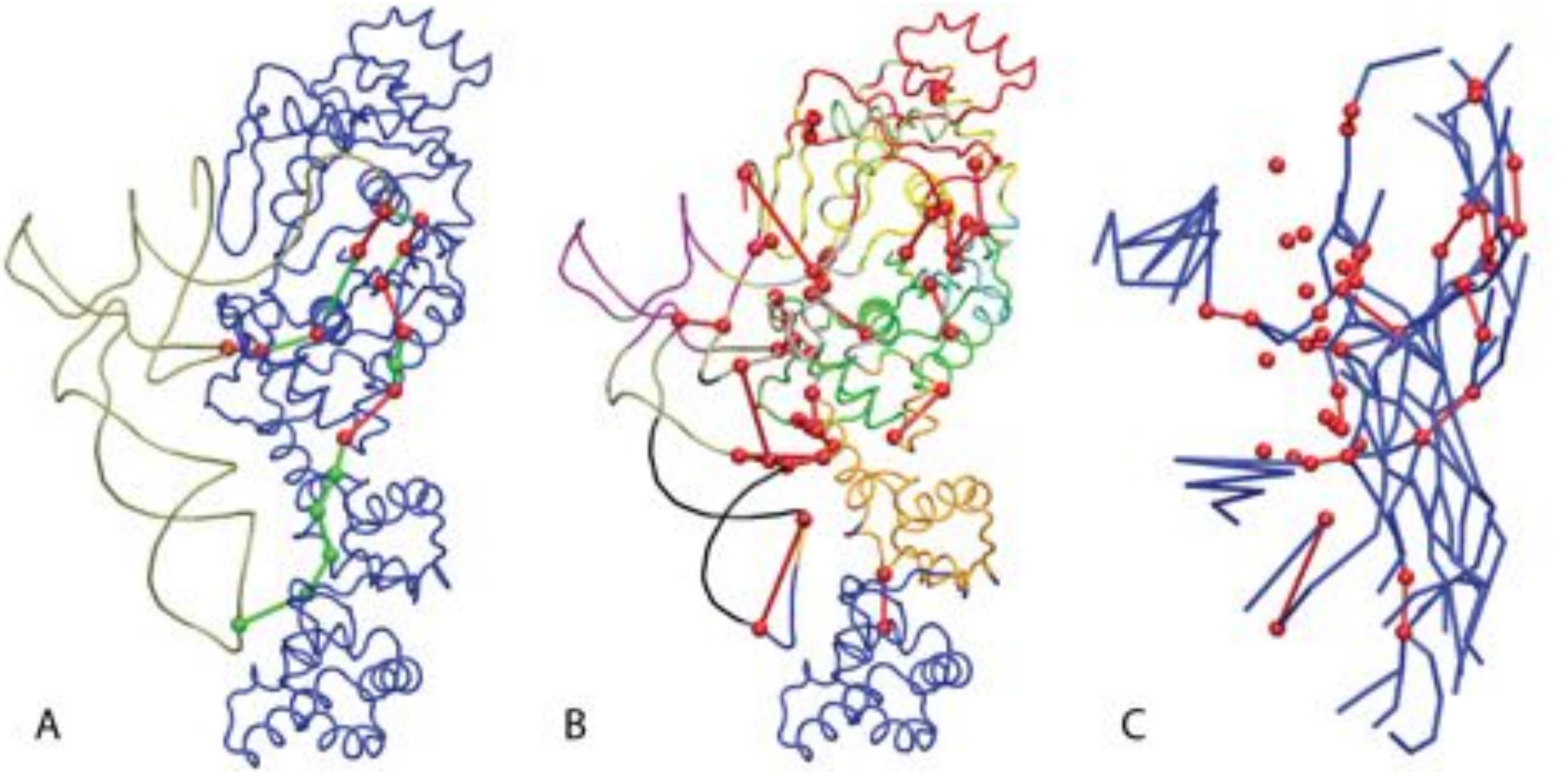


GlnRS



LeuRS

Dynamical Networks, Conservation, and Betweeness



Optimal signal pathways: U13, U35 to A76

Critical(conserved) nodes connecting communities

Betweeness routes - highest density pair optimal paths