

# **Overview & Applications**

Hands-on Workshop in Computational Biophysics

**Pittsburgh Supercomputing Center** 

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#### Full atomic simulations are computationally expensive





Bakan et al. Bioinformatics 2011.

Coarse-grained Elastic Network Models are fast

Lane et al. 2013





## **Elastic Network Model**



- Useful for predicting global motions of proteins
- Coarse-grained description (Cα-only usually)
- Residue pairs are connected via elastic springs with unified force constants
- You obtain a unique analytical solution for the spectrum of motion for each system – this is not a simulation





## Growth in structural data

#### **ONE HUNDRED THOUSAND PROTEIN STRUCTURES**

Biomolecular structures stored in the Protein Data Bank are getting bigger and more complex.



#### Multiple structures for a single sequence





## Dynamics may be inferred from structural data.





Experiment/Theory

## Leveraging the PDB since 2010

1008

- High-throughput analysis of structural data
- Application Programming Interface (API) for development of tools
- Suitable for interactive usage

User inputs a sequence

#### Usage example

>1A9U:A|PDBID|CHAIN GSSHHHHHHSSGLVPRGSHMSQ ERPTFYRQELNKTIWEVPERYQ NLSPVGSGAYGSVCAAFDTKTG

**ProDy** *identifies*, *retrieves*, *aligns*, and *analyzes* (*PCA*) structures matching input sequence

. . . . . .

Bakan, Meireles & Bahar. Bioinformatics 2011.

#### User can

Compare experimental and theoretical models

p38 ensemble

(PCA)

p38 network

model (ANM)

MD trajectory

analysis (EDA)

Sample conformations along normal modes





## An Interactive Tool







## Suite of tools











Elastic Network Model (ANM/GNM) Analysis Principal component analysis of experimentally resolved structures

Multiple Sequence Alignment Sequence conservation Correlated Mutation

Computational Drug Discovery Binding Site Prediction Affinity Estimation

A VMD plugin Visualization of collective motions Animations/movies





## Suite of tools





Modeling coupled protein-lipid dynamics Useful for membrane proteins

Response to external forces Identification of mechanical stiffness



ENM guided MD simulations Efficient sampling of energy landscape





## **Tutorials: ProDy & Structure Analysis**



- Retrieving PDB Files
- BLAST Searching the PDB
- Constructing Biomolecular Assemblies
- Determining functional motions
- Aligning and Comparing Structures
- Identifying Intermolecular Contacts





## **Tutorial: Elastic Network Models**



- Gaussian Network Model (GNM)
- Anisotropic Network Model (ANM)
- Normal Mode Analysis





## **Tutorial: Trajectory Analysis**



- Fast processing of long trajectories
- Enables comparison of MD trajectories and ENM predictions





## **Tutorial: Ensemble Analysis**



- NMR Models
- Homologous Proteins
- Multiple X-ray Structures
- Multimeric Proteins





## A better comparison:

Consider more than 2 end points for a given structure, but all the known structures for a given protein, or the structurally resolved

**Ensemble of structures** 





## What is Ensemble Analysis?

Principal component analysis

#### Input:

# An ensemble of structures for a given protein

- NMR models (~40)
- X-ray structures resolved under different conditions (ligand-bound/unbound, different stages of molecular machinery or transport cycle
- MD snapshots/frames

#### **Output:**

# Principal modes of conformational

- variations/differences between NMR models
- rearrangements/changes under different functional states
- dynamics/fluctuations observed in simulations





## What is Ensemble Analysis?

#### • Method:

- Superimpose of the structures
- Evaluate the covariance matrix (differences between individual coordinates and mean coordinates)
- Decompose it into a series of modes of covariance (3N-6 eigenvectors)

#### Principal component analysis

#### **Output:**

# Principal modes of conformational

- variations/differences between NMR models
- rearrangements/changes under different functional states
- dynamics/fluctuations observed in simulations





#### Average position vector <R<sub>i</sub>> of atom i

 $\{R_1(t_1), R_1(t_2), R_1(t_3), \dots, R_1(t_m)\}$  for atom/residue 1

Average position vector for atom *i* over all trajectory  $\langle \mathbf{R}_1 \rangle = (1/k) \sum_k \mathbf{R}_1(\mathbf{t}_k)$ , where the summation is k = 1, m

#### Instantaneous fluctuation vector

 $\Delta \mathbf{R}_{1}(t_{3}) = \mathbf{R}_{1}(t_{3}) - \langle \mathbf{R}_{1} \rangle$ 









# **RMSD** with respect to starting structure **R(O)**

Instantaneous deviation for atom i

 $\Delta \mathbf{R}_{i}(t_{k}) = \mathbf{R}_{i}(t_{k}) - \mathbf{R}_{i}(0)$ 

Average deviation over all atoms, at a given time,

**RMSD**(t<sub>k</sub>) = (1/N)  $[\sum_{i} (\Delta \mathbf{R}_{i}(t_{k}) . \Delta \mathbf{R}_{i}(t_{k}))]^{1/2}$  where i = 1, N





#### **Cross-correlations between fluctuations**

Example: cross-correlations between fluctuation vectors of residues i and j (average over *m* snapshots/conformations)

$$<\Delta \mathbf{R}_{i} \cdot \Delta \mathbf{R}_{j} > = \Sigma_{k} [\Delta \mathbf{R}_{i}(\mathbf{t}_{k}) \cdot \Delta \mathbf{R}_{j}(\mathbf{t}_{k})] / m$$

For i = j, this reduces to mean-square fluctuation

<(
$$\Delta \mathbf{R}_{\mathrm{i}})^{2}$$
>

## Covariance matrix (NxN)



 $\Delta \mathbf{R} = \mathbf{N}$ -dim vector of instantaneous fluctuations  $\Delta \mathbf{R}_i$  for all residues ( $1 \le i \le N$ )

 $< \Delta \mathbf{R}_1$ .  $\Delta \mathbf{R}_1 > =$  ms fluctuation of site 1 averaged over all *m* snapshots.





### Cross-correlations between Components of fluctuation vectors

Example: cross-correlations between the X-component of R<sub>i</sub> and Y component of R<sub>i</sub>

$$<\Delta X_i \Delta Y_j > = \Sigma_k [\Delta X_i(t_k) \Delta Y_j(t_k)] / m$$

To be organized in a 3x3 matrix as





#### Covariance matrix (3Nx3N)









$$\mathbf{C}^{(ij)} = \begin{bmatrix} \left\langle \Delta x_i \Delta x_j \right\rangle & \left\langle \Delta x_i \Delta y_j \right\rangle & \left\langle \Delta x_i \Delta z_j \right\rangle \\ \left\langle \Delta y_i \Delta x_j \right\rangle & \left\langle \Delta y_i \Delta y_j \right\rangle & \left\langle \Delta y_i \Delta z_j \right\rangle \end{bmatrix} \longrightarrow \begin{bmatrix} \mathbf{C} = \mathbf{P} \mathbf{S} \mathbf{P}^T = \sum_{i=1}^{3N} \sigma_i \ \mathbf{p}^i \mathbf{p}^T \end{bmatrix}$$





# Global motions inferred from theory and experiments



 $\rightarrow$  PCA of the ensemble of resolved structures  $\rightarrow$  ANM analysis of a single structure from the ensemble





# Global motions inferred from theory and experiments



The intrinsic dynamics of enzymes plays a dominant role in determining the structural changes induced upon inhibitor binding

Ahmet Bakan and Ivet Bahar<sup>1</sup>

Department of Computational Biology, School of Medicine, University of Pittsburgh, 3064 BST3, 3501 Fifth Avenue, Pittsburgh, PA 15213

#### Reference:

Bakan & Bahar (2009) PNAS 106, 14349-54





### Soft modes enable functional movements



#### References:

Bakan & Bahar (2009) PNAS 106, 14349-54.





## **Comparing PCA and ENM**

Structures of HIV-1 RT Unbound Inhibitor bound DNA bound









## **Example: Comparing PCA and ENM**

Structures of p38 MAPK Unbound Inhibitor bound Glucose bound Peptide bound









20

## Different types of spring 'constants'



Yang et al. PNAS 106 (2009).





## **Tutorial: Normal Mode Wizard**



- perform ANM, GNM, and PCA/EDA calculations
- draw customizable normal mode arrows
- make animations (sample conformations)
- make interactive squarefluctuations plots
- compare two structures and draw deformation arrows





## **Tutorial: Evol**



- identification of conserved and coevolving residues
- Retrieving multiple sequence alignments (MSAs) from Pfam DB
- extremely fast MSA I/O functions
- Generation of conservation profiles (1D plots) and coevolution maps (2D plots)





### Tutorial: collective Molecular Dynamics



- Sampling the conformational space near native state
- Identification of substates and accessible transitions
- Generating transition paths between substates
- Obtaining information on global dynamics at atomic resolution
- Generating the conformational energy landscape for the investigated system





## Tutorial: membrANM



- Evaluating membrane proteins' dynamics in the presence of lipid bilayer, also modeled as an elastic network model, explicit or implicit
- Comparing protein global motions in the presence and absence of membrane
- Understanding mechanisms of protein-membrane remodeling or coupling to facilitate function





## **Global transitions**







## **Global transitions**

Single subunit showing the transport domain moving across the membrane







## **Global transitions**

Single subunit showing the transport domain moving across the membrane







## Tutorial: MechStiff



- Identification of the anisotropic response of the structure to external perturbations
- determination of the weak/strong pairs of interactions depending on the direction of the external force and the sites that are subjected to perturbation (uniaxial tension)
- Determination of the effective spring constant observed macroscopically, to be compared with data from Molecule Force Spectroscopy (SMFS) or atomic force microscopy (AFM).
- Evaluating the contributions of each mode to deformations along selected directions





## **Tutorial: Druggability**



- Set up NAMD simulations
- Analyze trajectories to identify binding hot spots





### Exploring binding with probe molecules







## A few commands in ProDy

- Download NMR structures from PDB
- Calculate residue MSFs for each protein
- Determine ENM topology







## Fine-tuning force constants



Learn more at prody.csb.pitt.edu





## **Rotations-Translations of Blocks**



Smaller Hessian can be more easily diagonalized...



...and modes projected back into all-residue space



H: ANM Hessian (3 rows/cols per residue) P: Projection matrix from all-residue space to rigid block space H<sup>RTB</sup>: RTB Hessian (no internal motions of blocks) V'<sup>AA</sup>: Approximate ANM motions RTB.buildHessian()





# Exploring structural transitions: Glutamate transporter

ANM predicts large radial motions of the trimer. Can we design a better model?

$$\mathbf{H}_{ij} = -\frac{\gamma}{\left(R_{ij}^{0}\right)^{2}} \begin{bmatrix} \left(x_{ij}^{0}\right)^{2} & x_{ij}^{0}y_{ij}^{0} & x_{ij}^{0}z_{ij}^{0} \\ x_{ij}^{0}y_{ij}^{0} & \left(y_{ij}^{0}\right)^{2} & y_{ij}^{0}z_{ij}^{0} \\ x_{ij}^{0}z_{ij}^{0} & y_{ij}^{0}z_{ij}^{0} & \left(z_{ij}^{0}\right)^{2} \end{bmatrix}$$

Altered radial force constants:

$$\begin{split} \mathbf{H_{ij}} &= - \left( R_{ij}^{0} \right)^{-2} \begin{bmatrix} \left( x_{ij}^{0} \sqrt{\gamma_{x}} \right)^{2} & x_{ij}^{0} y_{ij}^{0} \sqrt{\gamma_{x} \gamma_{y}} & x_{ij}^{0} z_{ij}^{0} \sqrt{\gamma_{x} \gamma_{y}} \\ x_{ij}^{0} y_{ij}^{0} \sqrt{\gamma_{x} \gamma_{y}} & \left( y_{ij}^{0} \sqrt{\gamma_{y}} \right)^{2} & y_{ij}^{0} z_{ij}^{0} \sqrt{\gamma_{y} \gamma_{y}} \\ x_{ij}^{0} z_{ij}^{0} \sqrt{\gamma_{x} \gamma_{z}} & y_{ij}^{0} z_{ij}^{0} \sqrt{\gamma_{y} \gamma_{z}} & \left( z_{ij}^{0} \sqrt{\gamma_{z}} \right)^{2} \\ \end{bmatrix} \\ \mathbf{H_{ij}} &= - \frac{\gamma}{\left( R_{ij}^{0} \right)^{2}} \begin{bmatrix} \left( x_{ij}^{0} \right)^{2} & x_{ij}^{0} y_{ij}^{0} & c x_{ij}^{0} z_{ij}^{0} \\ & x_{ij}^{0} y_{ij}^{0} & \left( y_{ij}^{0} \right)^{2} & c y_{ij}^{0} z_{ij}^{0} \\ & c x_{ij}^{0} z_{ij}^{0} & c y_{ij}^{0} z_{ij}^{0} & \left( c z_{ij}^{0} \right)^{2} \end{bmatrix} \end{split}$$



Lezon & Bahar. Biophys J 102 (2012).





## Exploring structural transitions: Glutamate transporter





#### **ANM:** Large radial motions

imANM





## System/environment approximation



As the *environment* fluctuates randomly, the effective motion of the *system* is given by

$$V_{eff}(\mathbf{s}) = \frac{1}{2} \Delta \mathbf{s}^{T} (\mathbf{H}^{ss}') \Delta \mathbf{s}$$
$$\mathbf{H}^{ss}' = \mathbf{H}^{ss} - \mathbf{H}^{SE} (\mathbf{H}^{EE})^{-1} \mathbf{H}^{ES}$$
reduceModel()