

SUMMARY

- Structure-encoded **flexibility** of drug targets and significance in drug discovery and design
- Druggability assessment: a first step before selecting a target
- Modularity and promiscuity of proteins and quantitative systems pharmacology methods

Druggable Genome

A small subset of are 'disease-modifying' – and not all of them are druggable



Druggable genome 3,000 genes

430+ kinases

600+ GPCRs

70+ kinases 100+ GPCRs Drug

Drug
Targets

600-1,500

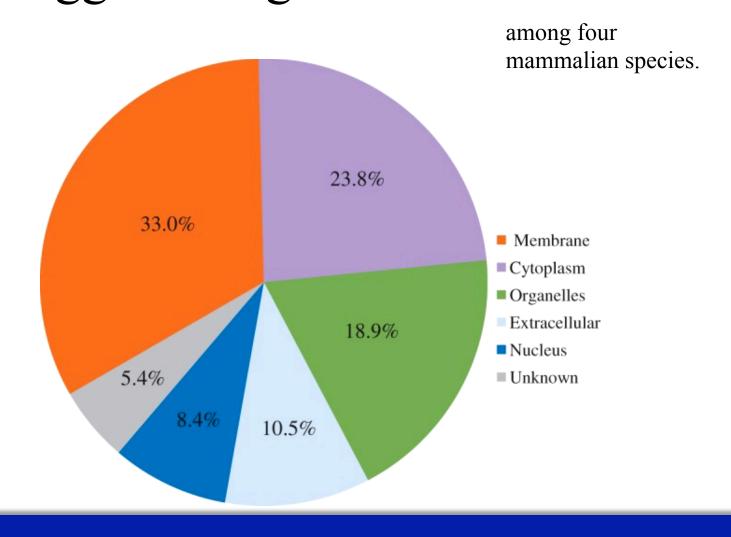
Disease-related genes ~3,000 genes

Hopkins and Groom, Nat Reviews Drug Disc, 2002

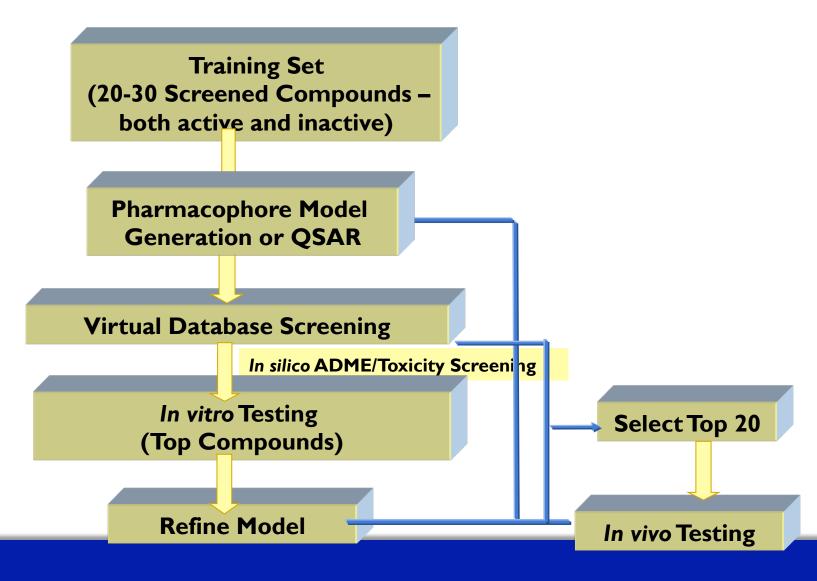


- Only 2% of human proteins interact with currently approved drugs.
- 10-15% of human proteins are disease-modifying
- 10-15% are druggable
- 5% are both disease-modifying and druggable

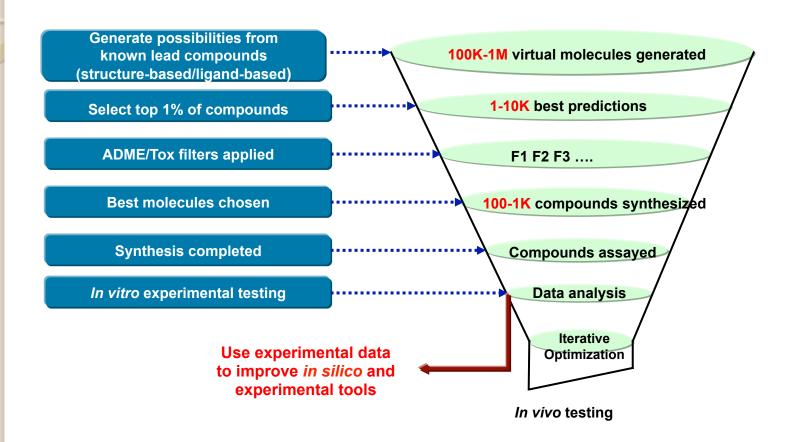
Subcellular distribution of 1,362 druggable targets



Ligand-Based Strategy



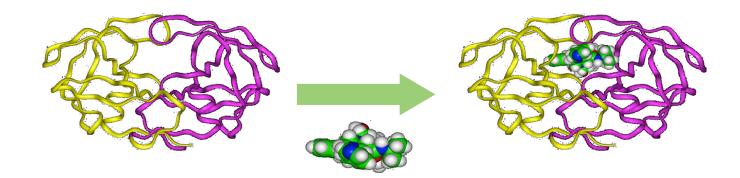
Screening Cascade



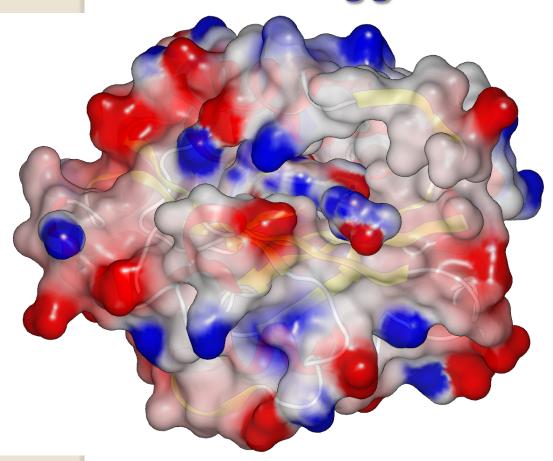
Rational Design of Inhibitors

3D structure of the target is used for

- Visual inspection/molecular graphics
- Docking (of both small molecules or fragments thereof)
- De novo methods
- Receptor properly mapping + database searching



Druggable or not?

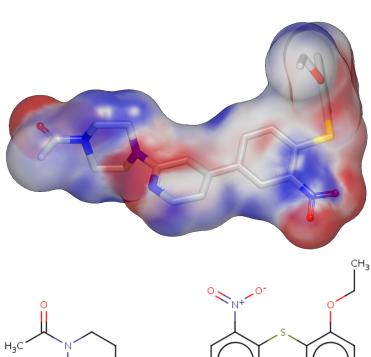


Lfa1 - a leukocyte glycoprotein that promotes intercellular adhesion and binds intercellular adhesion molecule 1

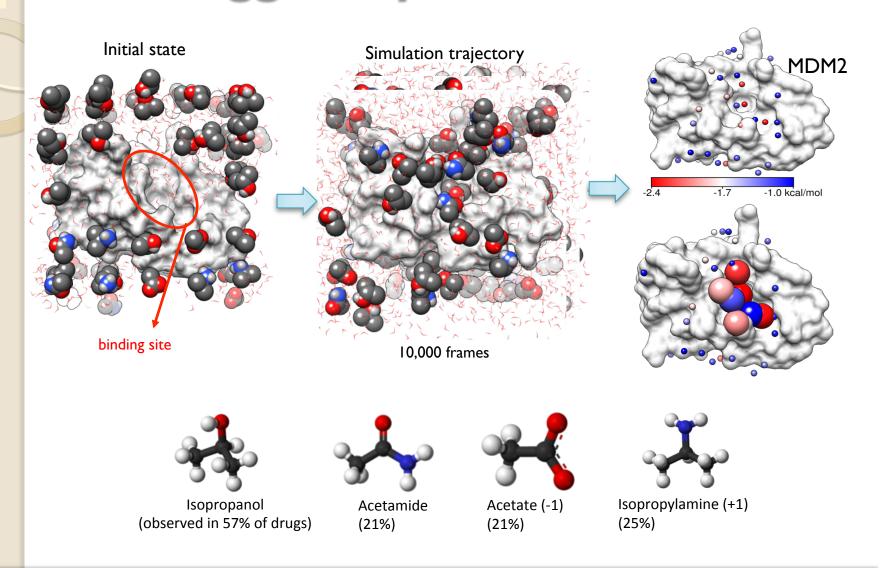
Active site druggability:

➤ Best known K_d 18.3 nM

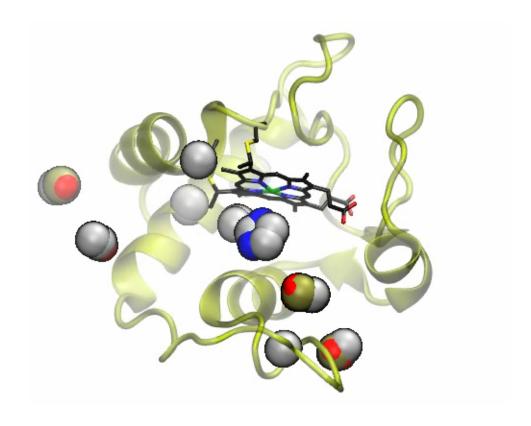
Simulation 0.03-0.5 nM



Druggability Simulations

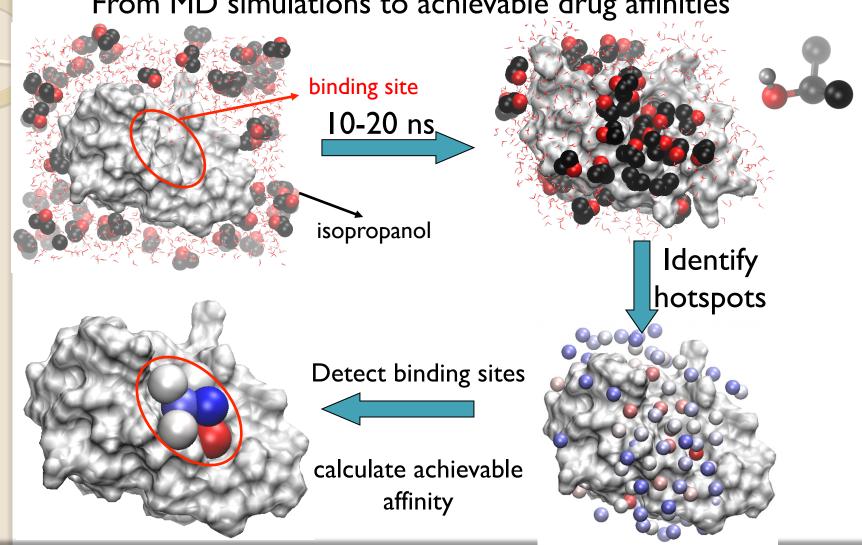


Cytochrome c druggability

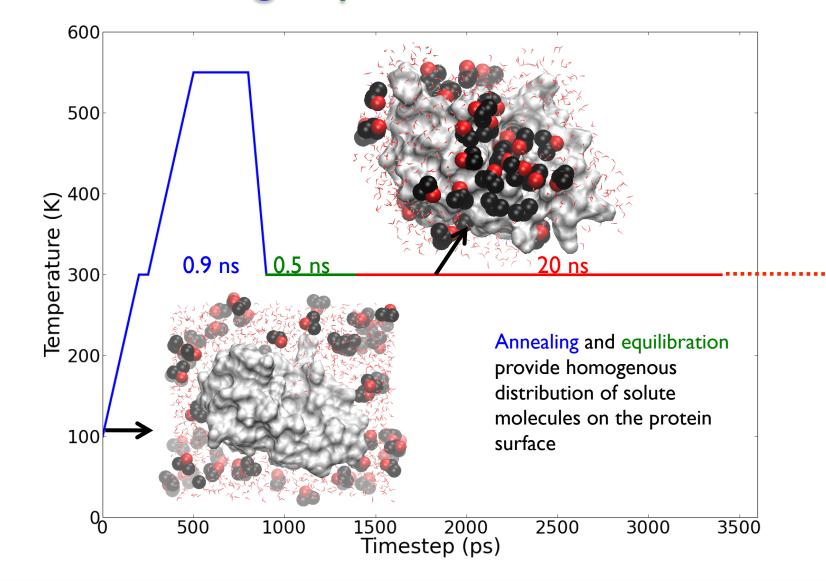


Methodology Overview

From MD simulations to achievable drug affinities



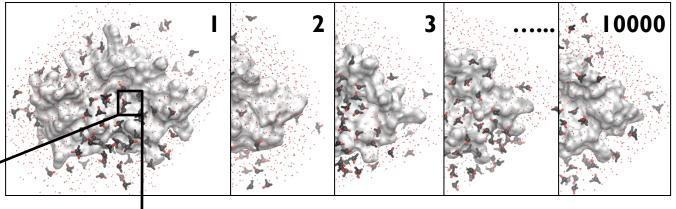
Annealing, Equilibration, Simulation

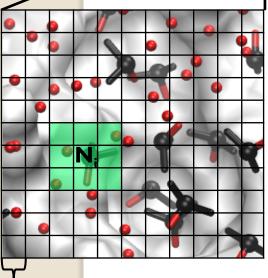


NAMD2 with CHARMM force field was used for simulations.

Free Energy of Binding for Isopropanol

Assuming that MD sampling converged to a **Boltzmann ensemble**





0.5 Å (not to scale)

 $\Delta G_{i} = -RT \ln(N_{i}/N_{0})$

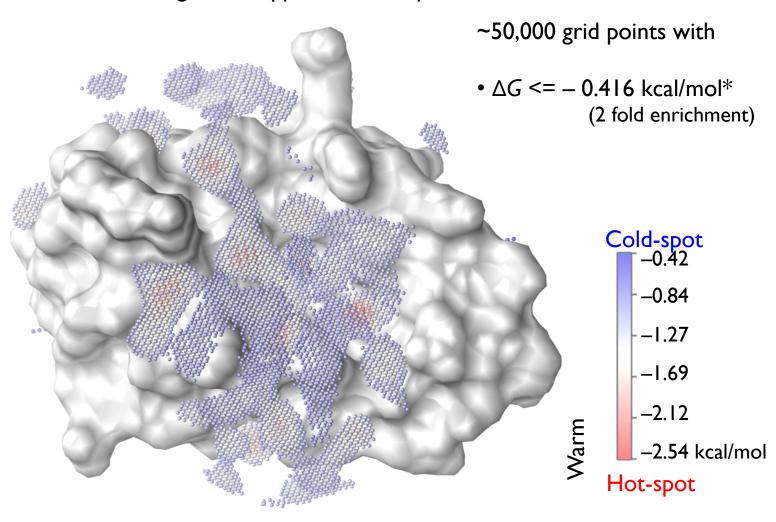
 $N_i = \underline{\text{observed number of isopropanols}}$ (# of frames) * (# of cubes)

 $N_0 = \underline{\text{total number of isopropanols}}$ total number of frames

N_i corresponds to the central highlighted grid element; number of cubes is introduced if multiple cubes are occupied by a single isopropanol

Isopropanol Binding Spots

 ΔG grid is mapped onto the protein structure



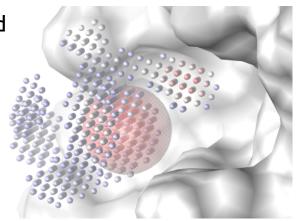
9/4/2009

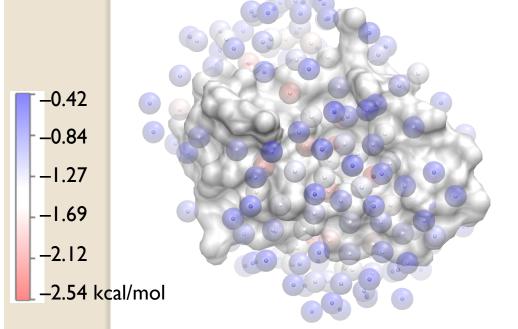


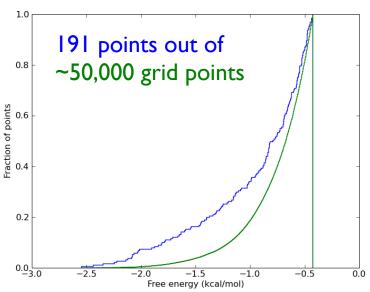
I. Grid element with lowest ΔG value is selected

 Other elements within 4 A are removed (elements inside the red sphere ->)

 I and 2 are repeated until no more points are left to remove







9/4/2009

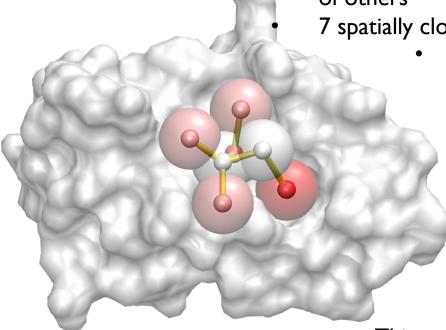
Affinity of a Drug-size Molecule

A heuristic approach for calculating achievable free energy of binding

Assuming binding of an isopropanol is independent of others

7 spatially close binding spots are selected

The sum of $\Delta G_{binding}$ of individual points is considered as a binding free energy estimate that is achievable by a drug-like molecule



This way, the highest affinity we can observe is 5 fM (10^{-15}) .

-0.42

-0.84

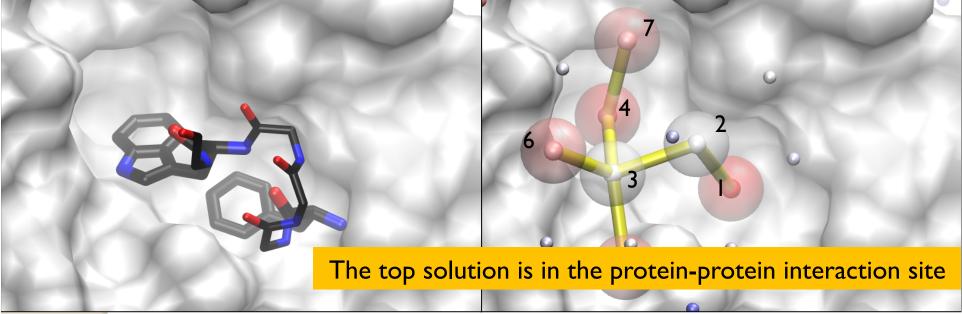
-1.27

-1.69

MDM2: p53 binding site

p53 peptide key interactions (X-ray)

Highest affinity solution (7 points)



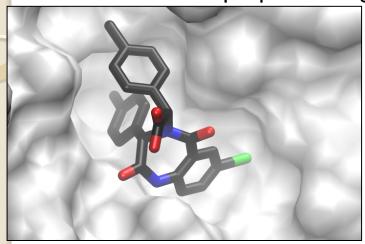
Numbers indicate the order that hot spots were merged by the growing algorithm

Predicted binding affinity range : 0.05-0.3 nM

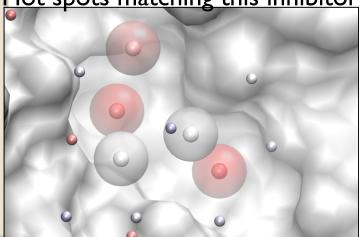
Predicted max. affinity by Seco et al. : 0.02 nM

MDM2: p53 binding site

An inhibitor that disrupts p53 binding



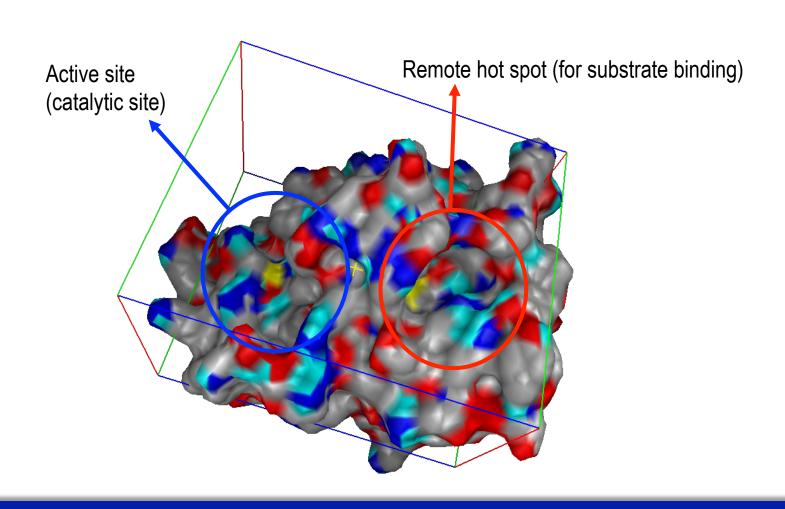
Hot spots matching this inhibitor



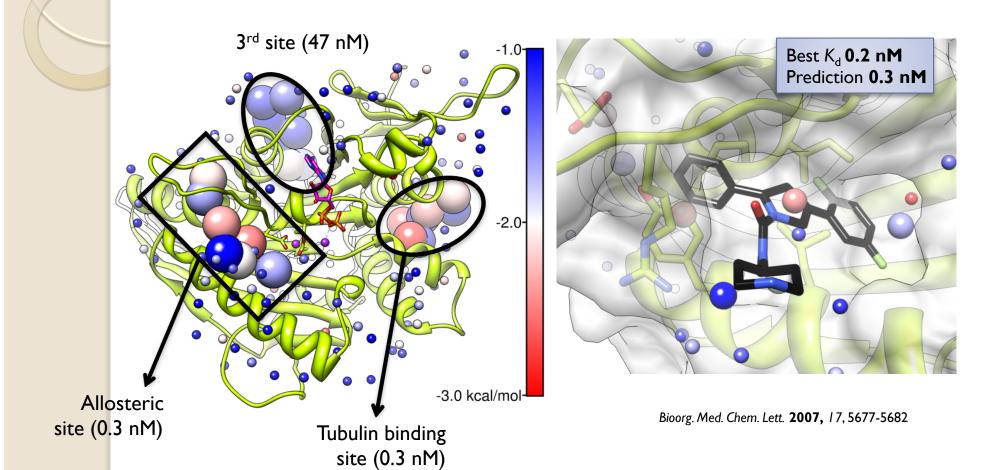
Correspondence of inhibitor in the hot spot volume

Predicted K_d : **47 nM** Known K_d : **80 nM**

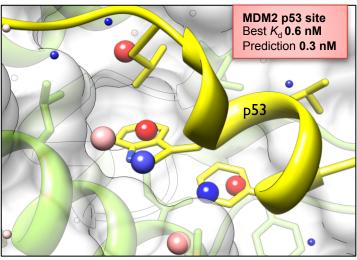
Proteins may have multiple target sites



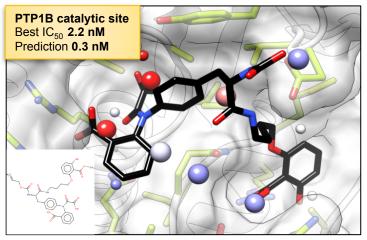
eg5 Druggable Sites



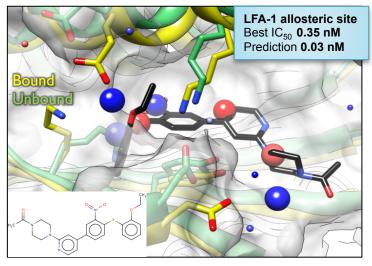
Assessment of druggable allosteric sites



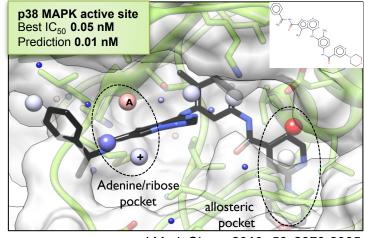
J Med. Chem. 2009, 52, 7970-7973



Bioorg. Med. Chem. Lett. 2003, 13, 3947-3950

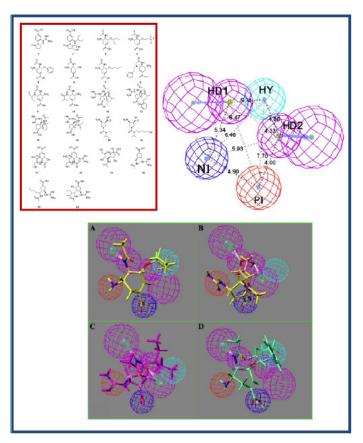


Biochemistry 2004, 43, 2394-2404



J Med. Chem. 2010, 53, 2973-2985

Probe distributions are used for building pharmacophore models



Zhang et al (2006) Bioorganic & Medicinal Chemistry Letters 16, 3009

Pharmacophore Model: PM

Starting point: a series of hits

Method: clustering, identifying commonalities, assigning weights

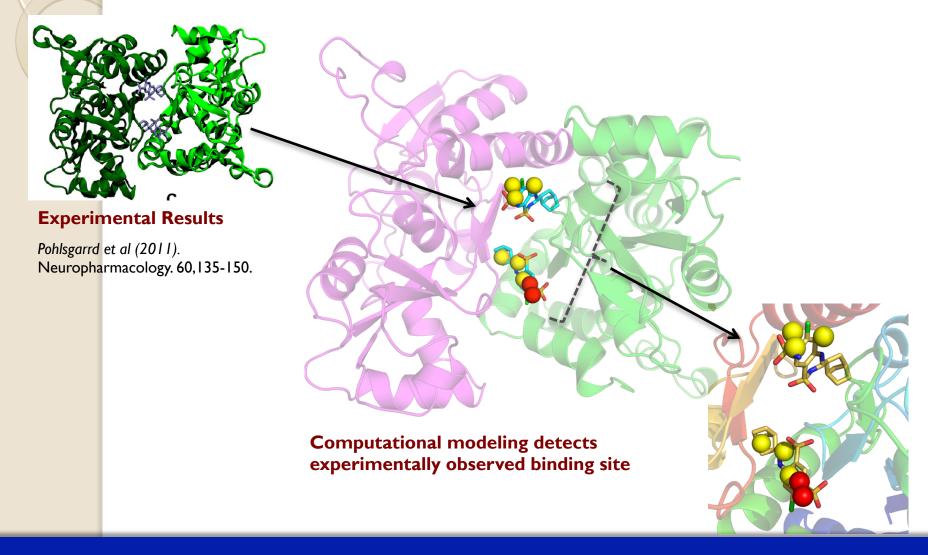
Used for screening

- approved drugs
- -libraries of small compounds

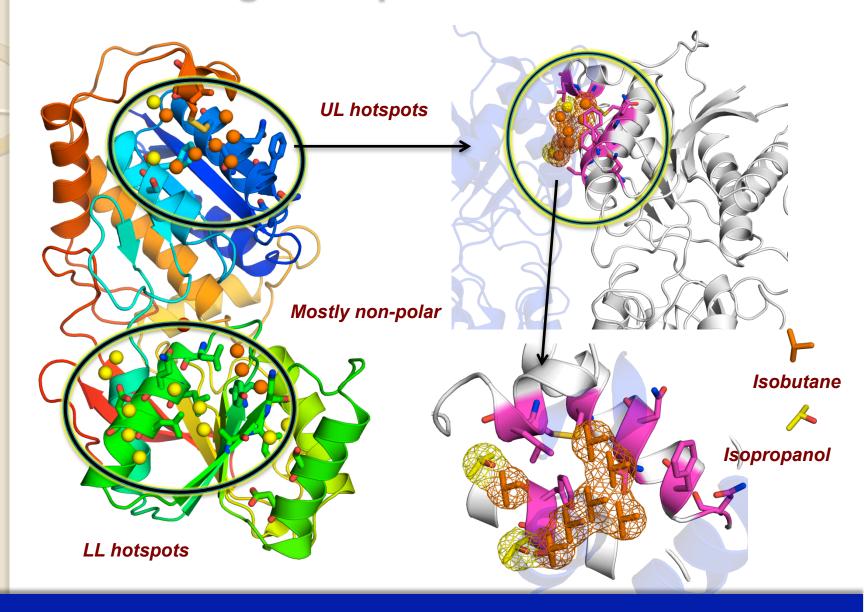
To identify repurposable or new drugs

2

Probes capture allosteric modulator site of AMPAR LBD Dimer



Interfacial regions captured in AMPAR NTD

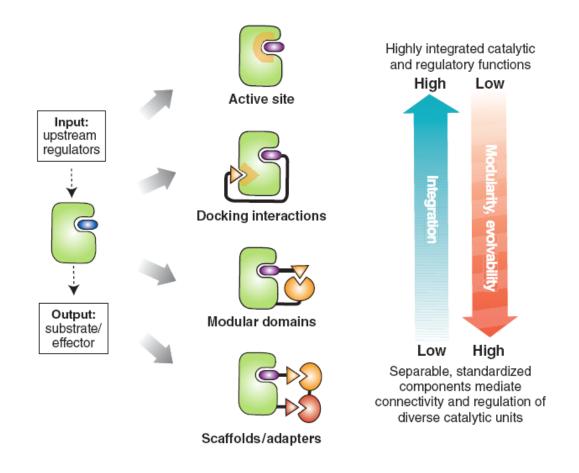




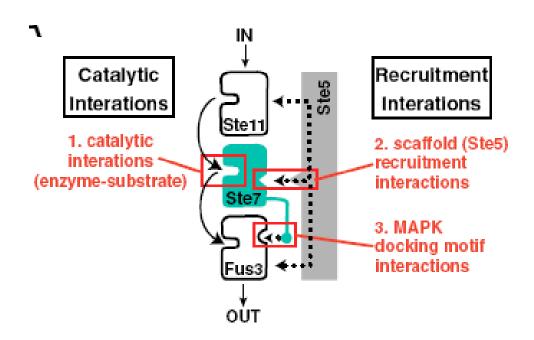
SUMMARY

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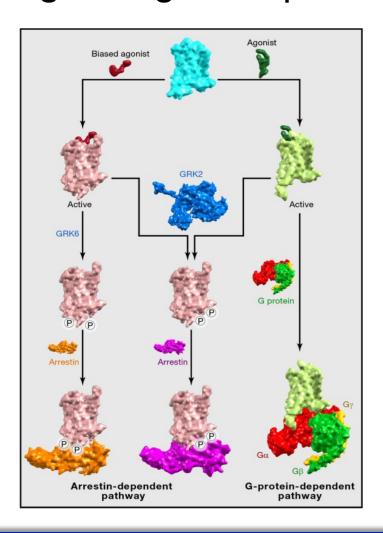
Diversity & complexity of phenotypes arise from combinations of proteins & modular domains



Significance of targeting a specific site, not only a target protein



Allostery Can Diversify Cellular Signaling Pathways through a Single Receptor



GPCRs use conformational selection to shape signaling.

Two (different) conformations of GPCR bind two (different) agonists, which branch into two pathways

Protein Promiscuity

Many proteins are involved in multiple pathways.

Depending on the targeted surface region, or on the accessible structural change/dynamics

the interactions with different (or multiple) upstream or downstream partners/substrates may be affected,

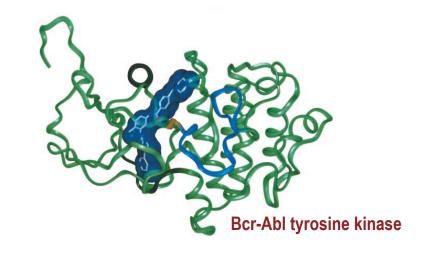
which in turn would impact different (or multiple) pathways, and may result in various phenotypes

Assessment of druggable allosteric sites

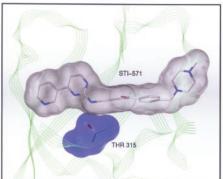
Imatinib (Gleevec)



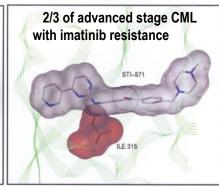
Imatinib was developed for chronic myelogenous leukemia (CML), but was also used for gastrointestinal stromal tumors (GISTs) and some other diseases.



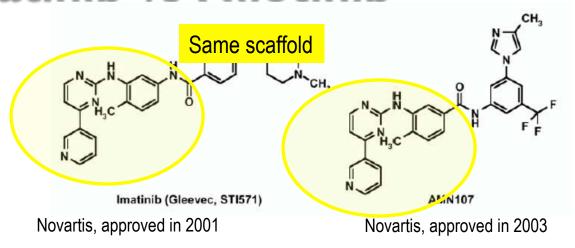
WILD-TYPE

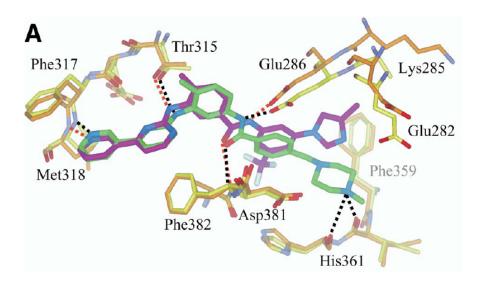


T3151 MUTANT



Imatinib vs Nilotinib



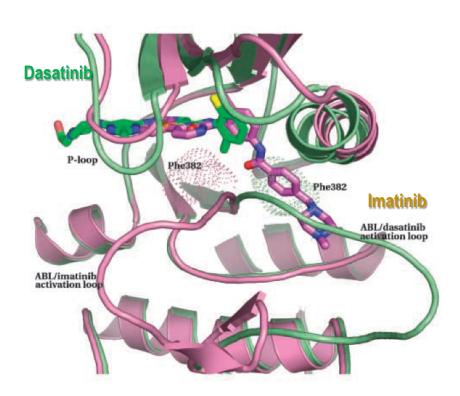


Dasatinib addresses imatinib resistance mutations, but fails with mutant T315I

Dasatinib

Bristol Myers Squibb, approved in 2011

Scaffold hopping via pharmacophore modeling



Cancer Res. 2006, 66: 5790-7.

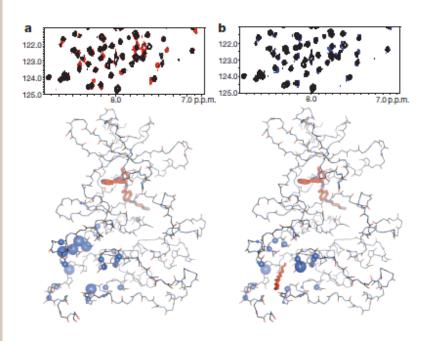
ARTICLES

Targeting Bcr-Abl by combining allosteric with ATP-binding-site inhibitors

Jianming Zhang¹*, Francisco J. Adrián²*, Wolfgang Jahnke³, Sandra W. Cowan-Jacob³, Allen G. Li², Roxana E. Iacob⁴, Taebo Sim^{1,5}, John Powers⁶, Christine Dierks², Fangxian Sun², Gui-Rong Guo², Qiang Ding², Barun Okram⁷, Yongmun Choi¹, Amy Wojciechowski¹, Xianming Deng¹, Guoxun Liu², Gabriele Fendrich³, André Strauss³, Navratna Vajpai⁸, Stephan Grzesiek⁸, Tove Tuntland², Yi Liu², Badry Bursulaya², Mohammad Azam⁶, Paul W. Manley³, John R. Engen⁴, George Q. Daley⁶, Markus Warmuth⁹ & Nathanael S. Gray¹

GNF-2 binds to the myristate-binding site of AbI, leads to changes in the structural dynamics of the protein, and thus inhibits allosteric interactions!

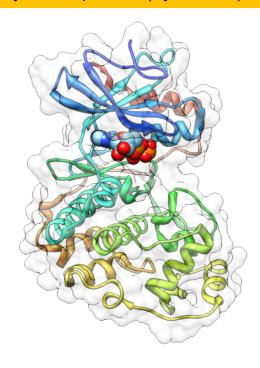
Polypharmacological strategy: Inhibition of allosteric interaction site in addition to catalytic site



Evidence for GNF-2 binding to the myristate pocket of Abl. HSQC spectrum of Abl/imatinib with (red) and without (black) GNF-2 (top) shows chemical shift changes induced by ligand binding. Mapping of chemical shift changes to structure (PDB 10PK8) identifies the myristate pocket as the GNF-2 binding site. b, Same as a except myristic acid used instead of GNF-2.

Simultaneously targeting of

- the ATP binding site (by Gleevec)
- the myristate pocket (by GNF-2)



Khateb et al. <u>BMC Cancer.</u> 2012 Overcoming Bcr-Abl T315I mutation by combination of GNF-2 and ATP competitors in an Abl-independent mechanism.

Quantitative Systems Pharmacology: Integrating Quantitative Models with Experimental Data for Drug Discovery Refined cellular **Signals** pathways extracted /processes from the data Models **Predicted Drug** Information binding to targets **DrugBank** Data **Predictions High-Content Scre Targeted Libraries** Clinical Trials Measurements of **Experimental** biological functions testing 3D or 4D images



A Role for Computational Biology

Box 2 A role for computational biology?

Predicting protein promiscuity is a problem of daunting complexity for bioinformaticians. Indeed, earlier work has shown that bioinformatics methods need improving to reliably uncover promiscuous reactions¹¹⁷, and our own *in silico* work in protein function prediction^{118,119} has curbed our optimism. Even so, we do not doubt that certain areas of bioinformatics research will be important for progress in this field. Some of these avenues have already been pursued by computational biologists, but referencing individual studies is outside the scope of this review.

Data analysis. There is a great deal of data on protein promiscuity to be found in function-related databases. Collating information on promiscuous proteins would be a necessary first step, and existing enzyme, pathway or ontology databases can provide a lot of information on proteins with multiple EC numbers, reactions or substrates, or function categories for genes. Clues as to moonlighting might also be found from expression data. Unexpected expression patterns that do not correlate with our knowledge of existing networks are often indicative of a moonlighting function for a protein. Such data would be complemented by data on alternative splicing of a single gene, which will give hints to any additional roles in the cell.

Sequence-based methods. Large protein families with many relatives may indicate a trend toward promiscuity. Is there a correlation between number of orthologs and number of paralogs and how could it be explained?

Structure-based methods. Analyzing binding site characteristics could reveal those that make proteins more amenable to promiscuity.

Docking profiles. Probing the binding site with panels of selected ligands or other proteins can assess how restrictive the site is toward different types of molecules.

Flexibility. *In silico* studies of the flexibility of proteins can reveal how this may contribute to recognizing multiple partners.

Redundancy in pathways. The evidence of redundancy in metabolic and regulatory networks should be examined carefully, as it may also contain evidence for protein functional promiscuity.

Calculation of promiscuity indices. This could be based on *in silico* or experimental data and could help rank proteins and their partners according to their interaction promiscuity.

Mapping of small-molecule space to protein space. This would reveal any preferences of protein families for sets of chemical groups and possibly allow the engineering of mutatants capable of binding small molecules from neighboring parts of the chemical space.

These are only some possible directions that could be explored to improve our chances of successfully exploiting promiscuity. Experimental verification of any rules learned and predictions made will be indispensable.