

Force Fields for Classical Molecular Dynamics simulations of Biomolecules

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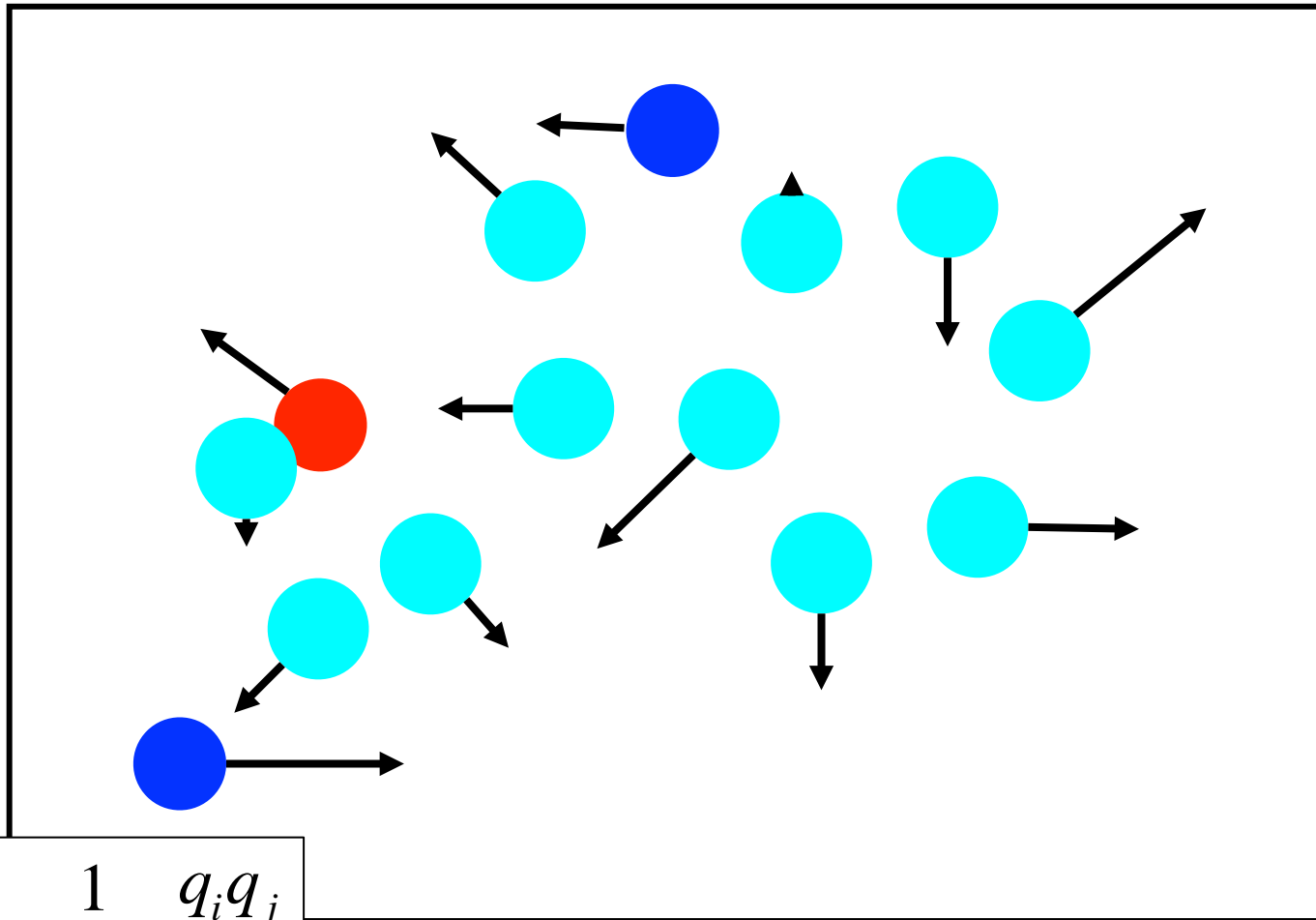
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University of Illinois at Urbana-Champaign

Classical Force Field Parameters

- Topology and structure files
- Parameter files
- Where do all the numbers needed by an MD code come from?
- Where to find these numbers and how to change them if needed.
- How to make topology files for ligands, cofactors, special amino acids, ...
- How to develop / put together missing parameters.

Classical Molecular Dynamics

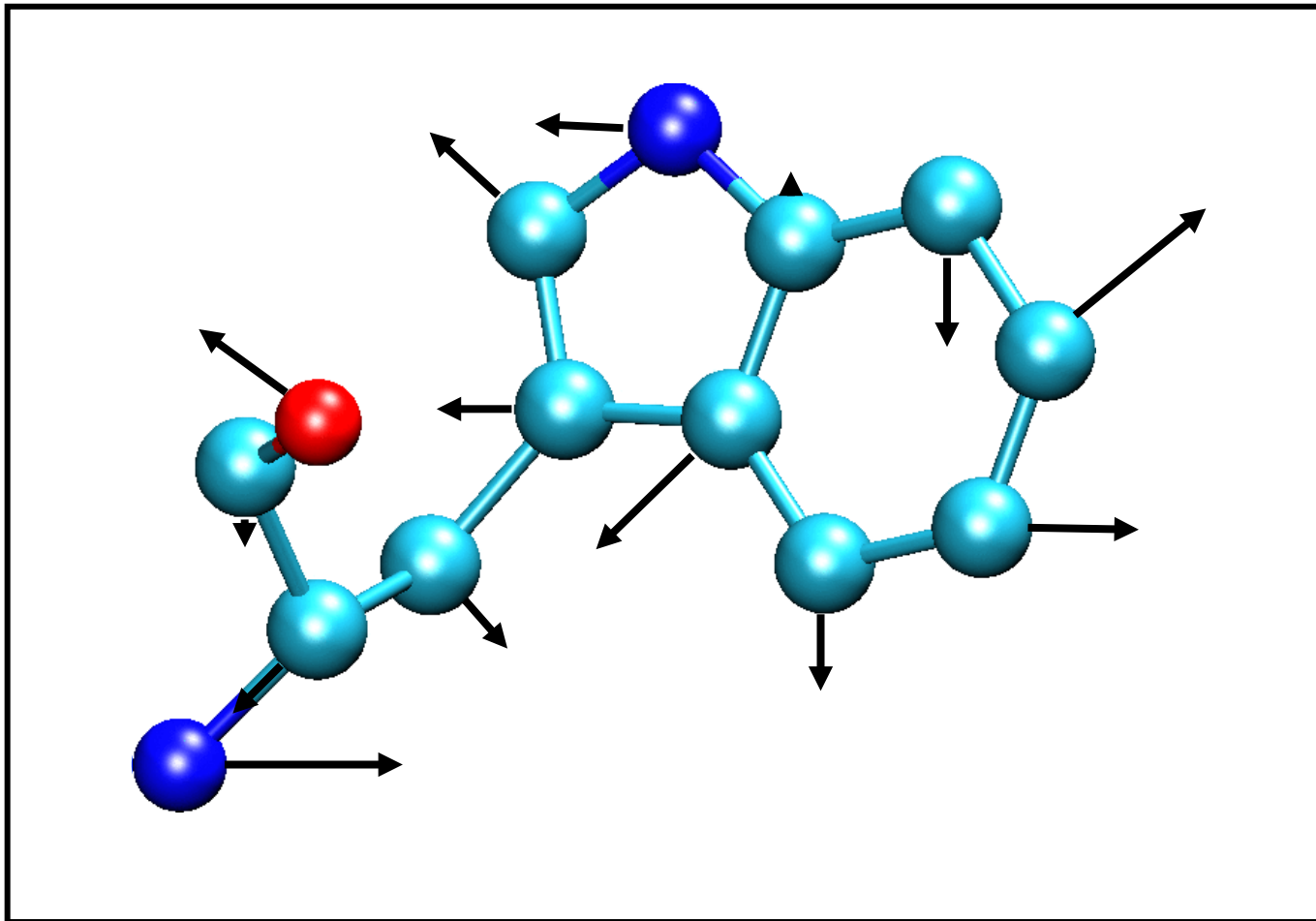


$$U(r) = \frac{1}{4\pi\epsilon_0} \frac{q_i q_j}{r_{ij}}$$

Coulomb interaction

$$U(r) = \epsilon_{ij} \left[\left(\frac{R_{min,ij}}{r_{ij}} \right)^{12} - \left(\frac{R_{min,ij}}{r_{ij}} \right)^6 \right]$$

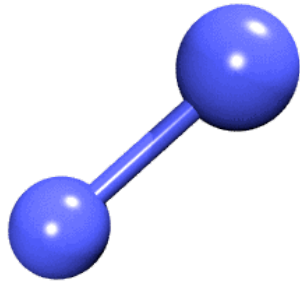
Classical Molecular Dynamics



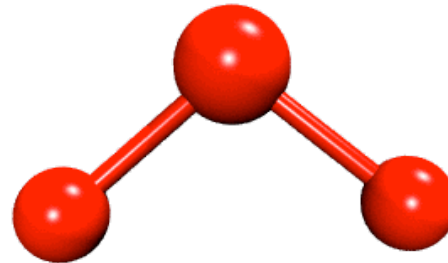
Bond definitions, atom types, atom names, parameters,

Energy Terms Described in

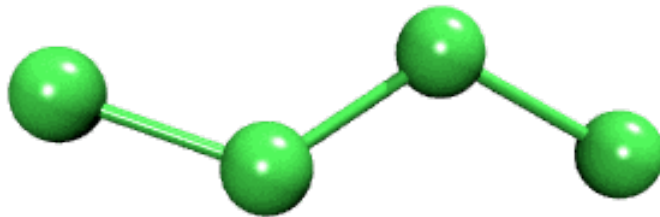
Bond



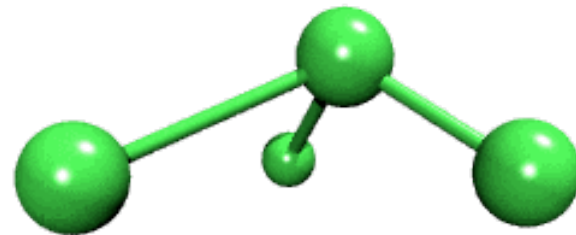
Angle



Dihedral



Improper



The Potential Energy Function

$$\begin{aligned}
 U(\vec{R}) = & \underbrace{\sum_{\text{bonds}} k_i^{\text{bond}} (r_i - r_0)^2}_{U_{\text{bond}}} + \underbrace{\sum_{\text{angles}} k_i^{\text{angle}} (\theta_i - \theta_0)^2}_{U_{\text{angle}}} + \\
 & \underbrace{\sum_{\text{dihedrals}} k_i^{\text{dihe}} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{\text{dihedral}}} + \\
 & \underbrace{\sum_i \sum_{j \neq i} 4\epsilon_{ij} \left[\left(\frac{\sigma_{ij}}{r_{ij}} \right)^{12} - \left(\frac{\sigma_{ij}}{r_{ij}} \right)^6 \right]}_{U_{\text{nonbond}}} + \sum_i \sum_{j \neq i} \frac{q_i q_j}{\epsilon r_{ij}}
 \end{aligned}$$

U_{bond} = oscillations about the equilibrium bond length

U_{angle} = oscillations of 3 atoms about an equilibrium bond angle

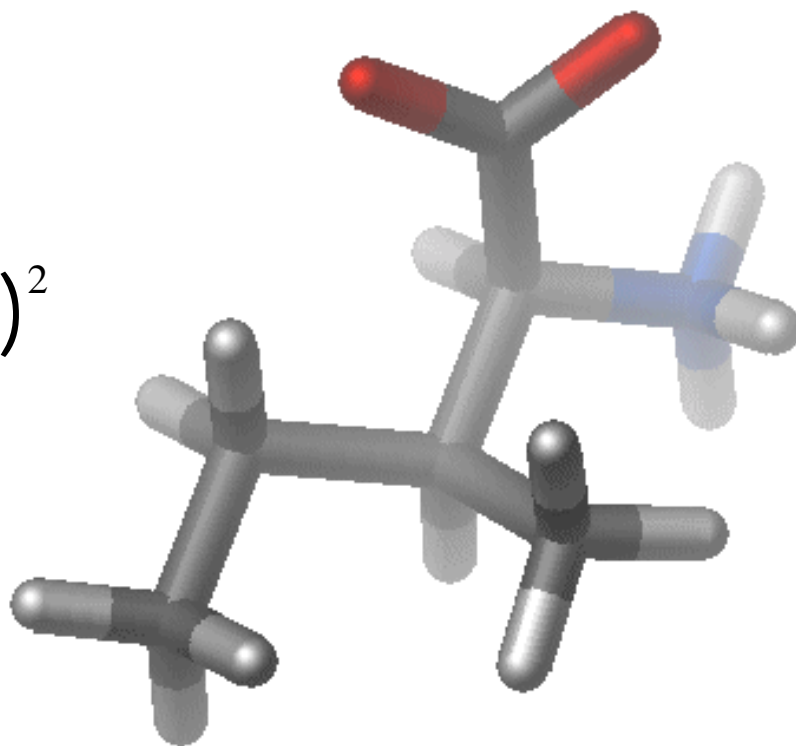
U_{dihedral} = torsional rotation of 4 atoms about a central bond

U_{nonbond} = non-bonded energy terms (electrostatics and Lenard-Jones)

Interactions between bonded atoms

$$V_{angle} = K_{\theta} (\theta - \theta_o)^2$$

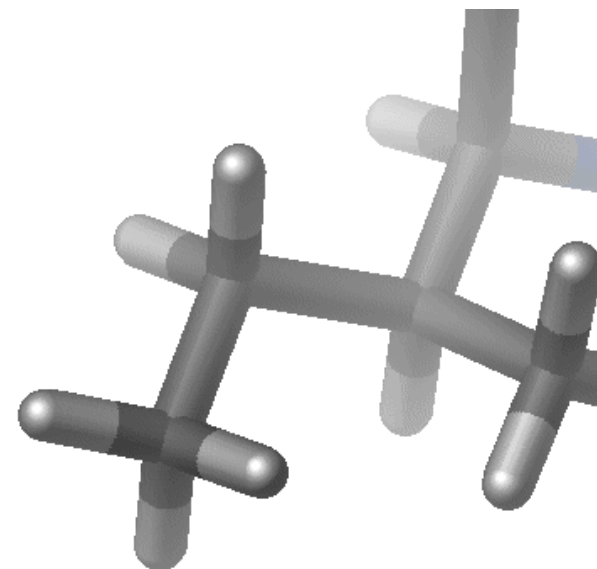
$$V_{bond} = K_b (b - b_o)^2$$



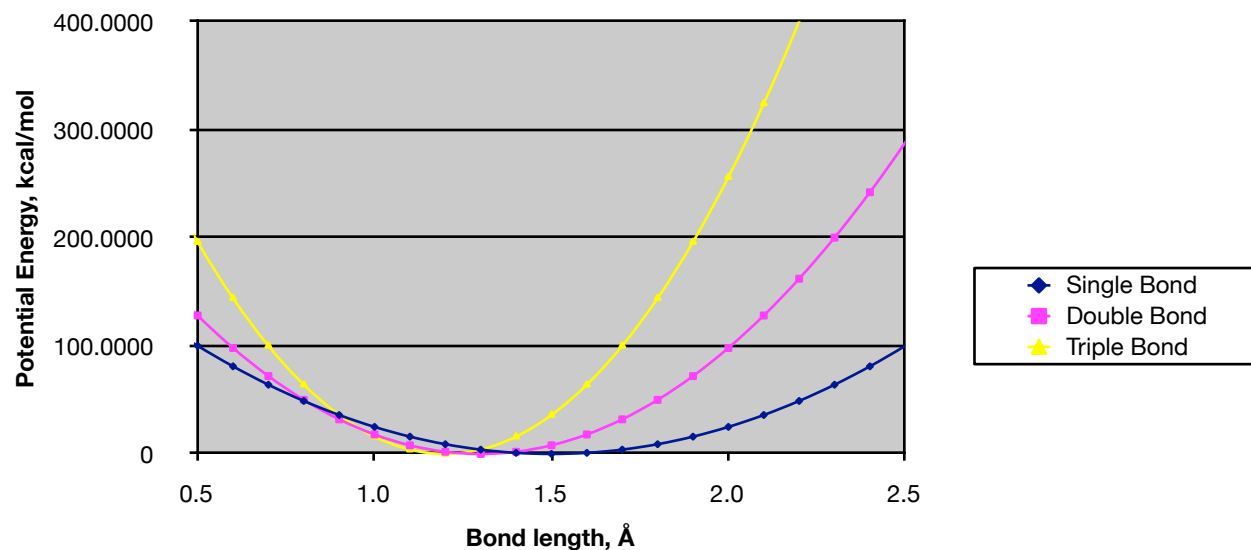
$$V_{dihedral} = K_{\phi} (1 + \cos(n\phi - \delta))$$

$$V_{bond} = K_b (b - b_o)^2$$

Chemical type	K_{bond}	b_o
C-C	100 kcal/mole/Å ²	1.5 Å
C=C	200 kcal/mole/Å ²	1.3 Å
C≡C	400 kcal/mole/Å ²	1.2 Å



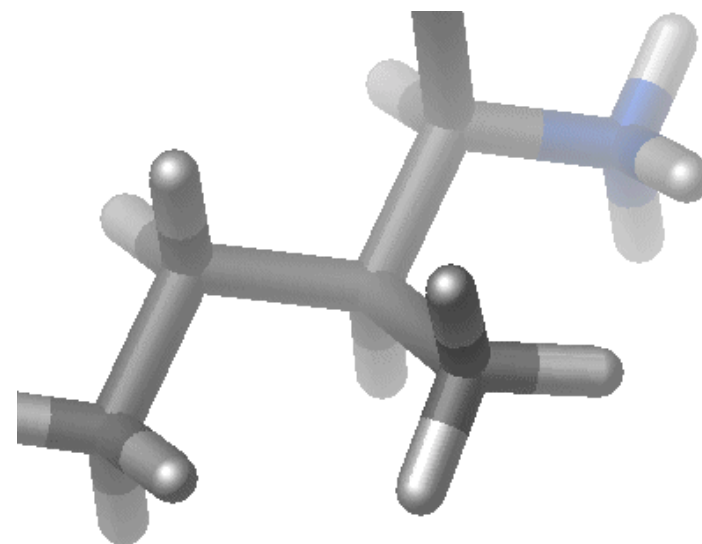
Bond Energy versus Bond length



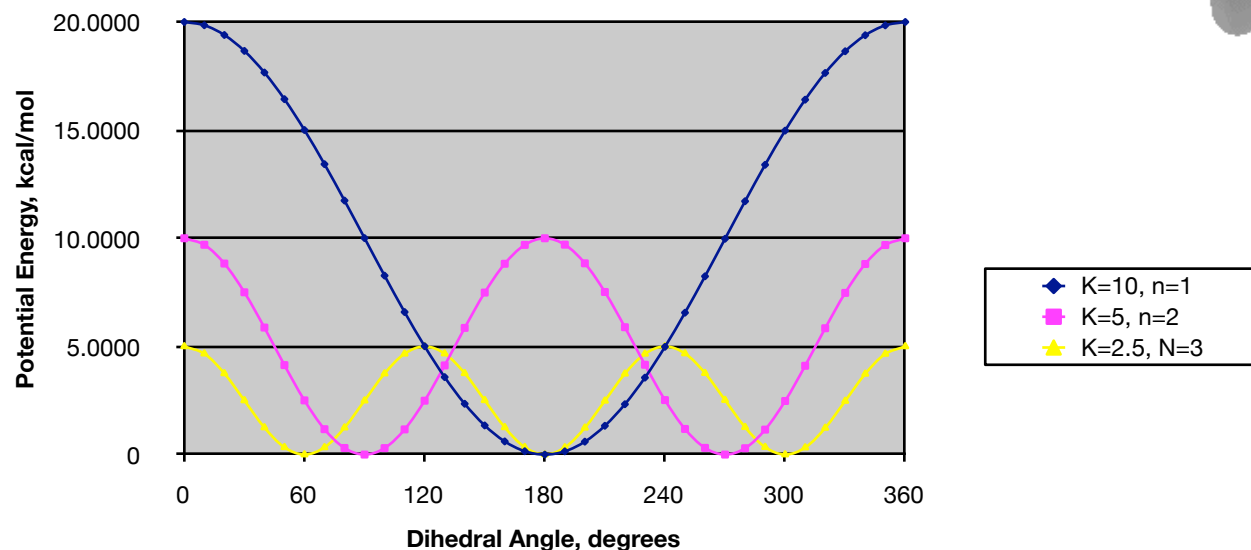
Bond angles and improper terms have similar quadratic forms, but with softer spring constants. The force constants can be obtained from vibrational analysis of the molecule (experimentally or theoretically).

Dihedral Potential

$$V_{dihedral} = K_{\phi} (1 + \cos(n\phi - \delta))$$



Dihedral energy versus dihedral angle



$$\delta = 0^\circ$$

Nonbonded Parameters

$$\sum_{\text{non-bonded}} \frac{q_i q_j}{4\pi D r_{ij}} + \epsilon_{ij} \left[\left(\frac{R_{\min,ij}}{r_{ij}} \right)^{12} - \left(\frac{R_{\min,ij}}{r_{ij}} \right)^6 \right]$$

q_i : partial atomic charge

D : dielectric constant

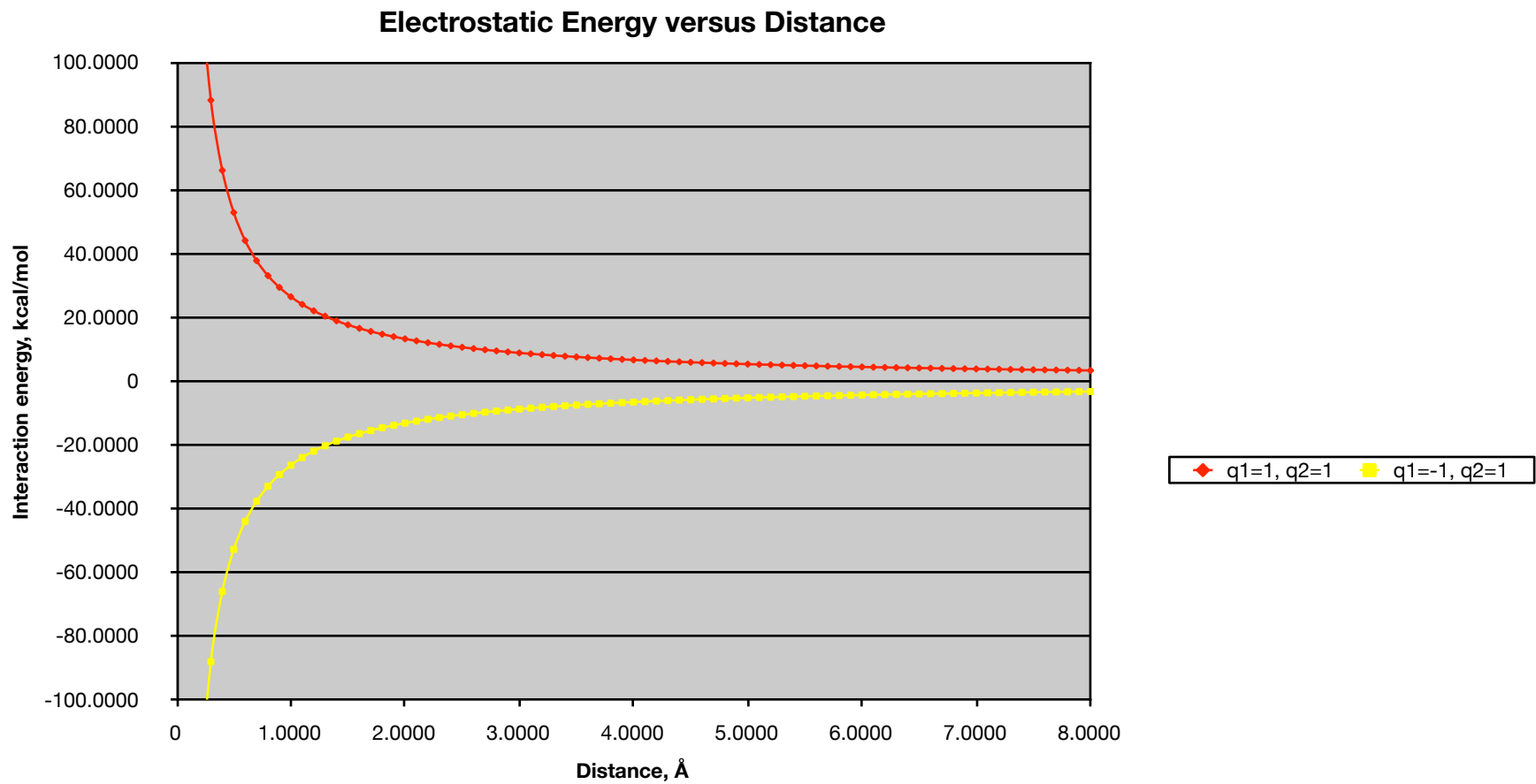
ϵ : Lennard-Jones (LJ, vdW) well-depth

R_{\min} : LJ radius ($R_{\min}/2$ in CHARMM)

Combining rules (CHARMM, Amber)

$$R_{\min i,j} = R_{\min i} + R_{\min j}$$

$$\epsilon_{i,j} = \text{SQRT}(\epsilon_i * \epsilon_j)$$



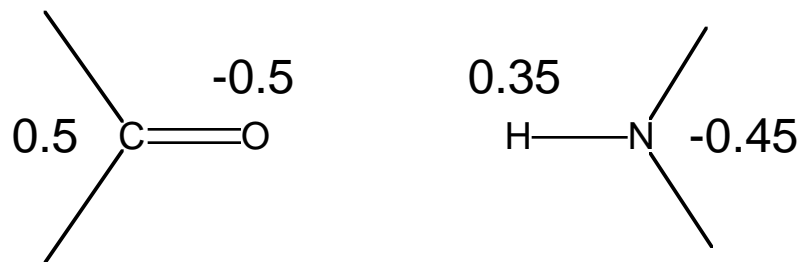
Note that the effect is long range.

Charge Fitting Strategy

CHARMM- Mulliken*

AMBER(ESP/RESP)

Partial atomic charges



*Modifications based on interactions with TIP3 water

CHARMM Potential Function

$$\begin{aligned}
 U(\vec{R}) = & \underbrace{\sum_{\text{bonds}} k_i^{\text{bond}} (r_i - r_0)^2}_{U_{\text{bond}}} + \underbrace{\sum_{\text{angles}} k_i^{\text{angle}} (\theta_i - \theta_0)^2}_{U_{\text{angle}}} + \\
 & \underbrace{\sum_{\text{dihedrals}} k_i^{\text{dihe}} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{\text{dihedral}}} + \underbrace{\sum_i \sum_{j \neq i} 4 \epsilon_{ij} \left[\left(\frac{\sigma_{ij}}{r_{ij}} \right)^{12} - \left(\frac{\sigma_{ij}}{r_{ij}} \right)^6 \right]}_{U_{\text{nonbond}}} + \sum_i \sum_{j \neq i} \frac{q_i q_j}{\epsilon r_{ij}}
 \end{aligned}$$

PDB file
geometry
 Topology
 PSF file
parameters
 Parameter file

File Format/Structure

- The structure of a pdb file
- The structure of a psf file
- The topology file
- The parameter file
- Connection to potential energy terms

Looking at File Structures

- PDB file
- Topology file
- PSF file
- Parameter file

Parameter Optimization Strategies

Check if it has been parameterized by somebody else

Literature

Google

Minimal optimization

By analogy (direct transfer of known parameters)

Quick, starting point

Maximal optimization

Time-consuming

Requires appropriate experimental and target data

Choice based on goal of the calculations

Minimal

database screening

NMR/X-ray structure determination

Maximal

free energy calculations, mechanistic studies,

subtle environmental effects

Getting Started

- Identify previously parameterized compounds
- Access topology information – assign atom types, connectivity, and charges – **annotate changes**

CHARMM topology (parameter files)

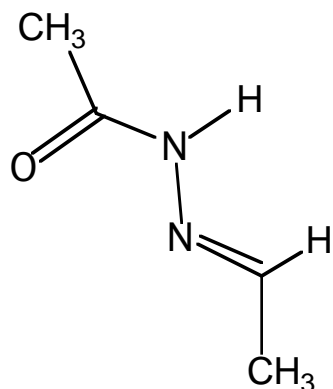
top_all22_model.inp (par_all22_prot.inp)
top_all22_prot.inp (par_all22_prot.inp)
top_all22_sugar.inp (par_all22_sugar.inp)
top_all27_lipid.rtf (par_all27_lipid.prm)
top_all27_na.rtf (par_all27_na.prm)
top_all27_na_lipid.rtf (par_all27_na_lipid.prm)
top_all27_prot_lipid.rtf (par_all27_prot_lipid.prm)
top_all27_prot_na.rtf (par_all27_prot_na.prm)
toph19.inp (param19.inp)

NA and lipid force fields have new LJ parameters for the alkanes, representing increased optimization of the protein alkane parameters. Tests have shown that these are compatible (e.g. in protein-nucleic acid simulations). For new systems is suggested that the new LJ parameters be used. Note that only the LJ parameters were changed; the internal parameters are identical

www.pharmacy.umaryland.edu/faculty/amackere/force_fields.htm

Partial Charge Assignment

- Most important aspect for ligands
- Different force fields might take different philosophies
 - AMBER: RESP charges at the HF/6-31G level
 - Overestimation of dipole moments
 - Easier to set up
 - CHARMM: Interaction based optimization
 - TIP3P water representing the environment
 - Could be very difficult to set up
- Conformation dependence of partial charges
- Lack of polarization
- Try to be consistent within the force field
- pKa calculations for titratable residues

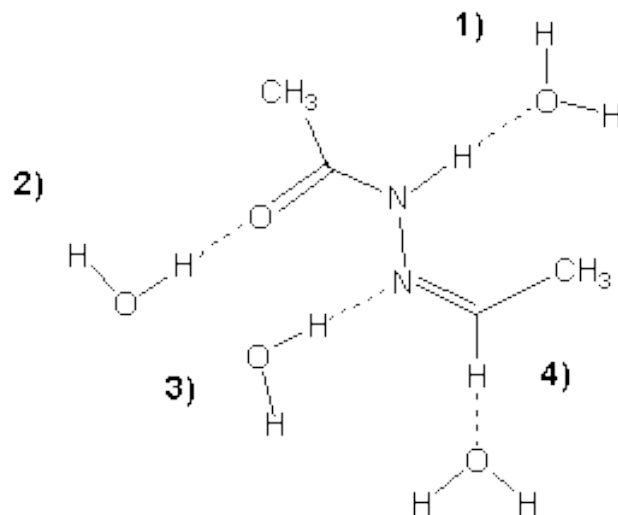


Starting charges??

Mulliken population analysis

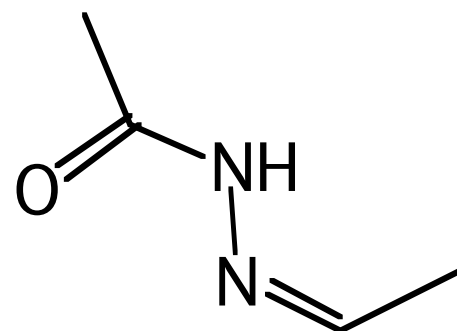
Analogy comparison

Final charges (methyl, vary q_C to maintain integer charge, $q_H = 0.09$)
interactions with water (HF/6-31G*, monohydrates!)

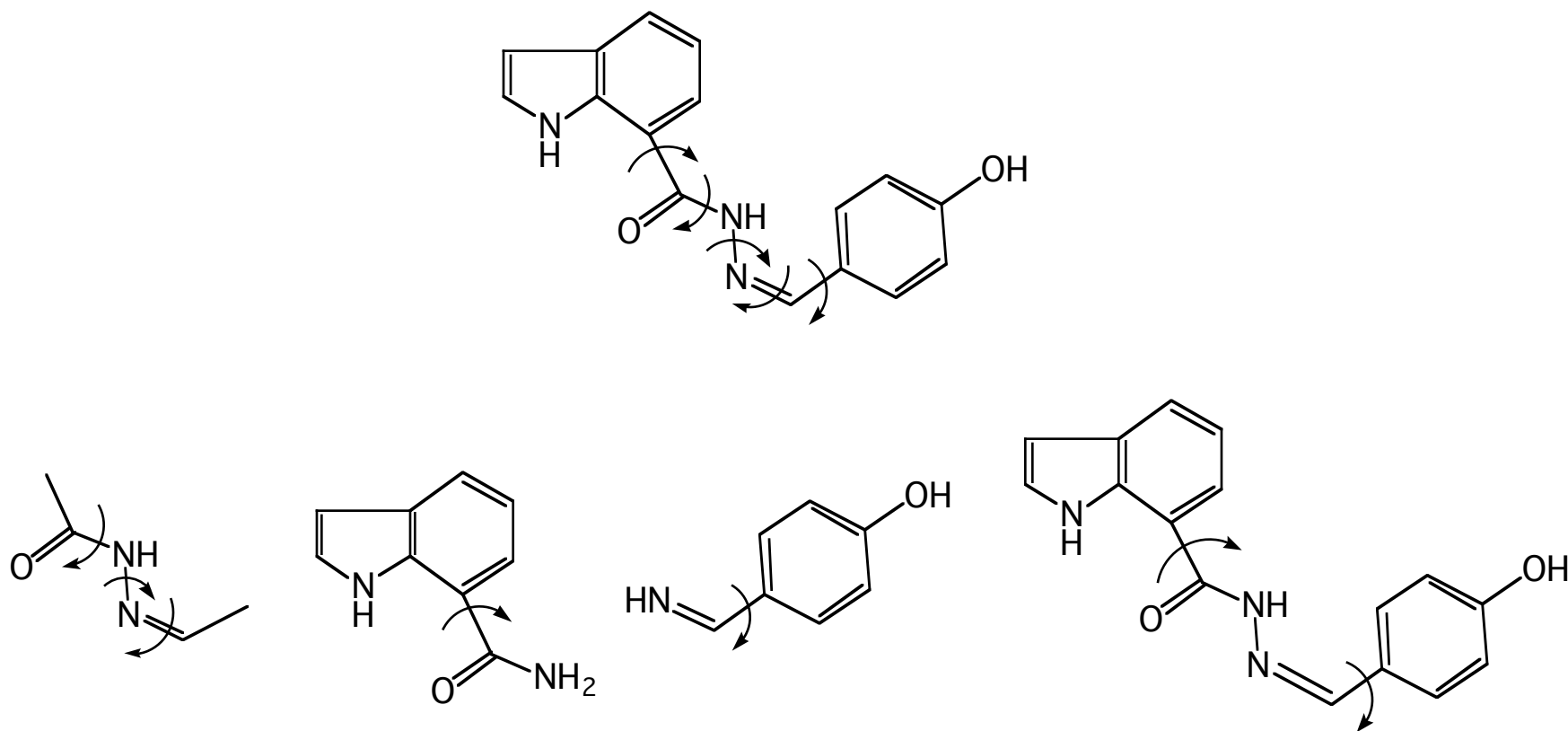


Comparison of analogy and optimized charges

Name	Type	Analogy	Optimized
C1	CT3	-0.27	-0.27
H11	HA3	0.09	0.09
H12	HA3	0.09	0.09
H13	HA3	0.09	0.09
C2	C	0.51	0.58
O2	O	-0.51	-0.50
N3	NH1	-0.47	-0.32
H3	H	0.31	0.33
N4	NR1	0.16	-0.31
C5	CEL1	-0.15	-0.25
H51	HEL1	0.15	0.29
C6	CT3	-0.27	-0.09
H61	HA	0.09	0.09
H62	HA	0.09	0.09
H63	HA	0.09	0.09



Dihedral optimization based on QM potential energy surfaces (HF/6-31G* or MP2/6-31G*).



From MacKerell

A Brief Overview of The Force Field Toolkit (*ffTK*)

Dr. Christopher G. Mayne
Tajkhorshid Group
February 13th, 2012

Available Methods for Obtaining Parameters

Analogy (Re-use of parameters from similar structures)

ParamChem <https://www.paramchem.org/>

SwissParam <http://swissparam.ch/>

Zoete *et al.*; *J. Comp. Chem.* 32(11) **2011**, pp.2359-2368

MATCH <http://brooks.chem.lsa.umich.edu/software>

Yesselman *et al.*; *J. Comp. Chem.* 33(2) **2012**, pp.189-202

Development

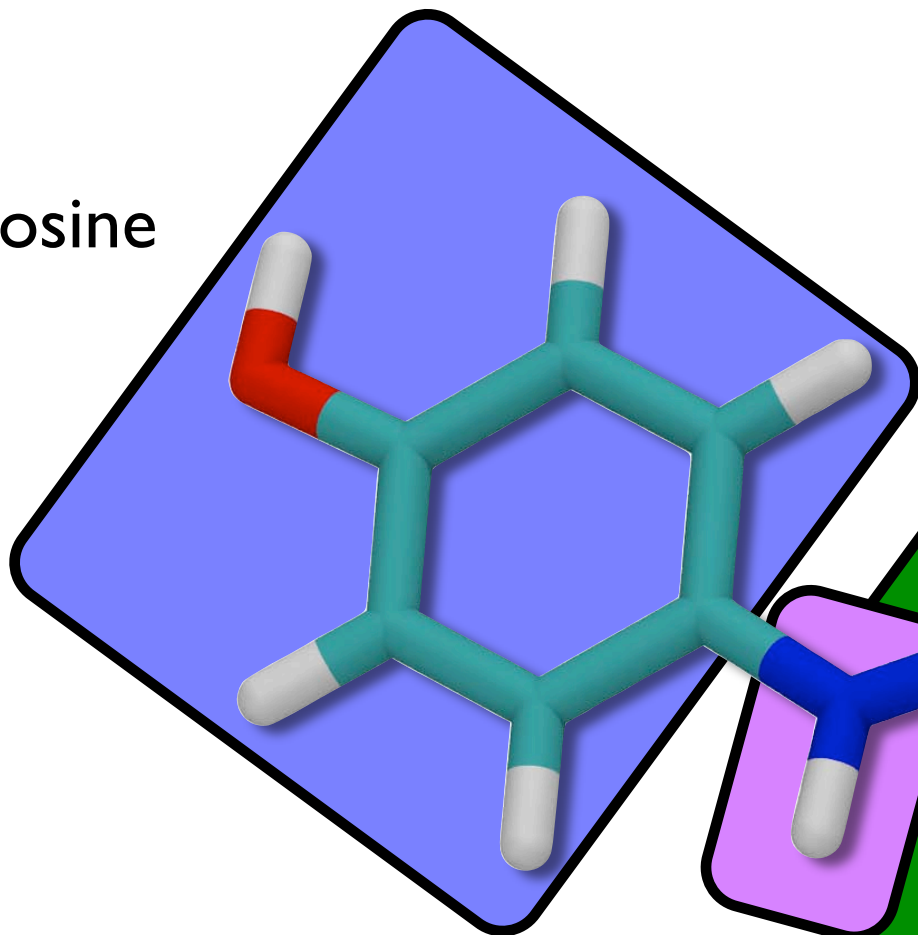
ParaTool <http://www.ks.uiuc.edu/Research/vmd/plugins/paratool/>

GAAMP <http://gaamp.lcrc.anl.gov/para-fit.html>

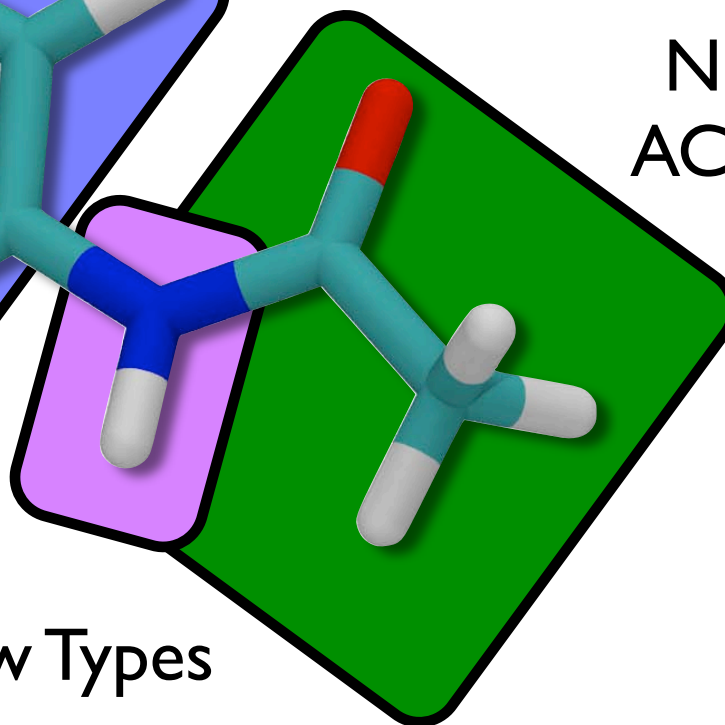
ffTK <http://www.ks.uiuc.edu/Research/vmd/plugins/fftk/>

An Example: Acetaminophen

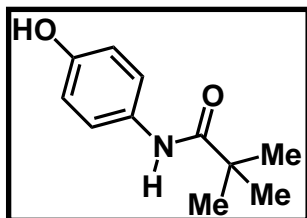
Tyrosine



NMA or
ACE-patch

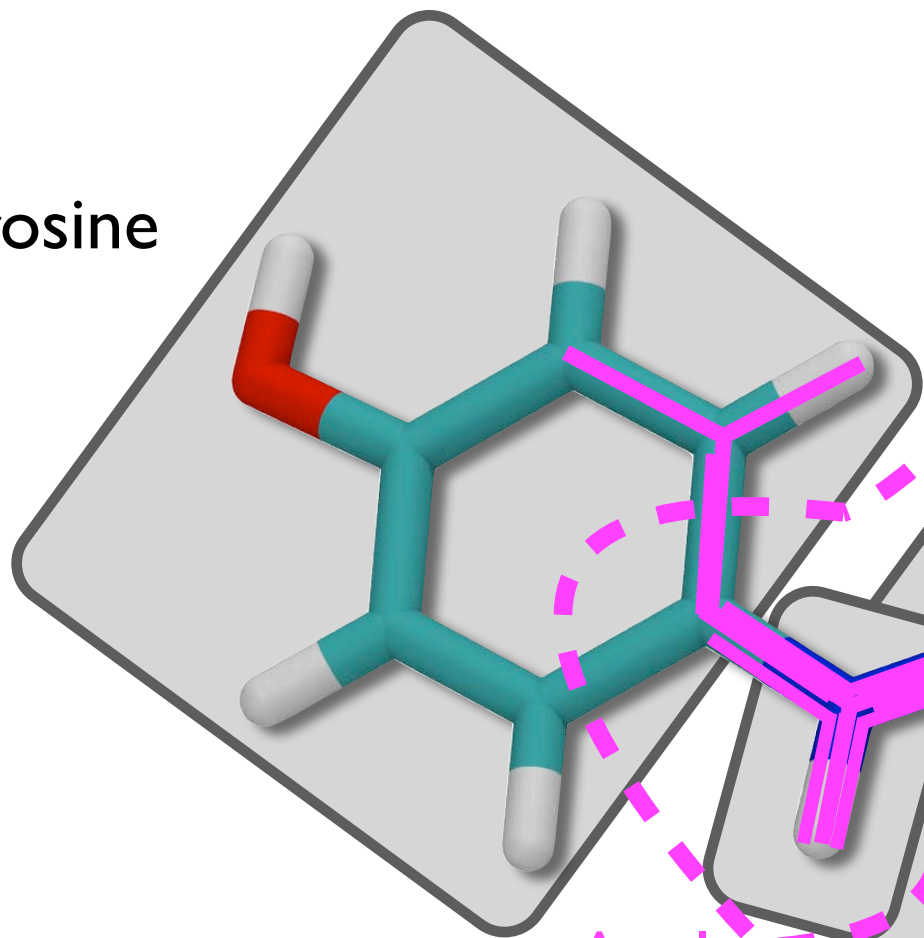


New Types



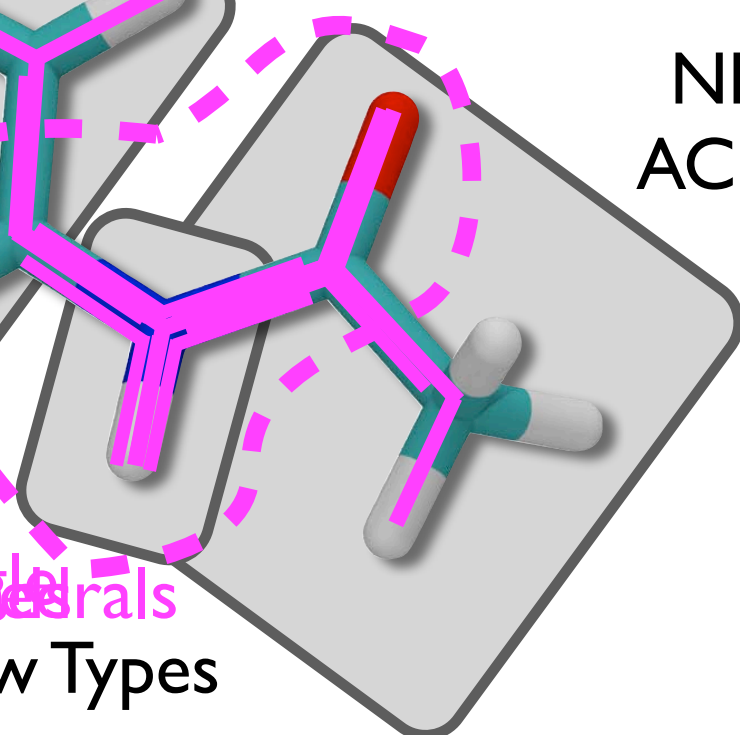
An Example: Acetaminophen

Tyrosine

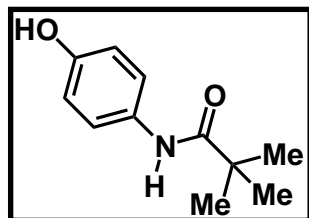


New Charges

NMA or
ACE-patch

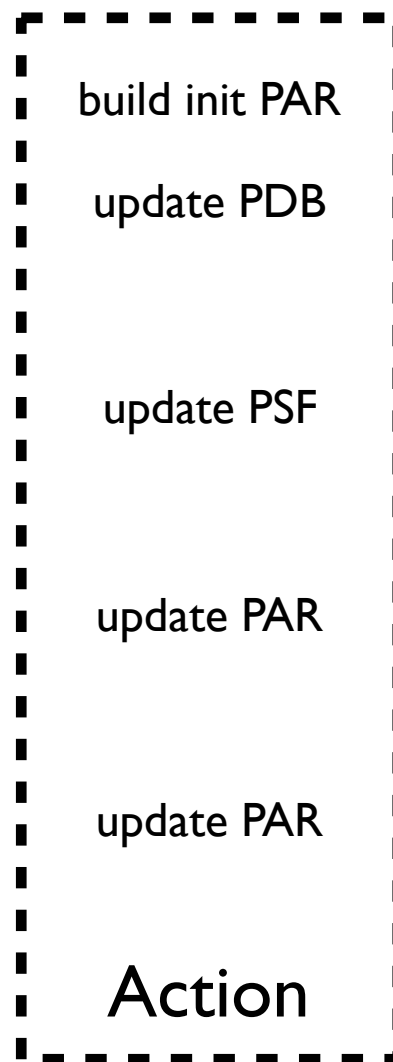
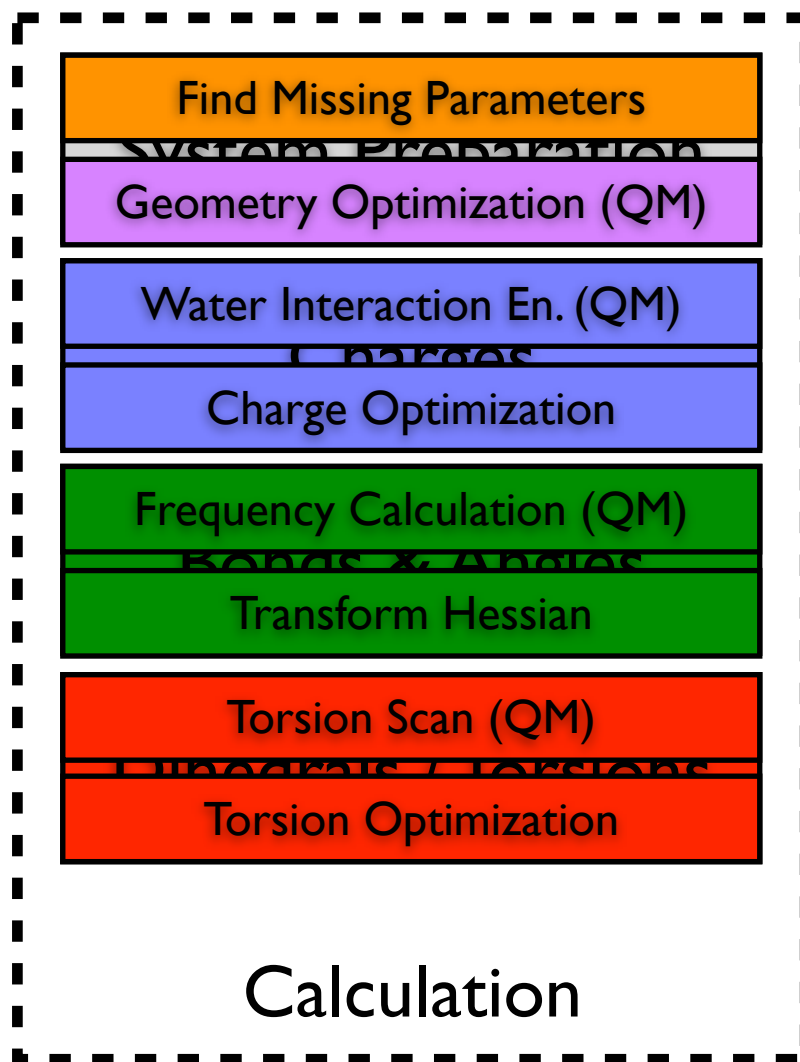


Angles
Bonds
New Types



ffTK Facilitates the Parameterization Workflow

PSF/PDB



PAR File

ffTK Interface

file dialog buttons

entry boxes to hold variables tasks divided into separate tabs action buttons

The image shows a screenshot of the ffTK interface with several annotations. At the top, there are colored tabs: orange, purple, blue, green, and red. Below these are tabs for different tasks: BuildPar, Opt. Geometry, Water Int., Opt. Charges (selected), Calc. Bonded, Scan Torsions, and Opt. Torsions. The main content area is divided into sections: 'Input' with fields for PSF File, PDB File, and Residue Name; 'Parameter Files' with a list of files and 'Add', 'Delete', and 'Clear' buttons; and 'NAMD binary' and 'Output LOG' fields. At the bottom, there is a treeview with expandable sections: Charge Constraints, QM Target Data, Advanced Settings, and Results. Annotations include: 'entry boxes to hold variables' pointing to the PSF File field; 'tasks divided into separate tabs' pointing to the top tabs; 'file dialog buttons' pointing to the 'Browse' buttons; 'action buttons' pointing to 'Load PSF/PDB', 'Add', 'Delete', and 'Clear'; 'action menus' pointing to the 'Label Atoms' dropdown; 'treeview boxes to hold lists' pointing to the bottom treeview; and 'arrow' pointing to the left side of the interface.

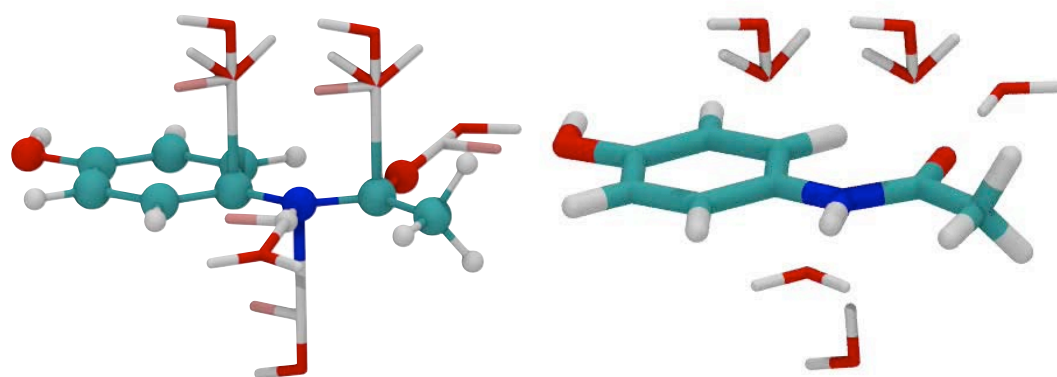
ffTK 1.0 is Available in VMD 1.9.1

Setup necessary QM calculations

Visualize calculation input/output

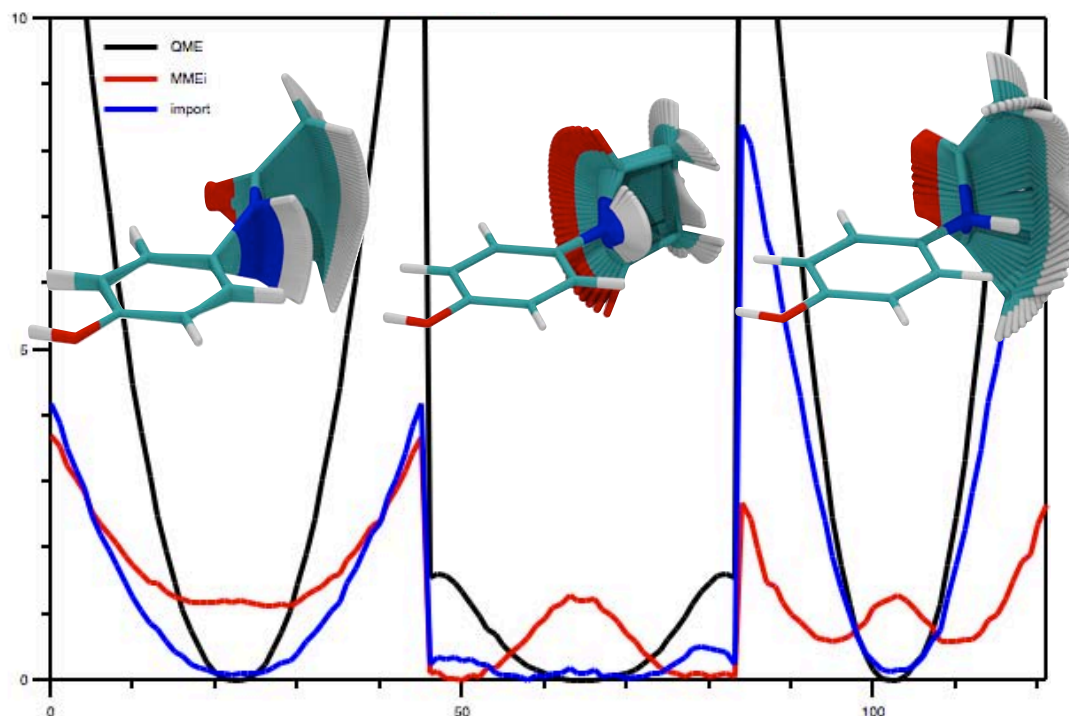
Multidimensional optimization
of developing parameters

Read/Write files (pdb, par, log, gau)



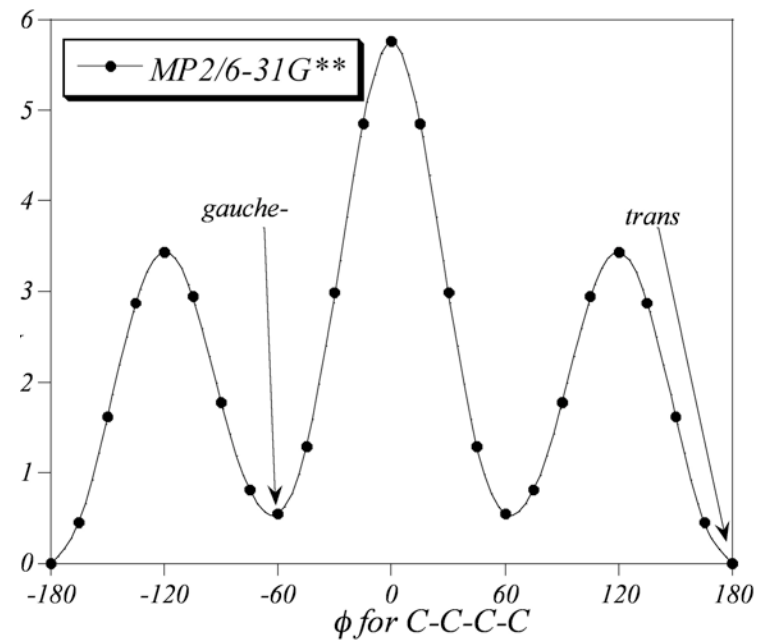
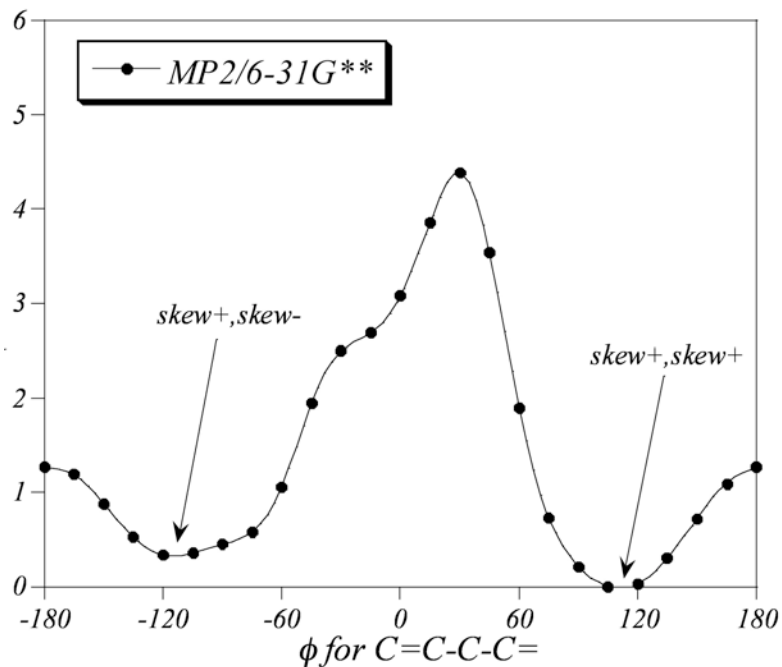
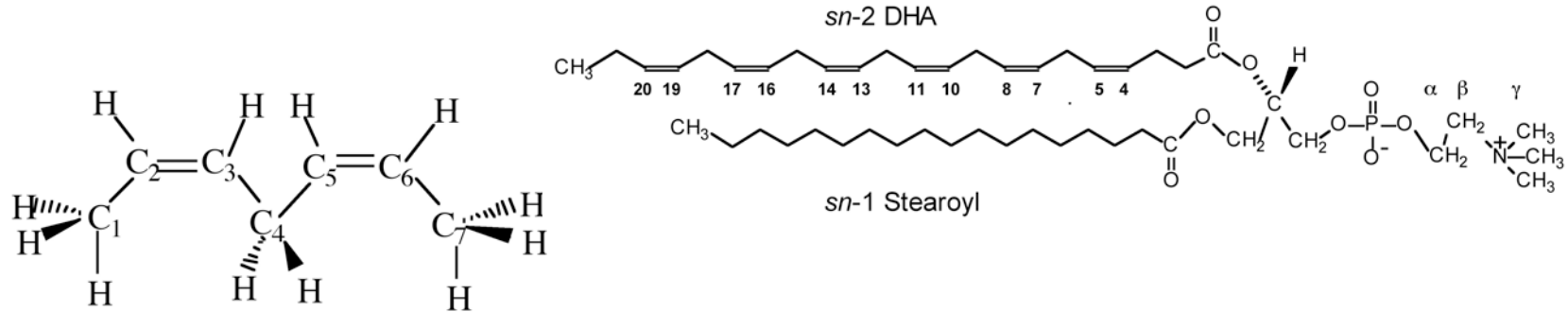
Full documentation online:
<http://www.ks.uiuc.edu/Research/vmd/plugins/fftk/>

Tutorial is under development



Parameterization of unsaturated lipids

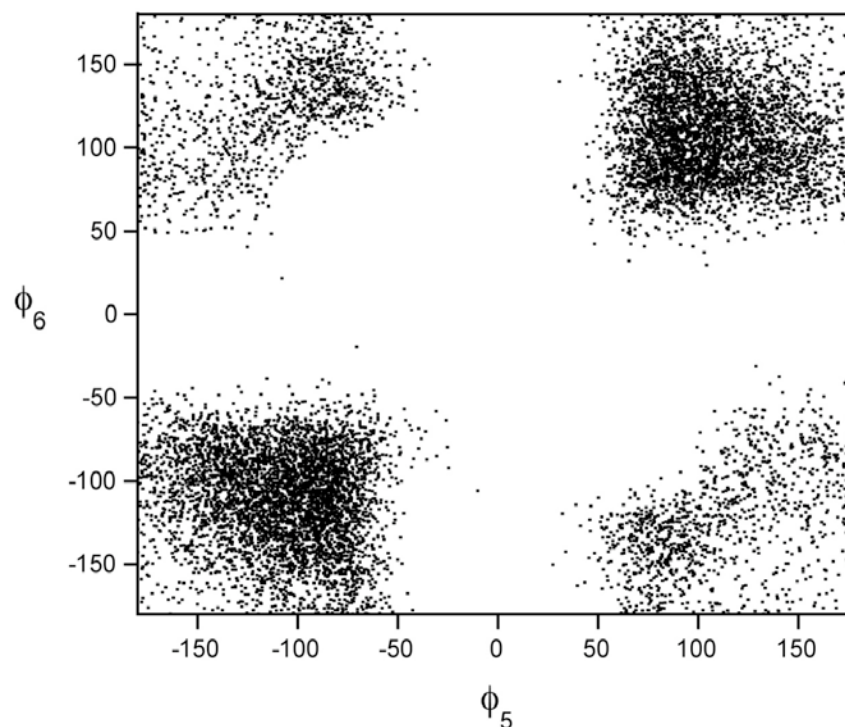
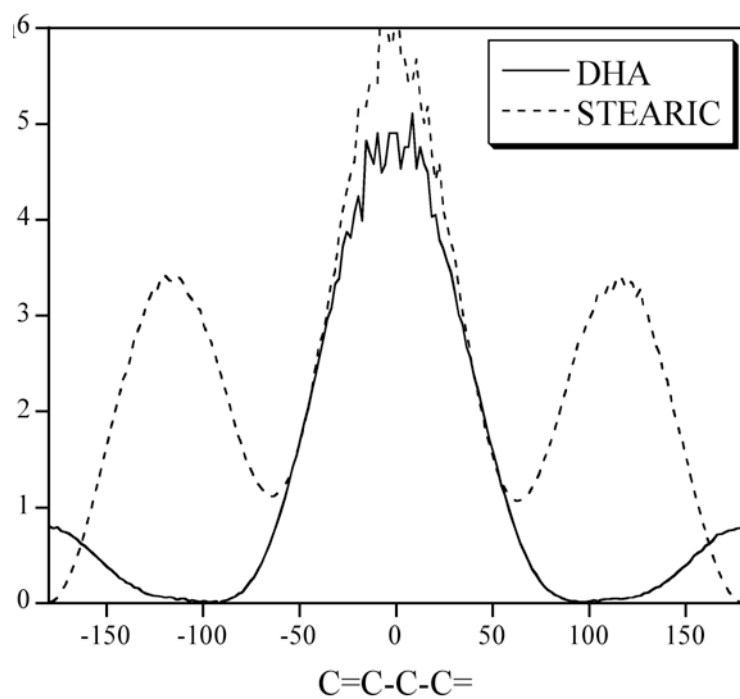
- All C=C bonds are cis, what does rotation about neighboring single bonds look like?



Courtesy of Scott Feller, Wabash College

DHA conformations from MD

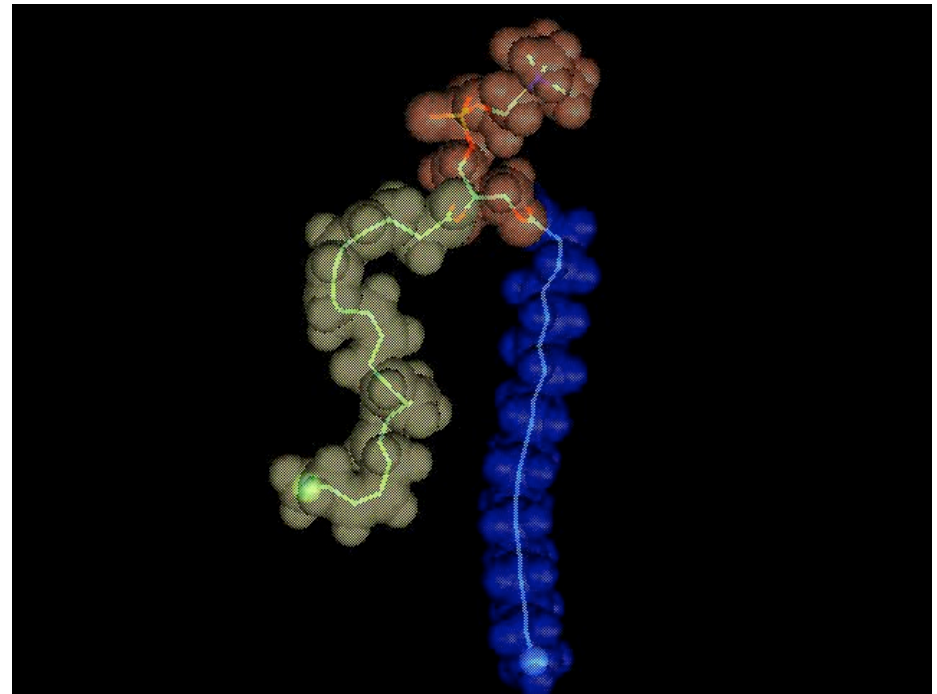
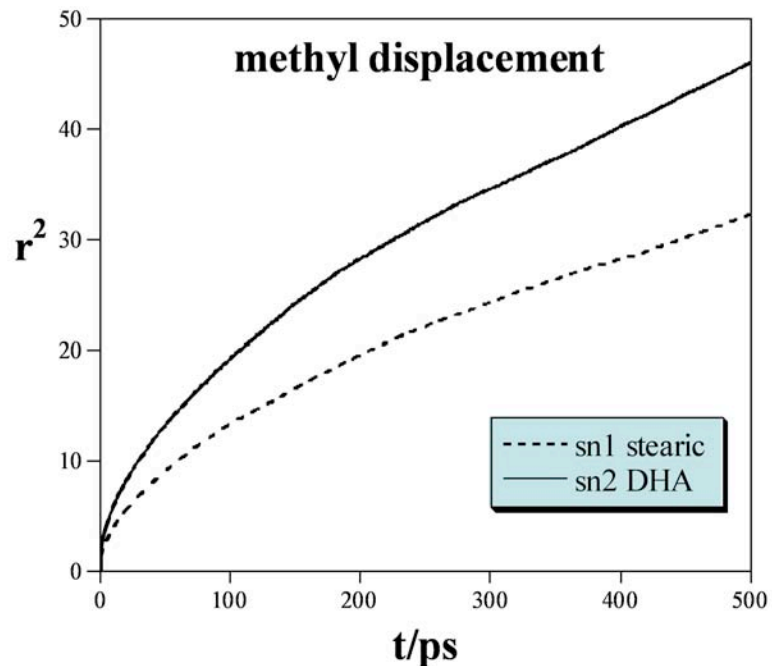
- rotational barriers are extremely small
- many conformers are accessible w/ short lifetimes



Courtesy of Scott Feller, Wabash College

Dynamics of saturated vs. polyunsaturated lipid chains

- *sn*1 stearic acid = blue
- *sn*2 DHA = yellow
- 500 ps of dynamics

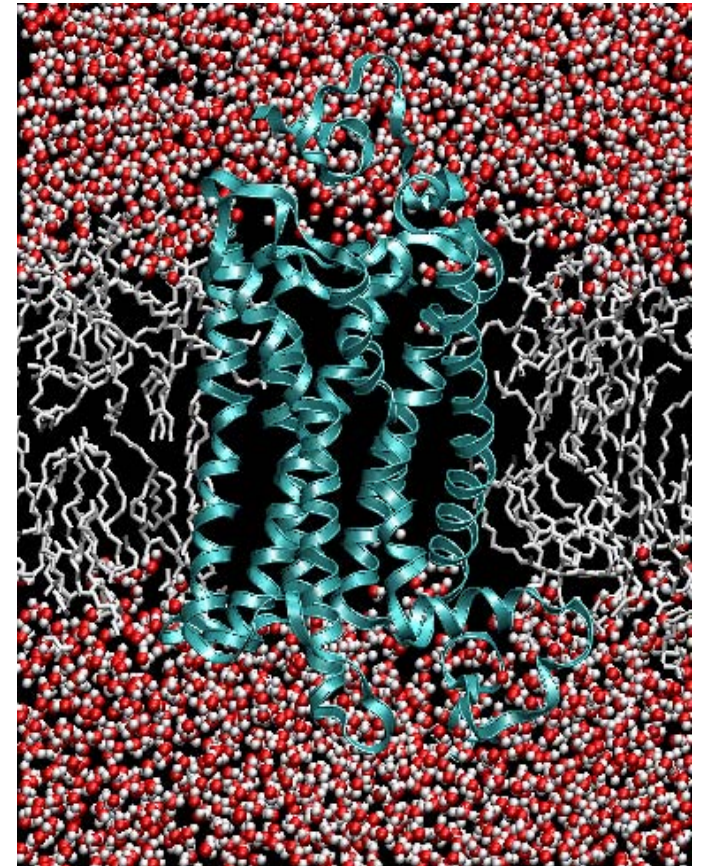
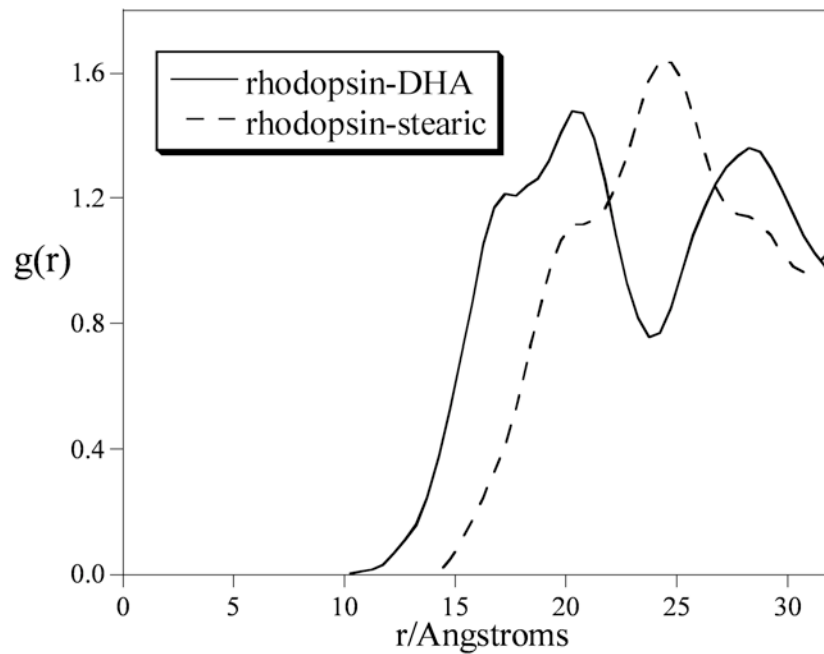


Movie courtesy of Mauricio Carrillo Tripp

Courtesy of Scott Feller, Wabash College

Lipid-protein interactions

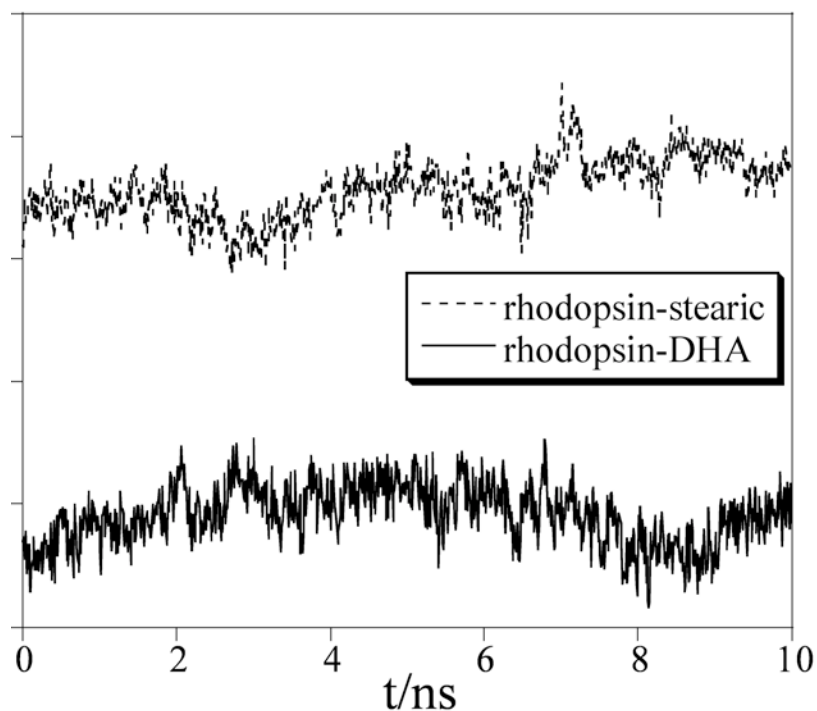
- Radial distribution around protein shows distinct layering of acyl chains



Courtesy of Scott Feller, Wabash College

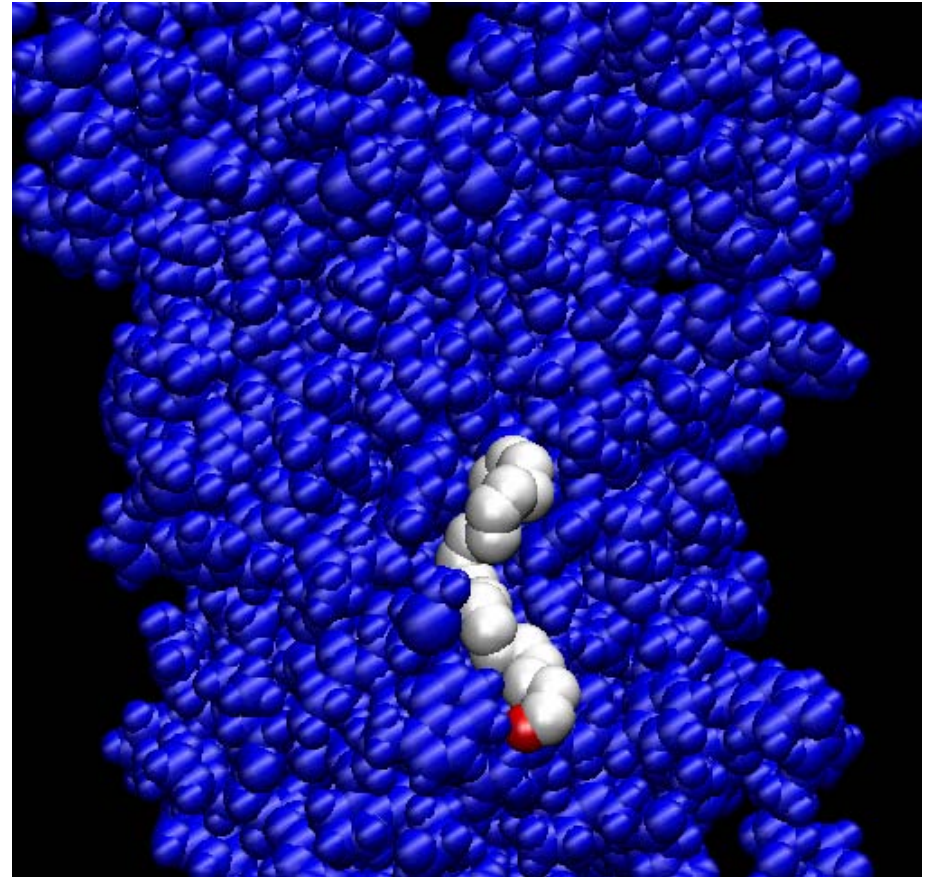
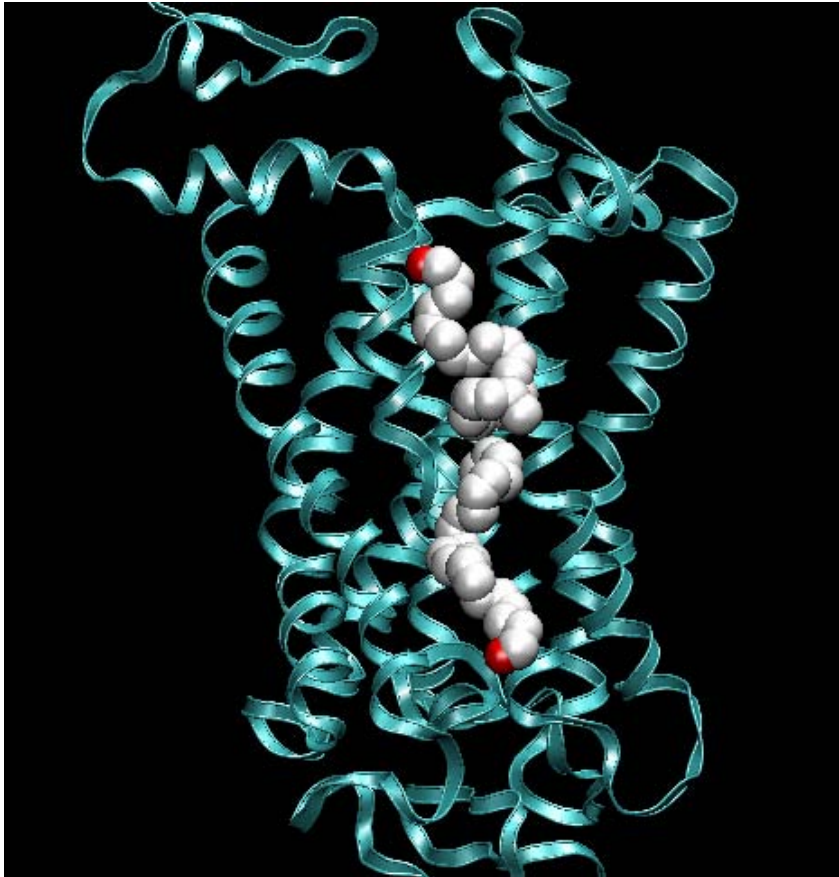
Lipid-protein interactions

- Decomposition of non-bonded interaction shows rhodopsin is strongly attracted to unsaturated chain
- All hydrophobic residues are stabilized by DHA



<u>resname</u>	\underline{U}_{DHA}	$\underline{U}_{stearic}$	<u>ratio</u>
PHE	-44.9	-22.6	2.0
ILE	-30.0	-10.1	3.0
VAL	-24.0	-9.6	2.5
LEU	-23.1	-13.0	1.8
MET	-22.8	-9.7	2.4
TYR	-18.6	-10.4	1.8
ALA	-11.4	-3.0	3.8
TRP	-10.3	-2.4	4.2

Origin of protein:DHA attraction

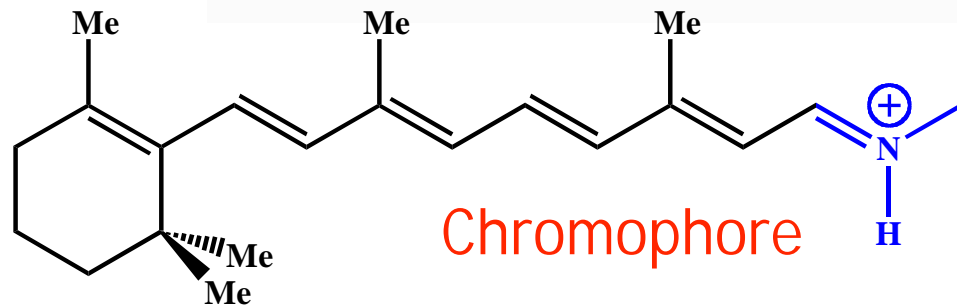
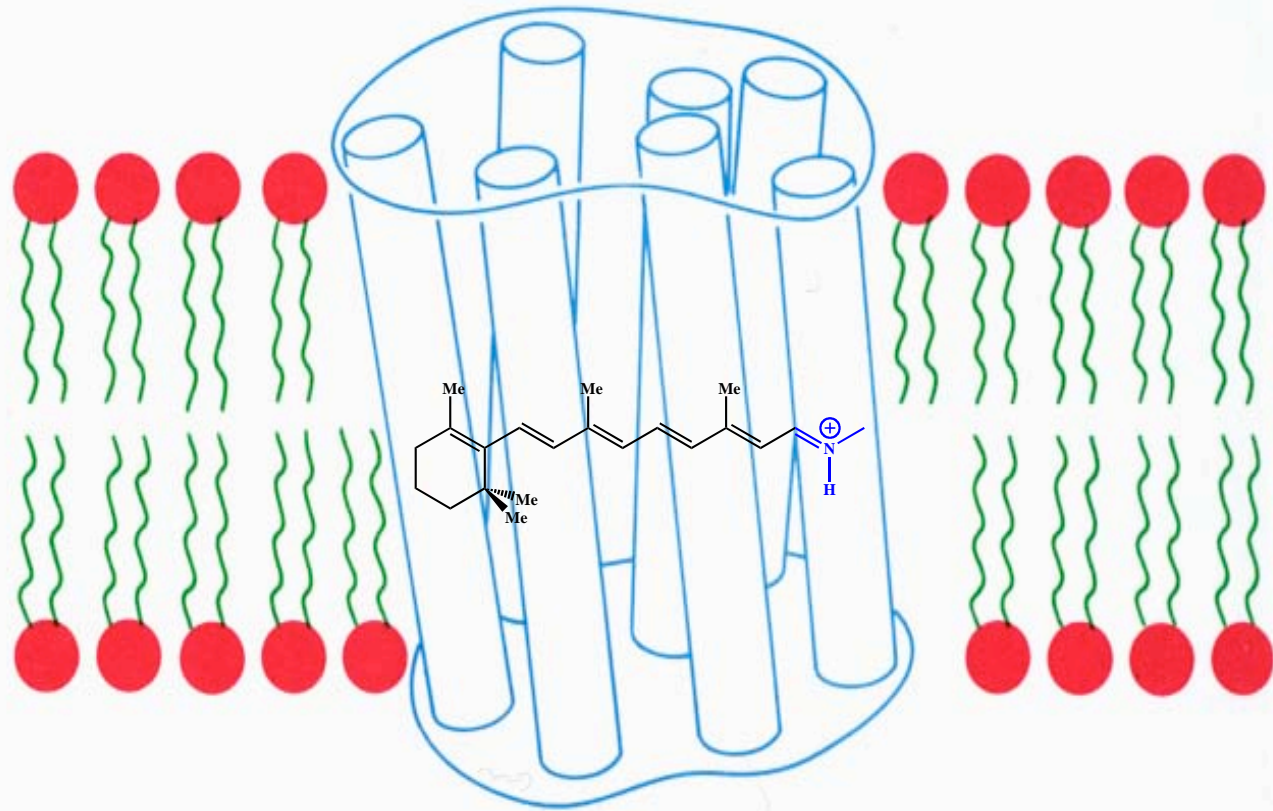


- Flexibility of the DHA chain allows solvation of the rough protein surface to occur with little intra-molecular energy cost

Major Recent Developments

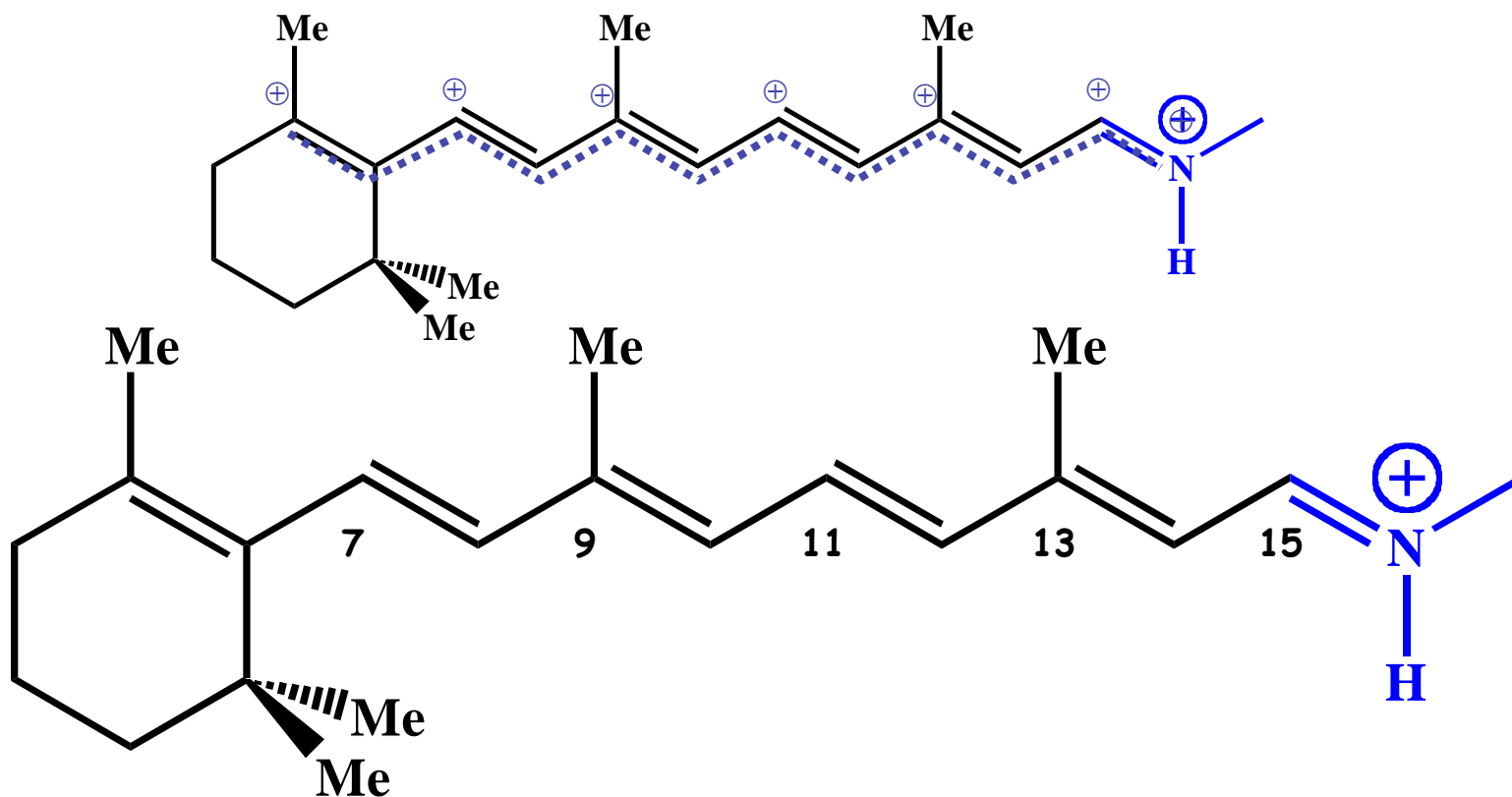
- New set of lipid force field parameters for CHARMM (CHARMM32+)
 - Pastor, B. Brooks, MacKerell
- Polarizable force field
 - Roux, MacKerell

Retinal Proteins -- Rhodopsins

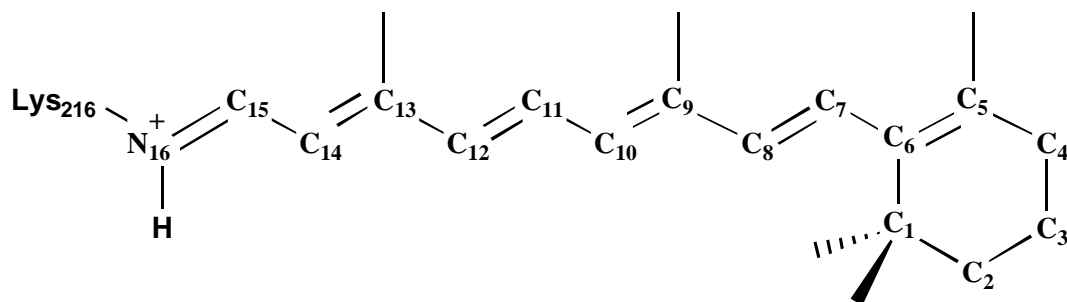


- Covalently linked to a lysine
- Usually protonated **Schiff base**
- all-trans and 11-cis isomers

Unconventional chemistry



Isomerization Barriers in retinal



DFT/6-31G**

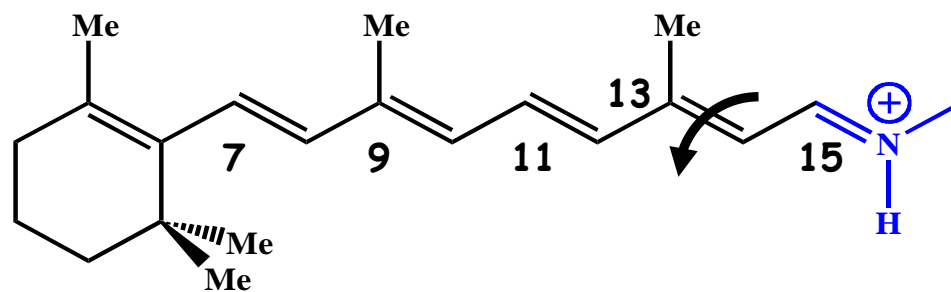
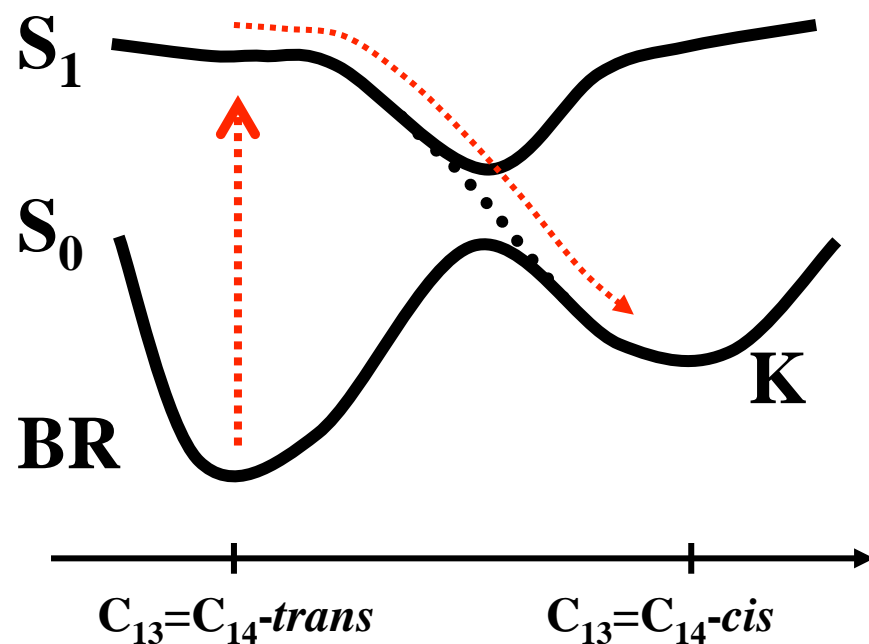
TABLE 2 The parameter set B used for the torsional potentials of the main polyene chain of the retinal Schiff base

ϕ_i	k_i (kcal/mol)*	n_i	δ_i (deg)
$C_5=C_6-C_7=C_8$	11.24	2.0	180.00
$C_6-C_7=C_8-C_9$	39.98	2.0	180.00
$C_7=C_8-C_9=C_{10}$	17.03	2.0	180.00
$C_8-C_9=C_{10}-C_{11}$	37.28	2.0	180.00
$C_9=C_{10}-C_{11}=C_{12}$	22.50	2.0	180.00
$C_{10}-C_{11}=C_{12}-C_{13}$	35.08	2.0	180.00
$C_{11}=C_{12}-C_{13}=C_{14}$	28.30	2.0	180.00
$C_{12}-C_{13}=C_{14}-C_{15}$	29.46	2.0	180.00
$C_{13}=C_{14}-C_{15}=N_{16}$	30.43	2.0	180.00
$C_{14}-C_{15}=N_{16}-C_{\epsilon}$	28.76	2.0	180.00

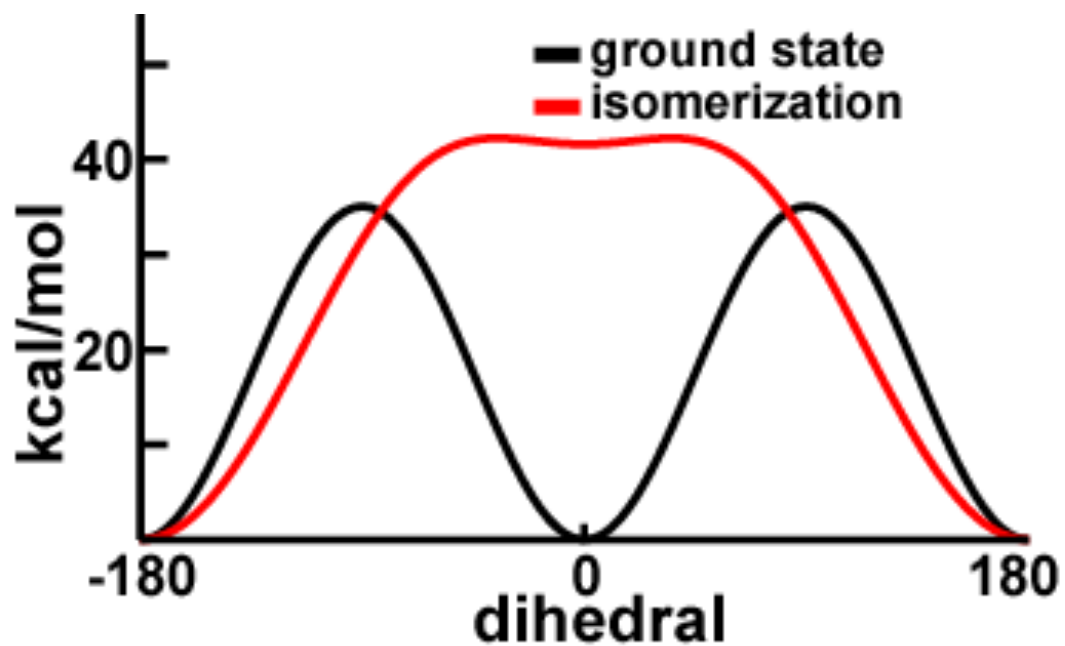
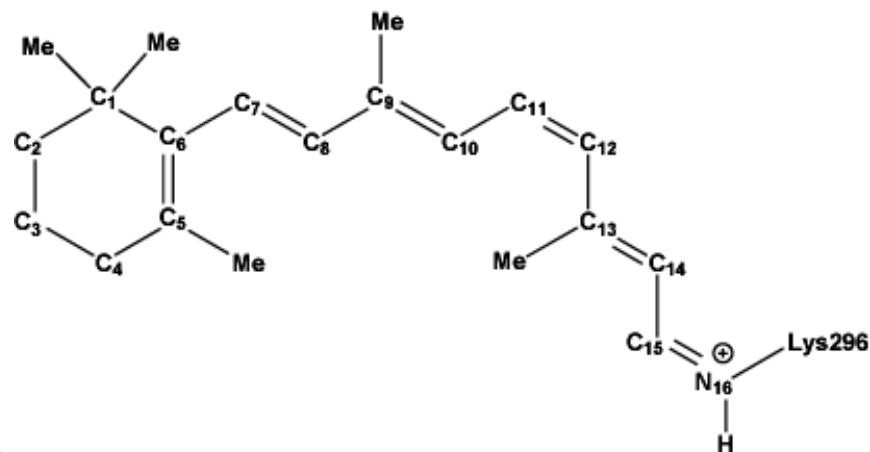
Tajkhorshid et al., 1999.

$$*E_i^{\text{dihedral}} = (1/2)k_i[1 + \cos(n_i\phi_i - \delta_i)].$$

Coupling of electronic excitation and conformational change in bR

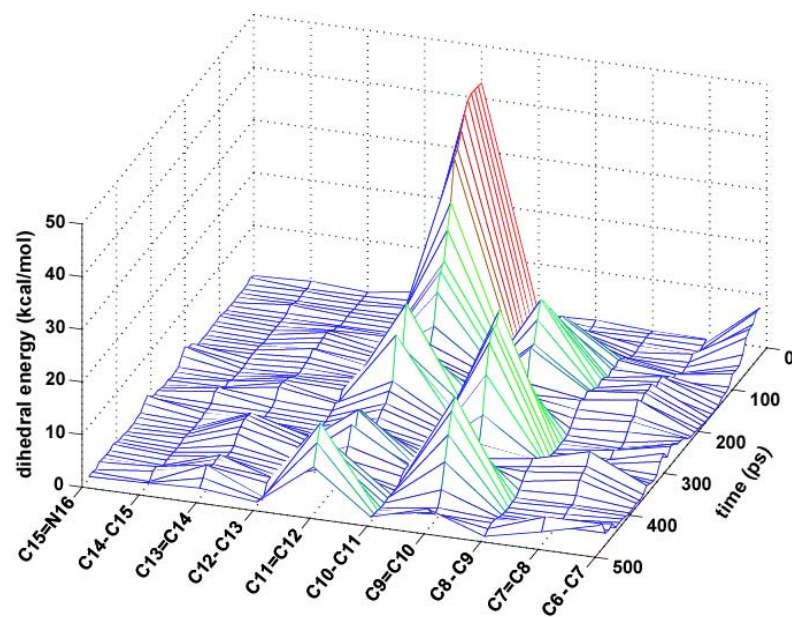
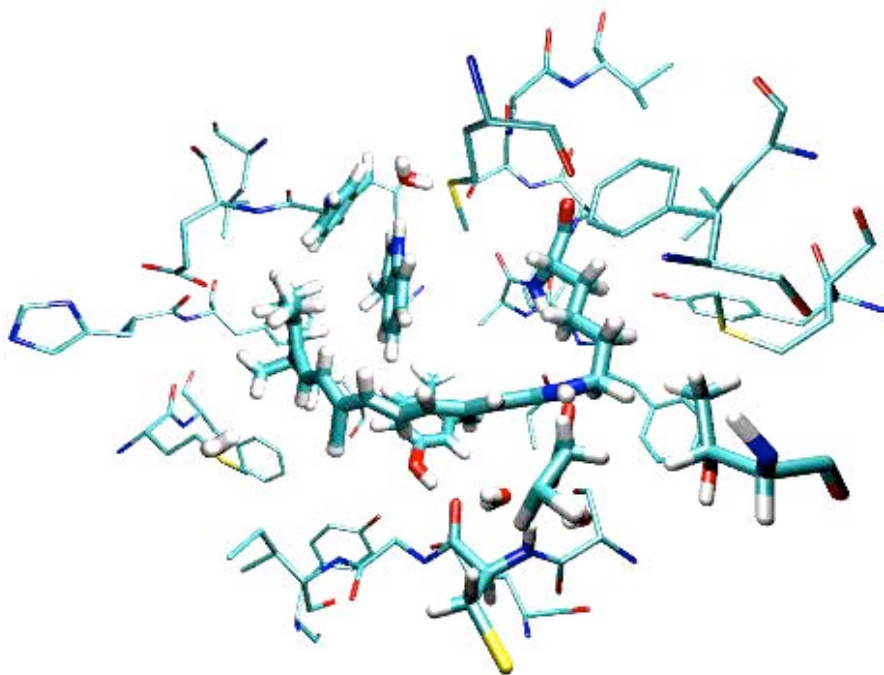


Inducing isomerization



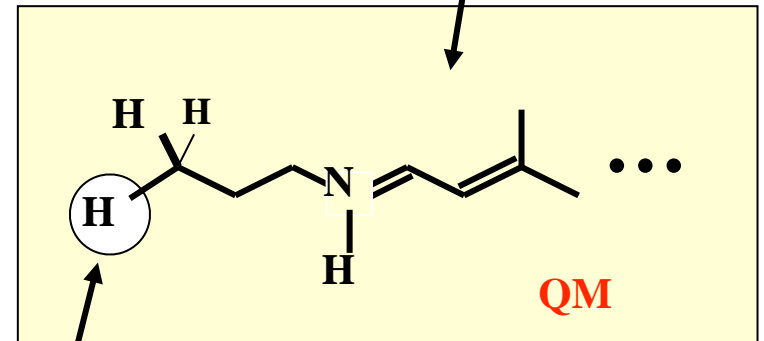
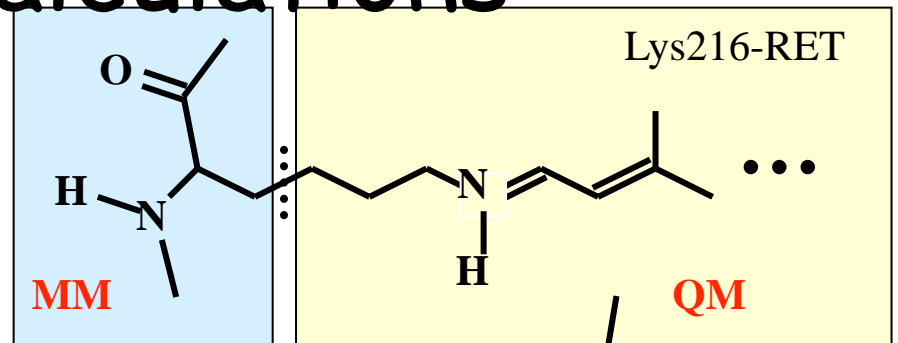
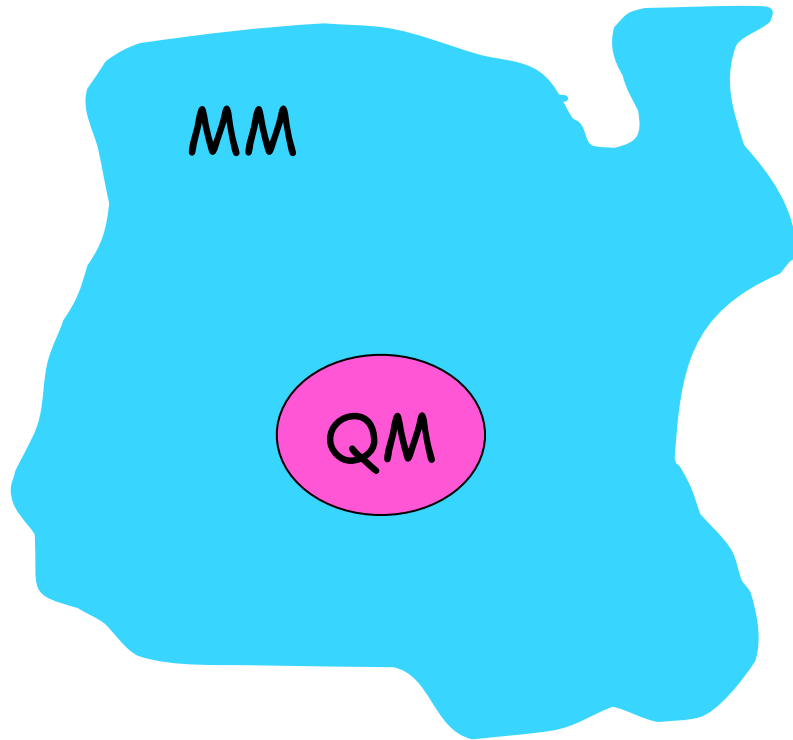
500 nm
~50 kcal/mole

Classical Retinal Isomerization



Twist Propagation

QM/MM calculations

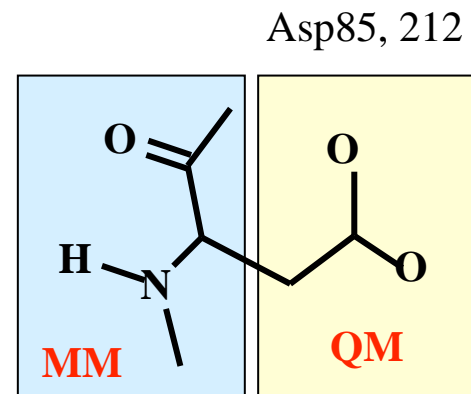


dummy atom

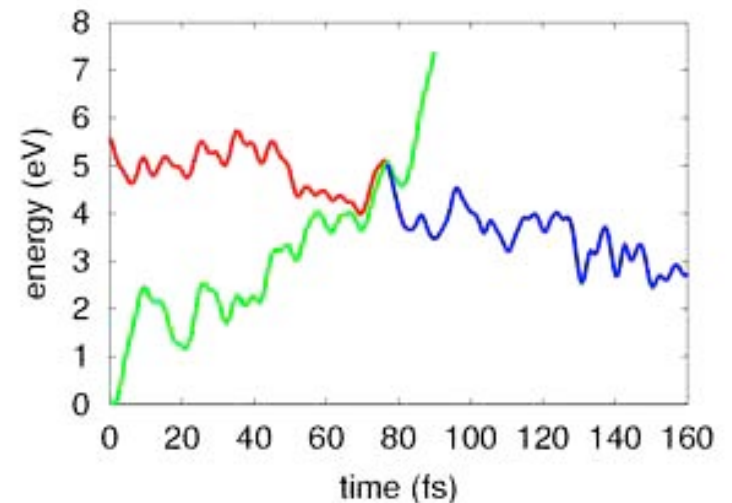
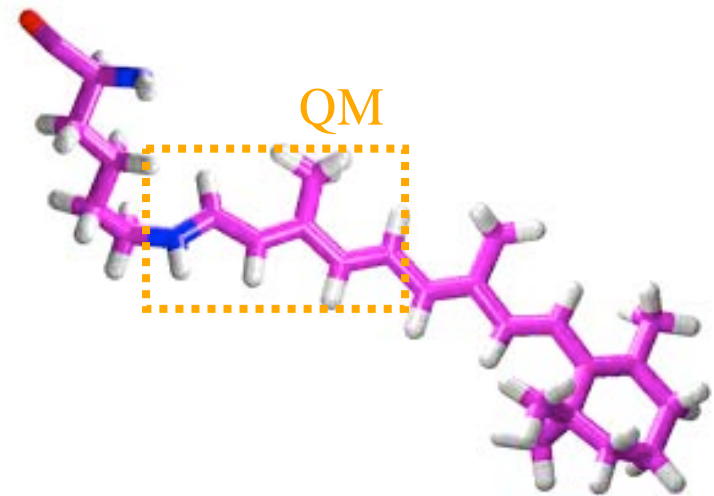
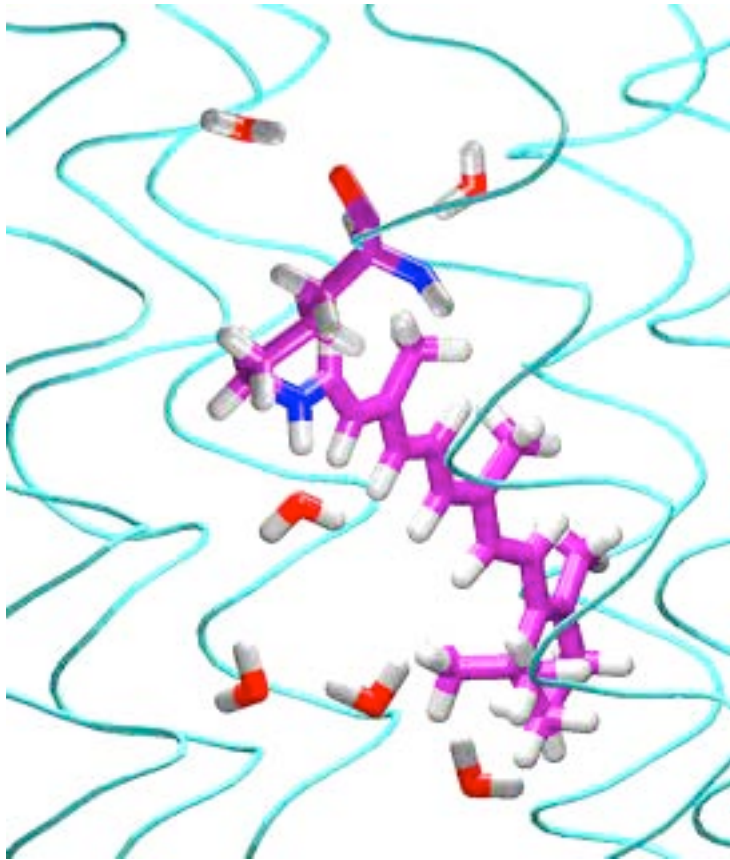
$$\hat{H} = \sum_i \frac{1}{2} p_i^2 + \sum_i \sum_A \frac{Z_A}{r_{iA}} + \sum_{i>j} \frac{1}{r_{ij}} + \sum_{A>B} \frac{Z_A Z_B}{r_{AB}}$$

$$+ \sum_i \sum_p \frac{q_p}{r_{ip}} + \sum_A \sum_p \frac{Z_A q_p}{r_{Ap}}$$

$$+ V_{QM-MM}^{MM} + V_{MM}^{MM}$$

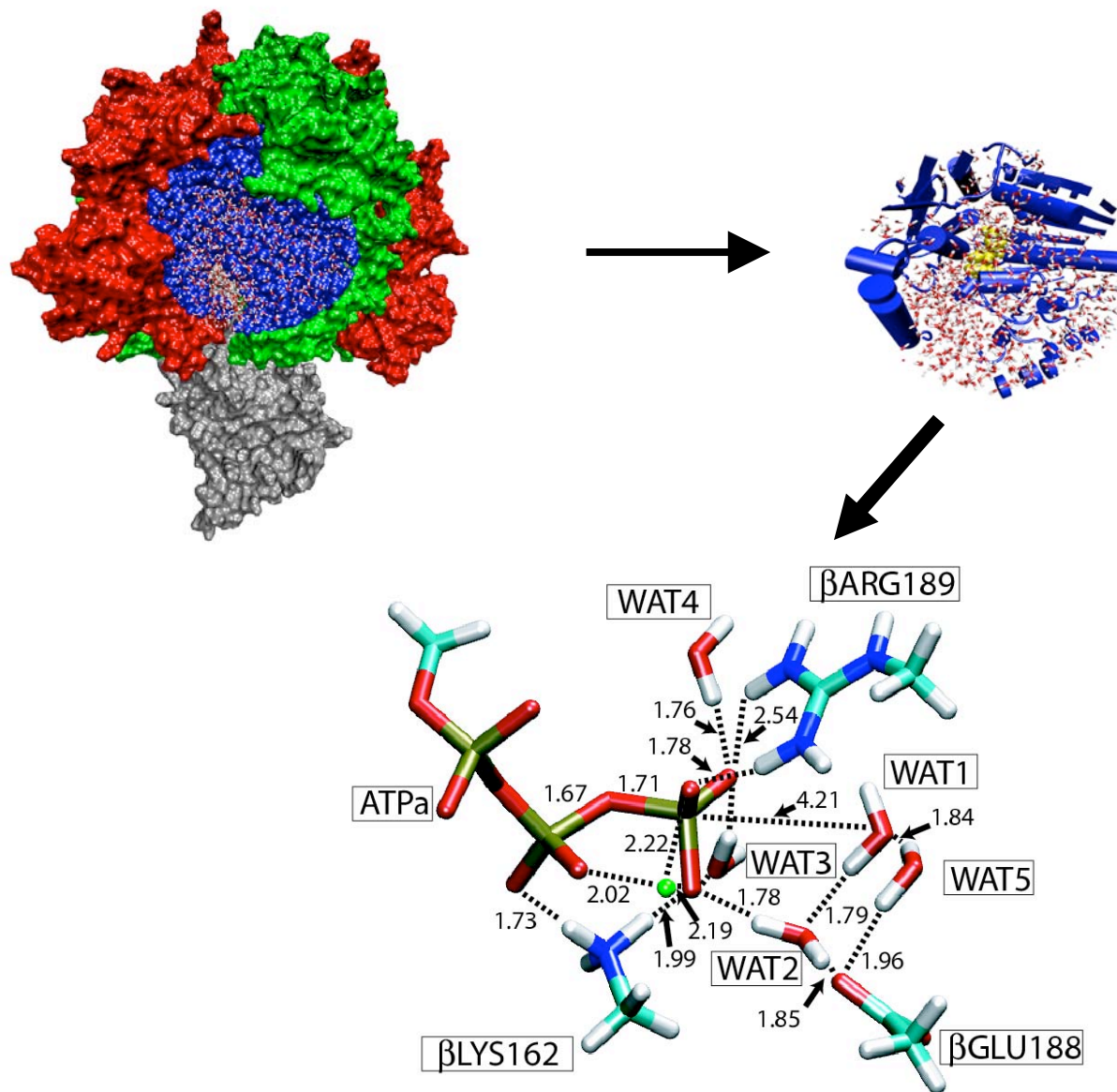


Ab Initio QM/MM Excited State MD Simulation



Quantum mechanical (QM)
treatment of the chromophore,
and force field (MM) treatment of
the embedding protein

QM/MM calculation of ATP hydrolysis



Coarse grain modeling of lipids

150 particles



9 particles!

(A)

