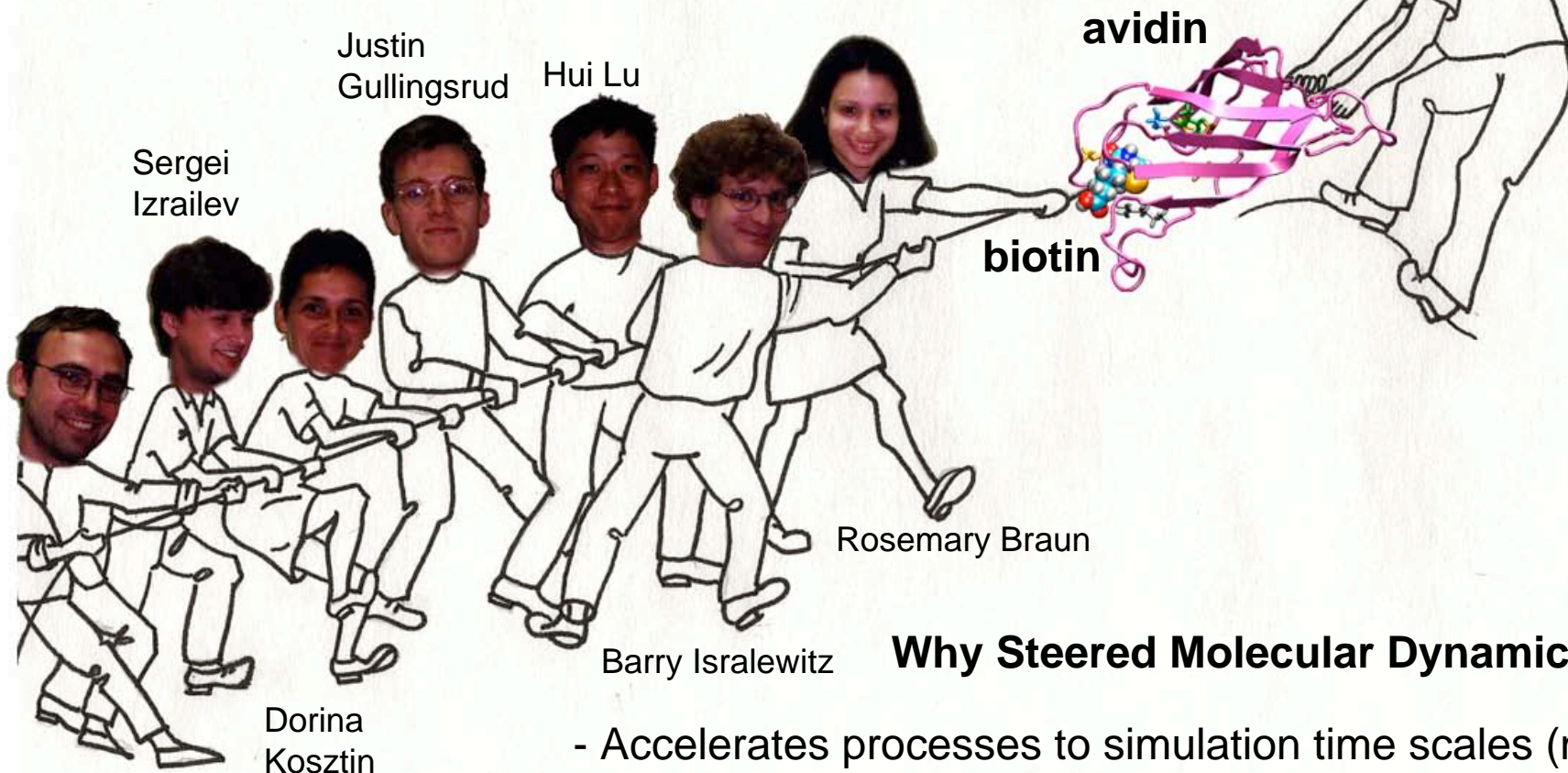


# Steered Molecular Dynamics Introduction and Examples

Klaus  
Schulten



## Why Steered Molecular Dynamics?

- Accelerates processes to simulation time scales (ns)
- Yields explanations of biopolymer mechanics
- Complements Atomic Force Microscopy
- Finds underlying unbinding potentials
- Generates and tests Hypotheses

Acknowledgements:

Fernandez group, Mayo C.; Vogel group, U. Washington  
NIH, NSF, Carver Trust

# Mechanical Functions of Proteins

## Forces

naturally arise in cells

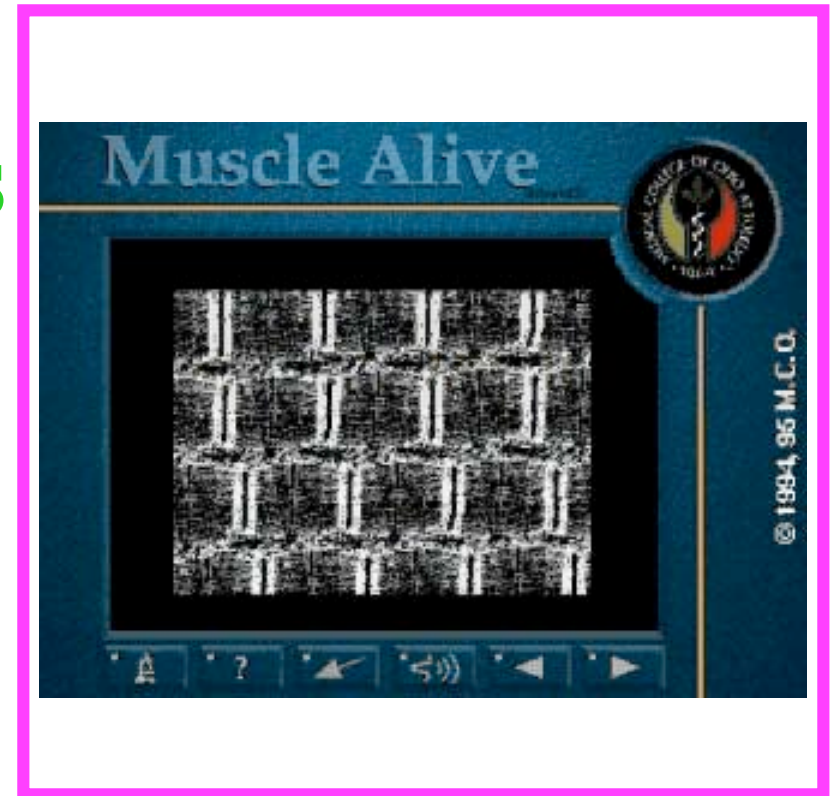
and can also be

substrates (ATPase)

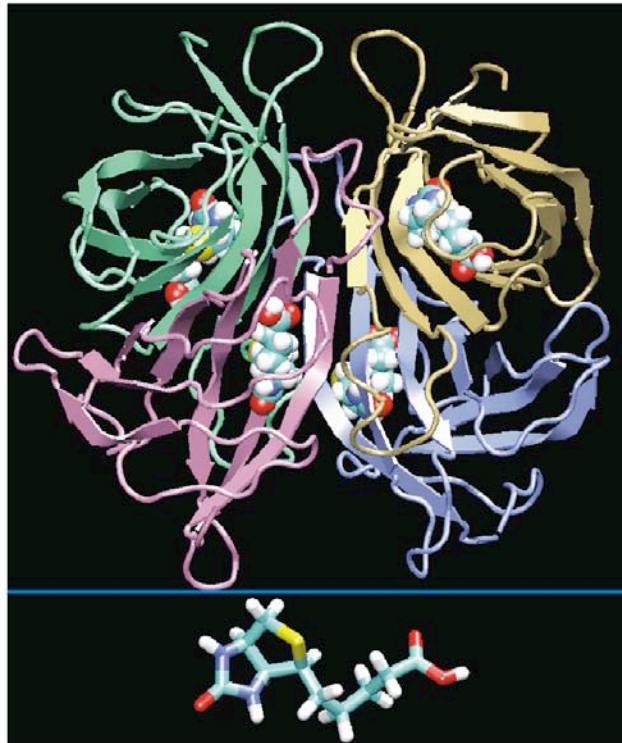
products (myosin)

signals (integrin)

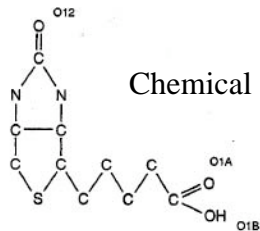
of cellular processes



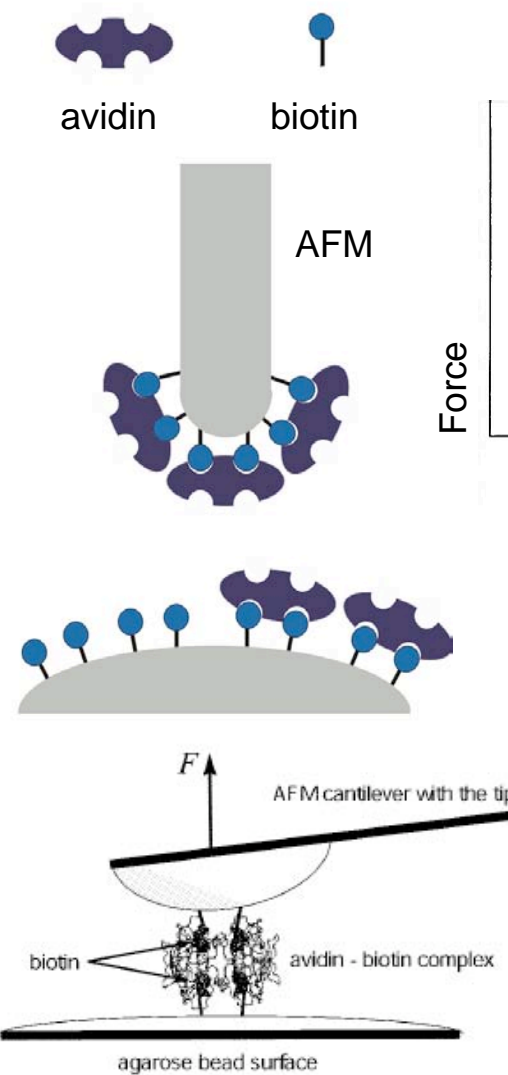
# Atomic Force Microscopy Experiments of Ligand Unbinding



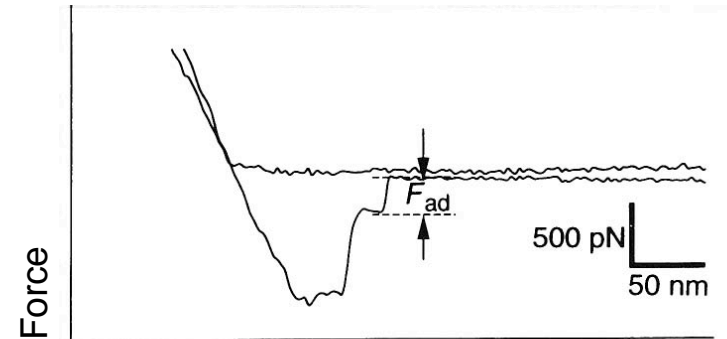
**Biotin**



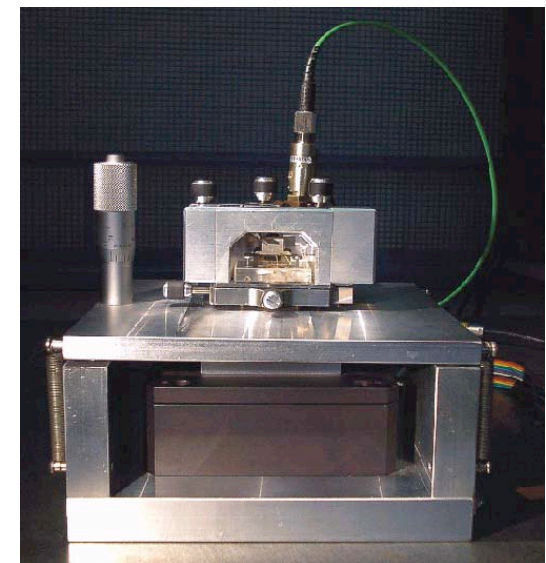
Chemical structure of biotin



Florin et al., Science 264:415 (1994)

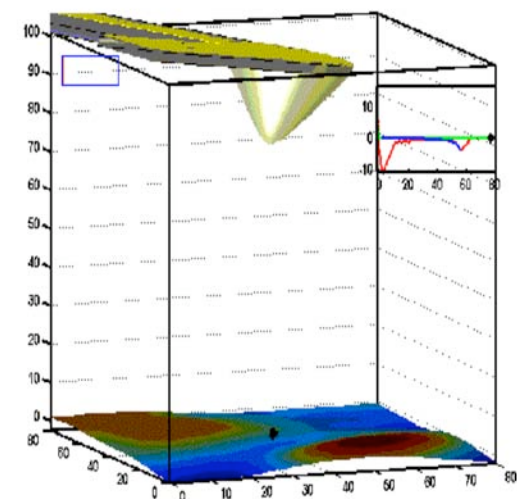
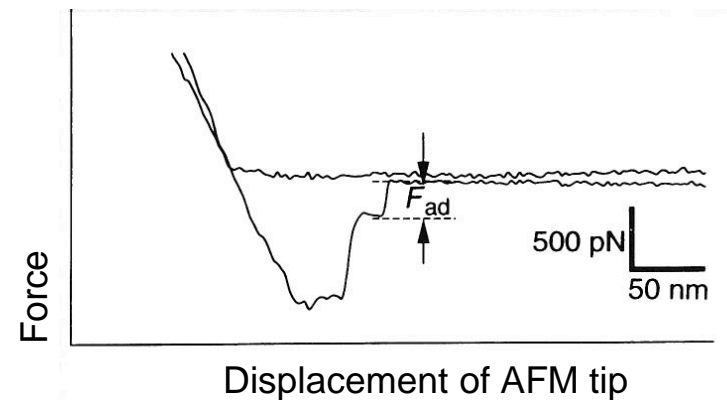
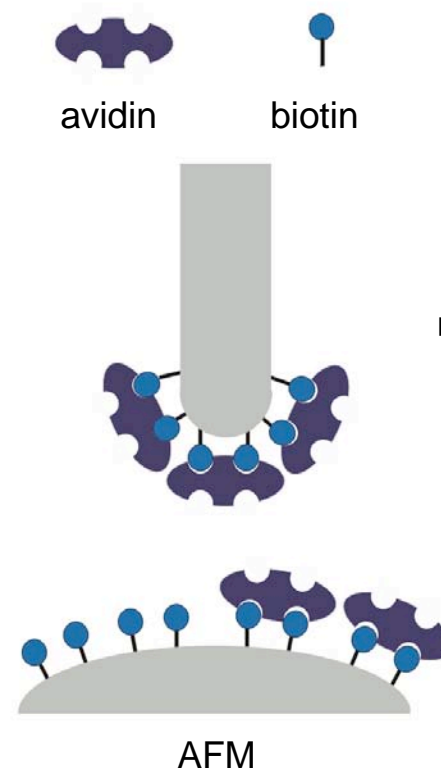
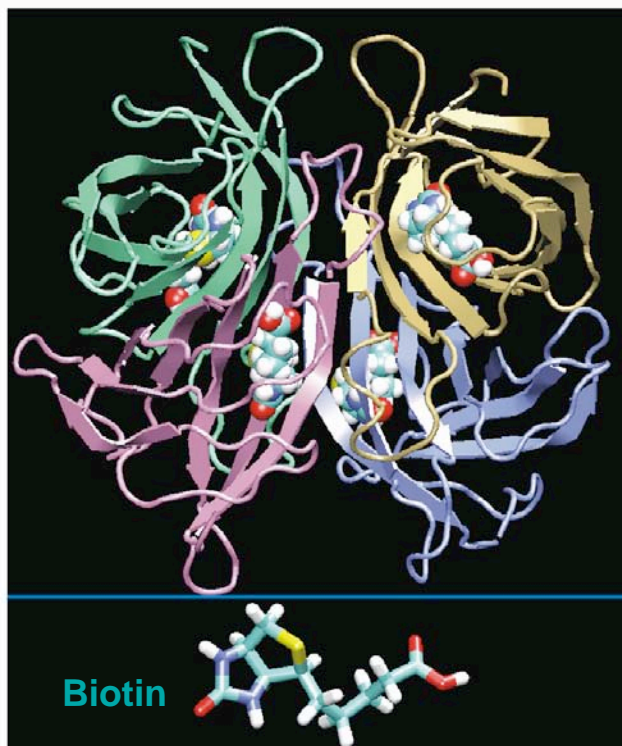


Displacement of AFM tip



# Atomic Force Microscopy Experiments of Ligand Unbinding

Florin et al., Science 264:415 (1994)



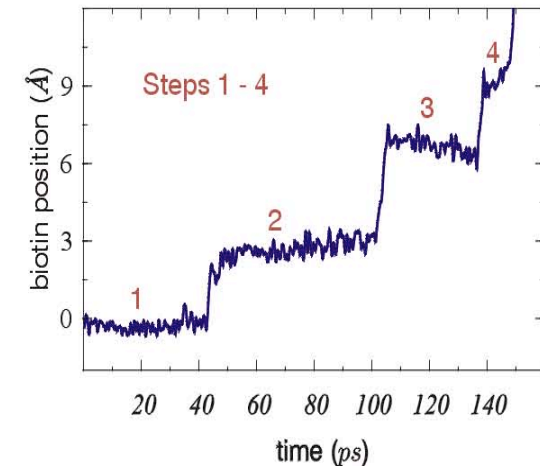
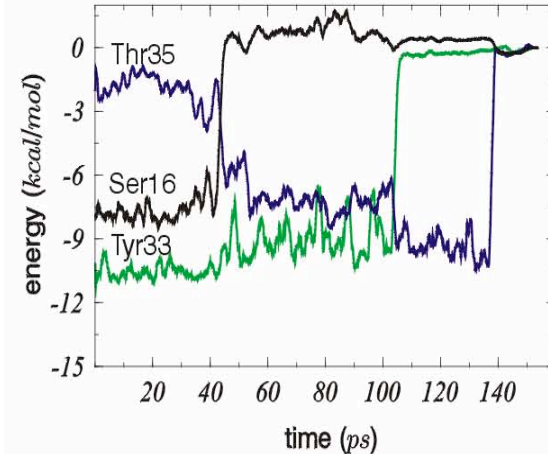
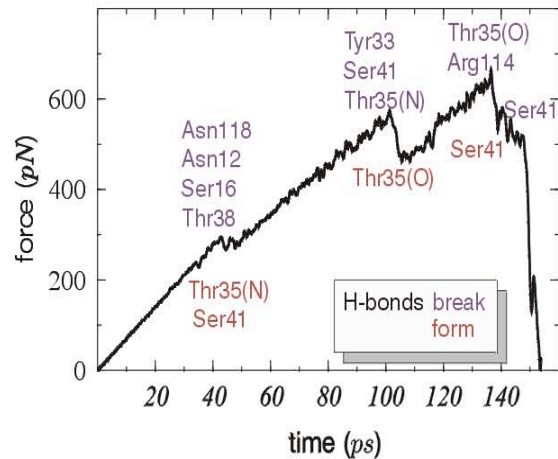
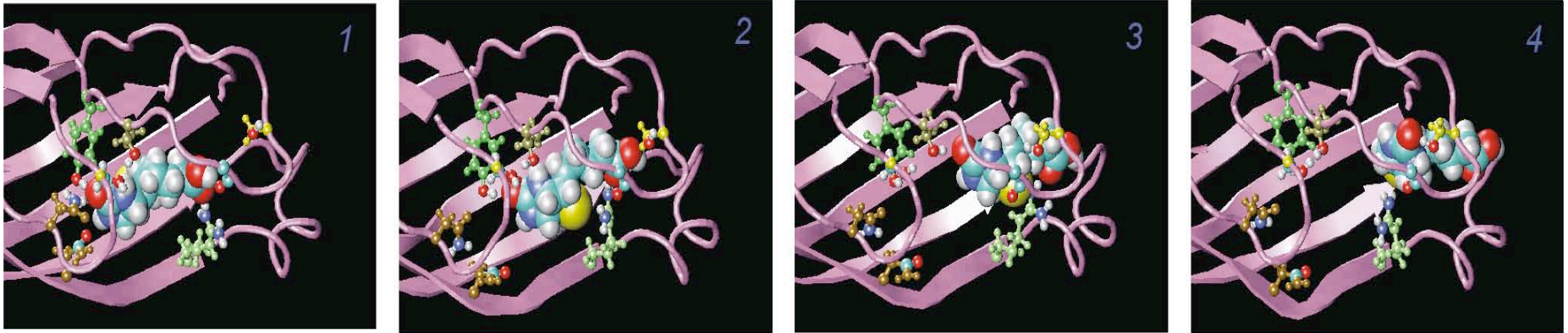
# Pulling Biotin out of Avidin



Molecular dynamics study of unbinding of the avidin-biotin complex. Sergei Izrailev, Sergey Stepaniants, Manel Balsera, Yoshi Oono, and Klaus Schulten. *Biophysical Journal*, 72:1568-1581, 1997.

# SMD of Biotin Unbinding: What We Learned

biotin slips out in steps, guided by amino acid side groups, water molecules act as lubricant, MD overestimates extrusion force



<http://www.ks.uiuc.edu>

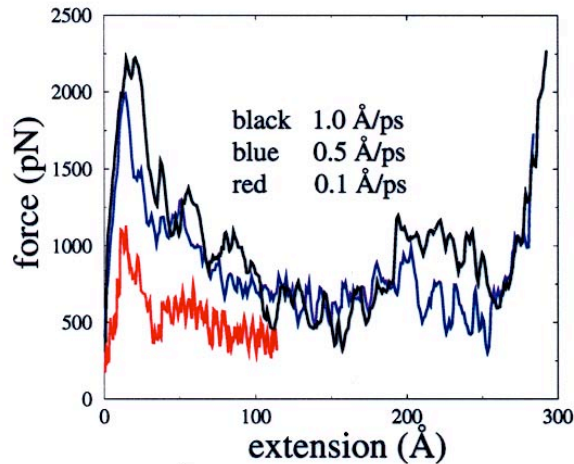
Israilev *et al.*, *Biophys. J.*, **72**, 1568-1581 (1997)

NIH Resource for Macromolecular Modeling and Bioinformatics  
Theoretical Biophysics Group, Beckman Institute, UIUC

# Quantitative Comparison

Bridging the gap between SMD and AFM experiments

Force-extension curve



Schematic potentials

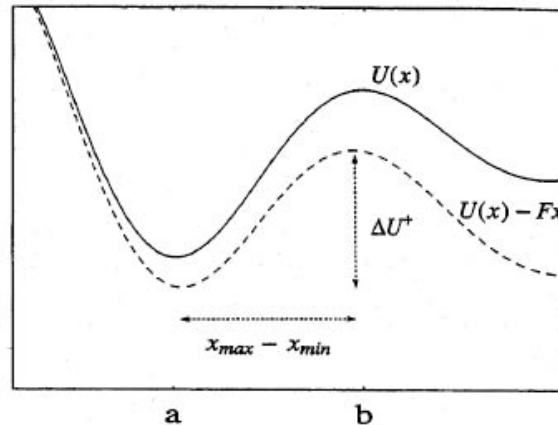


FIGURE 9 Schematic potentials  $U(x)$ , and  $U(x) - Fx$ .

$$\delta(F) = \beta [\Delta U - F(b-a)]$$

AFM regime

$$e^{\delta(F)} \gg 1$$

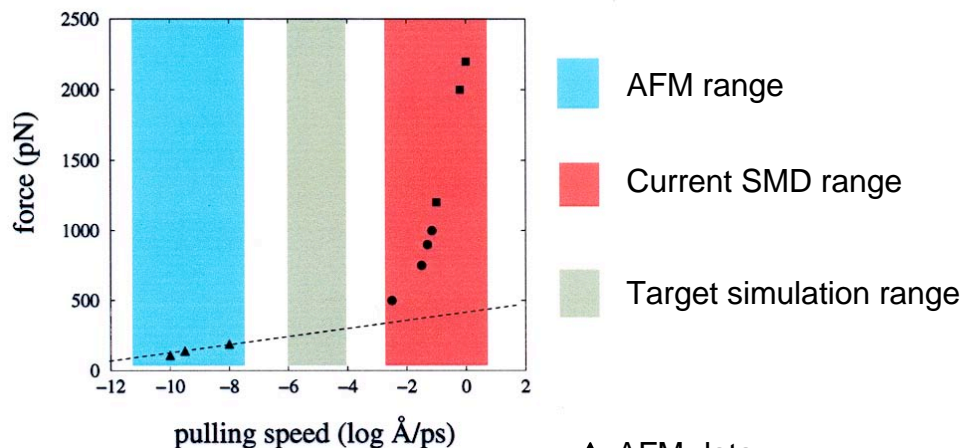
$$\tau_{AFM} \sim 2\tau_D \delta^{-2}(F) e^{\delta(F)}$$

SMD regime

$$e^{\delta(F)} \ll 1$$

$$\tau_{SMD} \sim 2\tau_D / |\delta(F)|^{-1}$$

Force-pulling velocity relationship



■

SMD data

●

▲ AFM data

----- Extrapolation of AFM data

# Rupture/Unfolding Force $F_0$ and its Distribution

$$\tau(F_0) = 1 \text{ ms} \quad \text{time of measurement}$$

$$\Rightarrow F_0 \quad \text{rupture/unfolding force}$$

Distribution of rupture/unfolding force

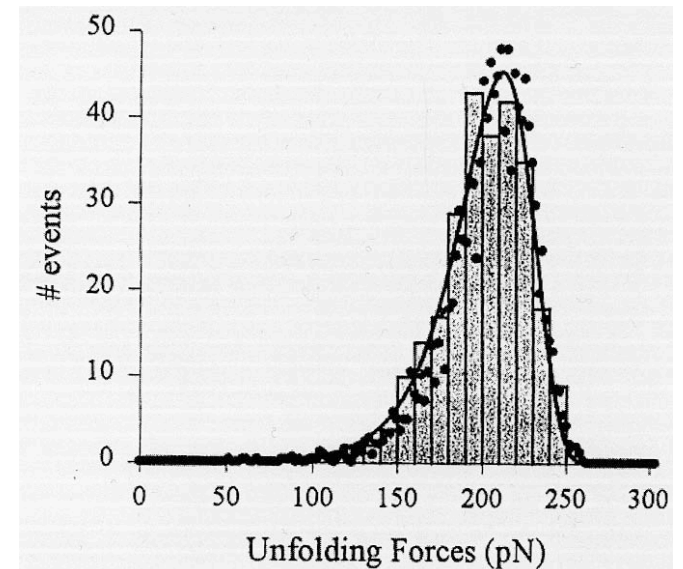
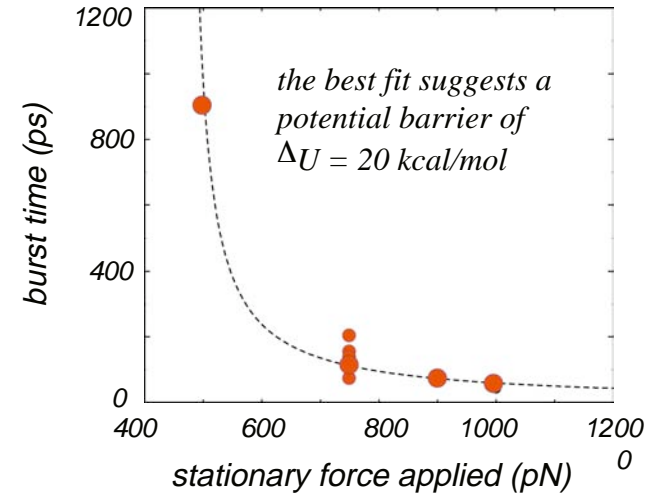
$$p(F_0) = \kappa \exp[\beta F_0(b - a) - \beta \Delta U - \frac{\kappa k_B T}{b - a} e^{-\beta \Delta U} (e^{\beta F_0(b - a)} - 1)]$$

$$\kappa = \delta^2(F)/2\tau_D k v$$

Israilev *et al.*, *Biophys. J.*, **72**, 1568-1581 (1997)

Balsera *et al.*, *Biophys. J.*, **73**, 1281-1287 (1997)

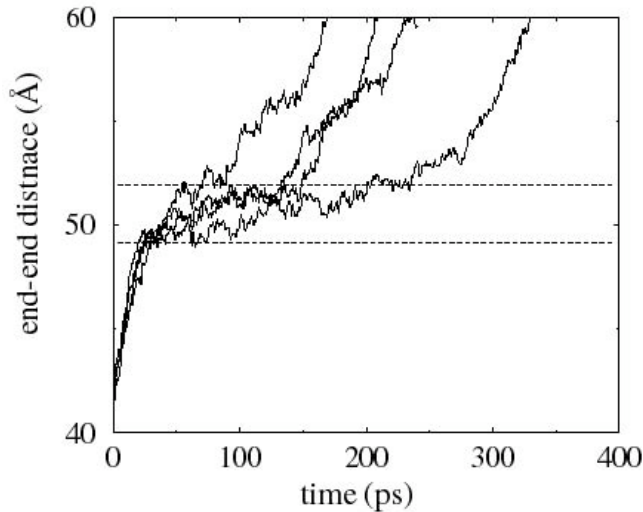
determination of barrier height based on mean first passage time



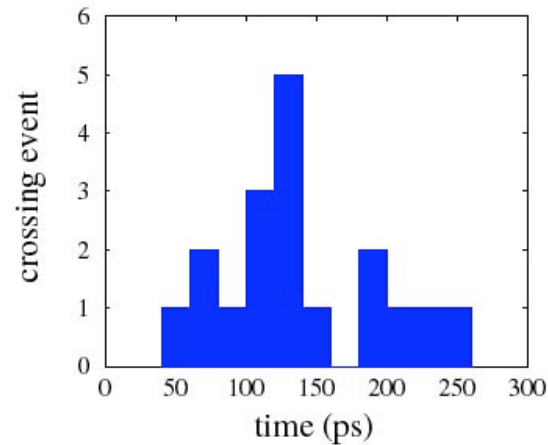


# Distribution of the Barrier Crossing Time

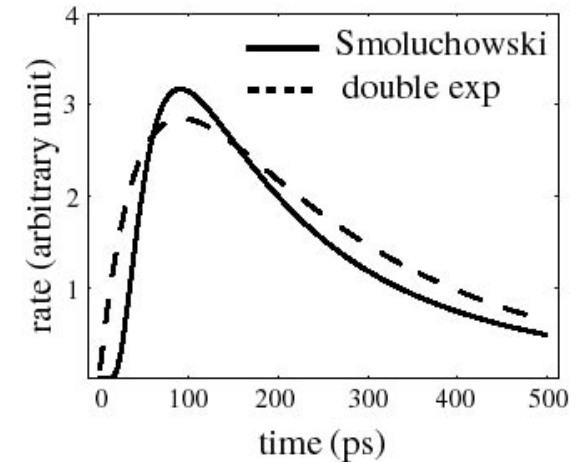
Multiple runs with same force of 750 pN



Barrier crossing times of 18 SMD simulations



Theoretical prediction of the barrier crossing times



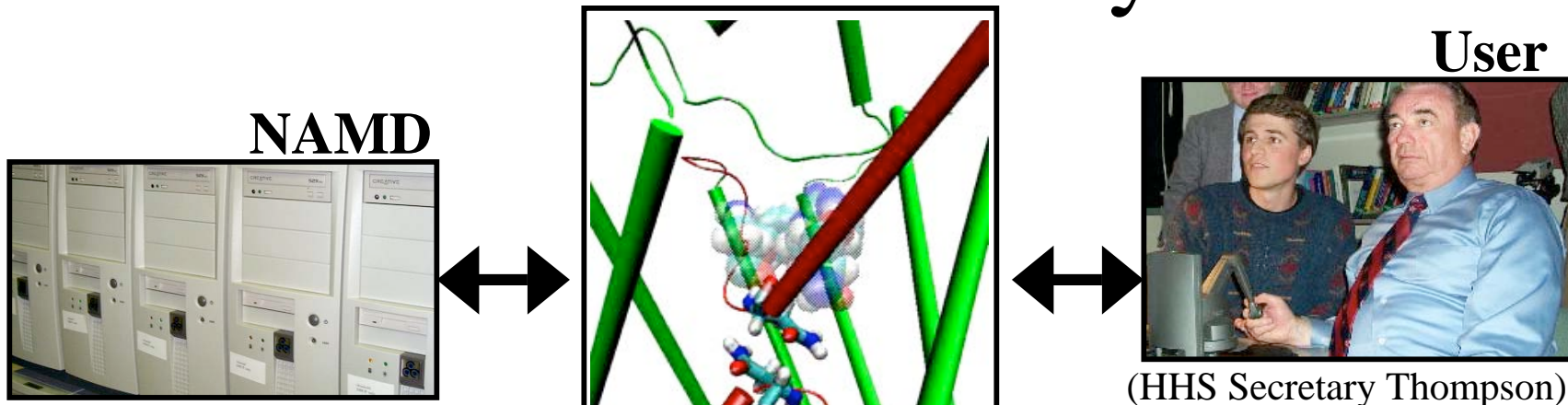
The fraction  $N(t)$  that has not crossed the barrier can be expressed through solving the Smoluchowski diffusion equation (linear model potential):

$$N(t) = \frac{1}{2} \operatorname{erfc} \left[ \frac{-a + \delta(F)Dt/(b-a)}{\sqrt{4Dt}} \right] - \frac{1}{2} \exp \left[ \frac{\delta(F)a}{b-a} \right] \operatorname{erfc} \left[ \frac{-a + \delta(F)Dt/(b-a)}{\sqrt{4Dt}} \right]$$

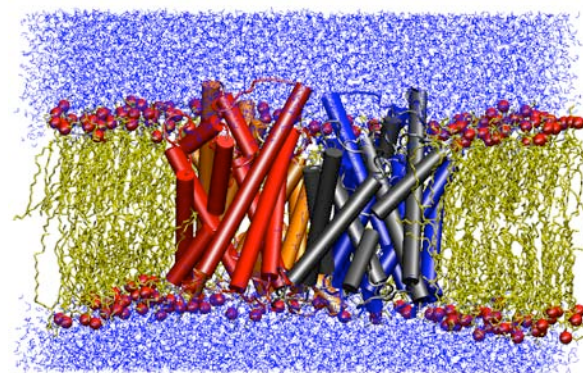
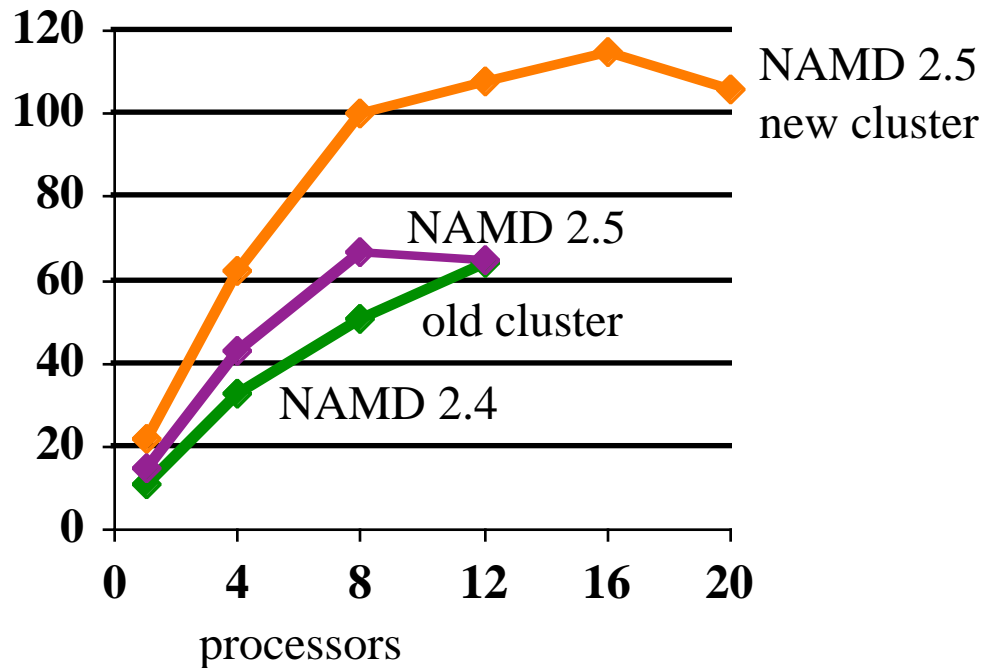
Or approximated by double exponential (general potential):

$$N(t) = [t_1 \exp(-t/t_1) - t_2 \exp(-t/t_2)] / (t_1 - t_2), \text{ Nadler \& Schulten, JCP., 82, 151-160 (1985)}$$

# Interactive Molecular Dynamics



steps per second (more is better)



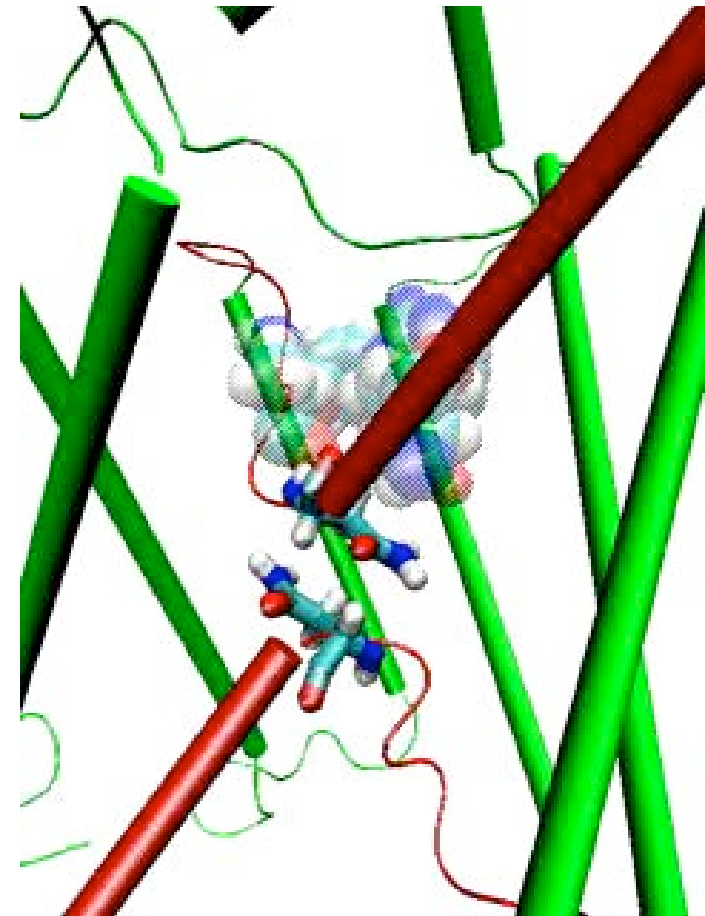
GlpF IMD Benchmark:

- 4210 atoms
- 3295 fixed atoms
- 10Å cutoff, no PME
- Limited by network latency

# Interactive Molecular Dynamics

VMD ←·····→ NAMD

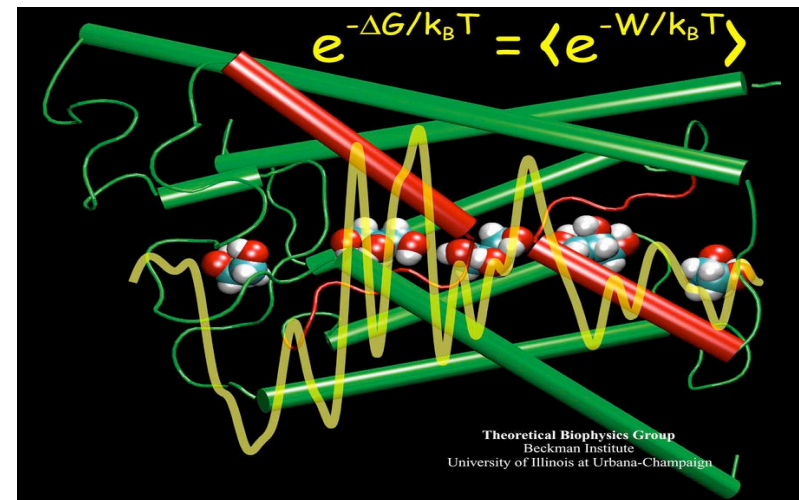
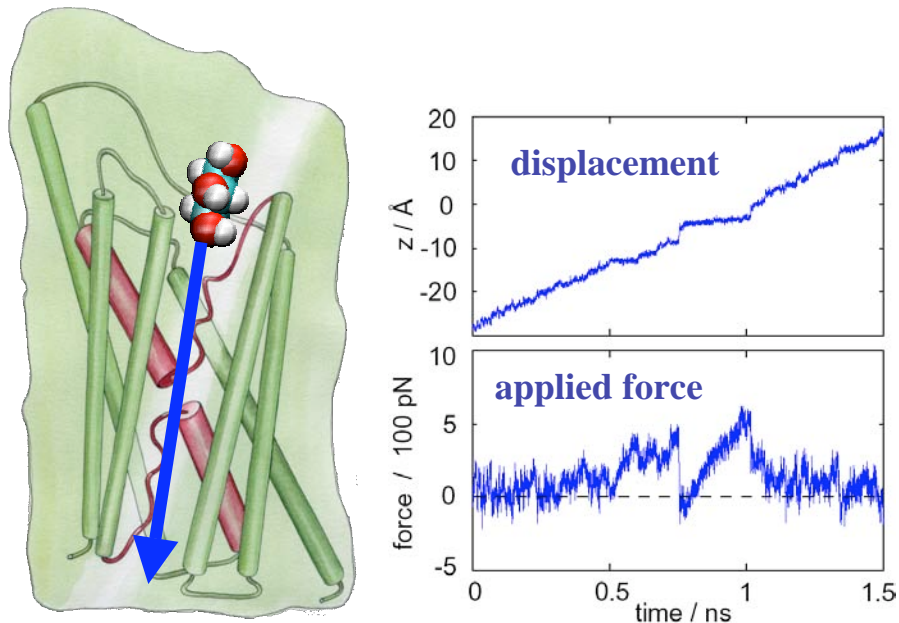
- Any PC/Workstation
- Supports 3D force-feedback devices for interaction



**J. Stone, J. Gullingsrud, K. Schulten, and P. Grayson.**  
*A System for Interactive Molecular Dynamics Simulation.*  
2001 ACM Symposium on Interactive 3D Graphics,  
pp.191-194, ACM SIGGRAPH

**P. Grayson, E. Tajkhorshid, and K. Schulten.**  
*Biophysical J*, **83**: 36 (2003)

# Quantitative Analysis of Substrate Permeation



Jensen et al, *PNAS* **99**: 6731-6736 (2002)

Calculation of the free energy profile of sugar transport from SMD simulations by Jarzynski's identity

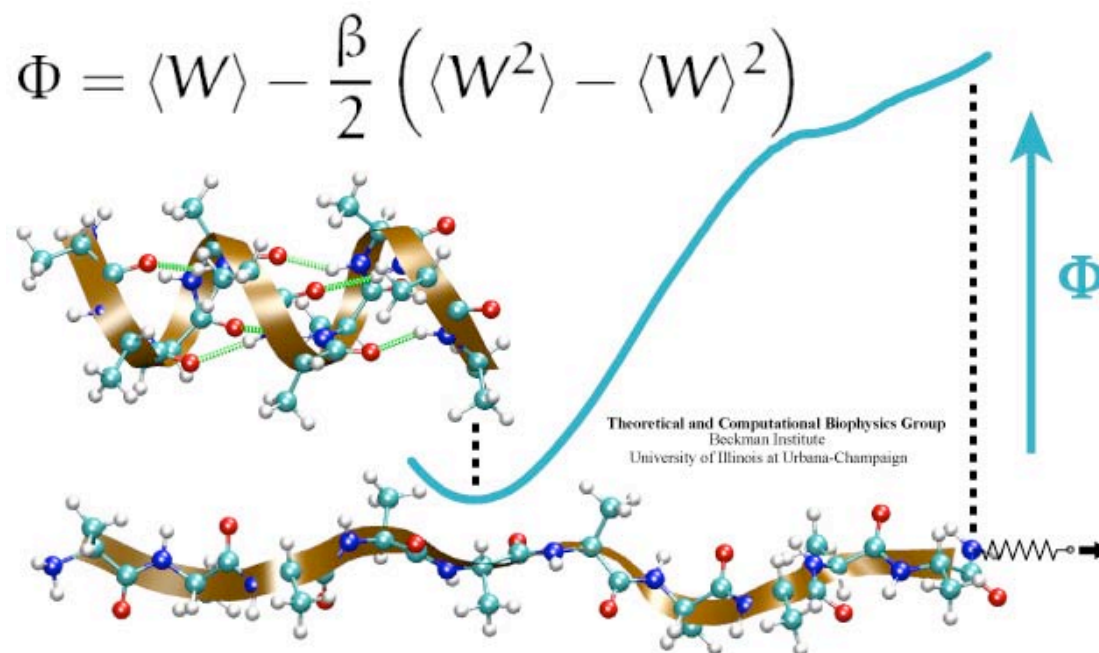
**Thermodynamics:**  $\Delta G \leq \langle W \rangle$

**Is there any chance to discount the irreversible work? Yes!**

# Free Energy of Stretched Alpha-Helix (Deca-alanine)

**Thermodynamics:**  $\Delta G \leq \langle W \rangle$

**Jarzynski (1997):**  $e^{-\Delta G/k_B T} = \langle e^{-W/k_B T} \rangle$



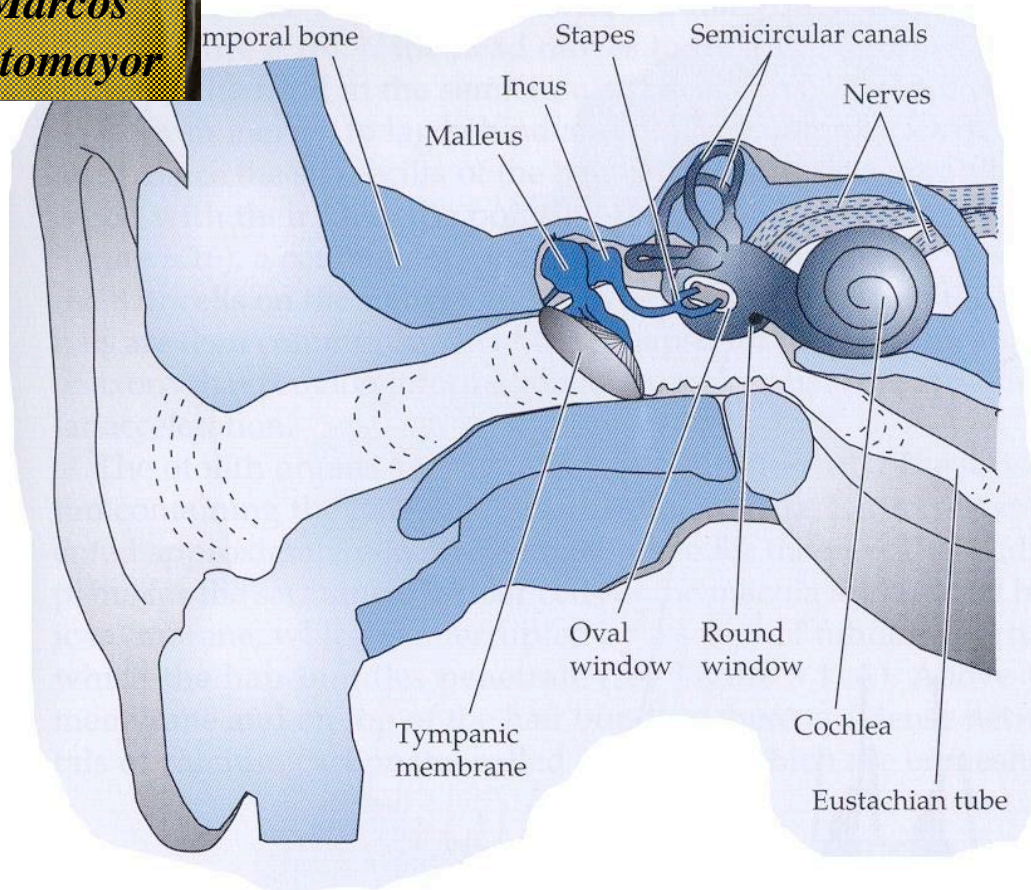
**Free energy calculation from steered molecular dynamics simulations using Jarzynski's equality.** S. Park, F. Khalili-Araghi, E. Tajkhorshid, and K. Schulten. *Journal of Chemical Physics*, 119:3559-3566, 2003

**Calculating potentials of mean force from steered molecular dynamics simulations.** S. Park and K. Schulten. *Journal of Chemical Physics*, 120: 5946-5961, 2004

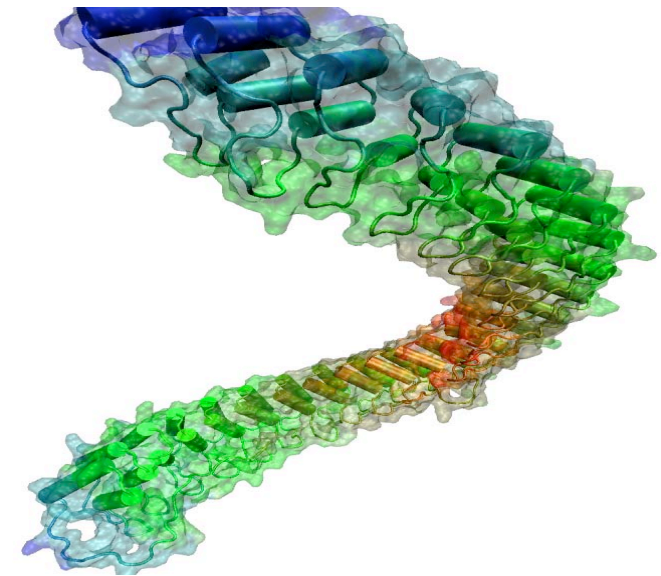


# Molecular Basis of Hearing

## *Molecular Modeling Ahead of Observation*



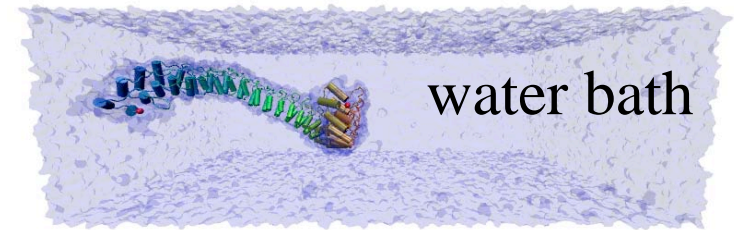
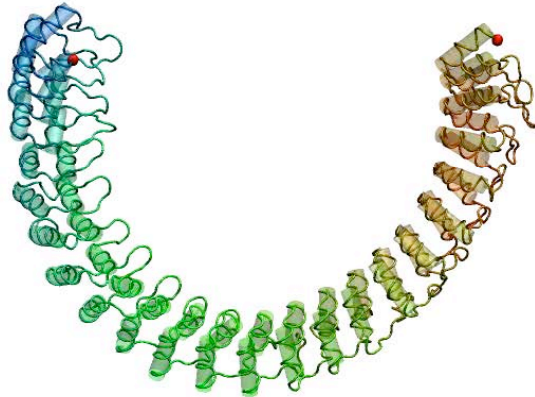
340,000 atoms



Ankyrin -  
gating spring in the  
inner ear hair cells

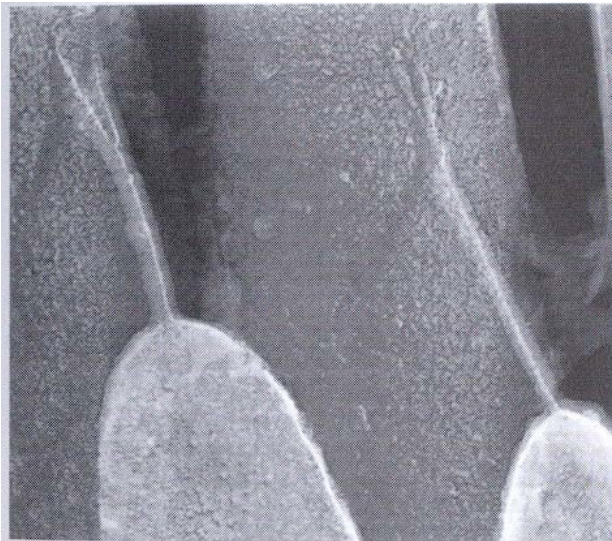
Mammalian Inner Ear (from Sensory Transduction, G. L. Fain).

# Ankyrin - Tertiary Structure Spring at weak force (100 pN)



**340,000 atom simulation  
of 24 repeat ankyrin**

Experimentally verified, papers submitted



**Tip Links** (Kachar et al., 2000; Corey Lab)

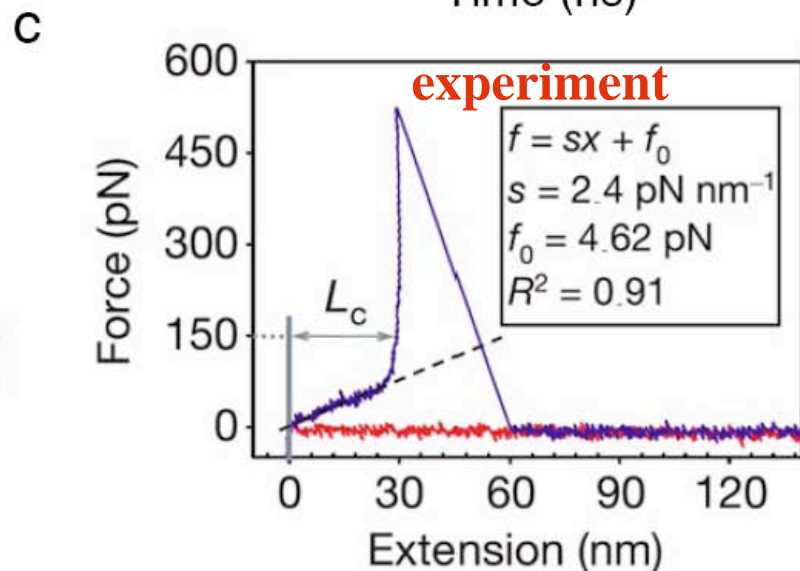
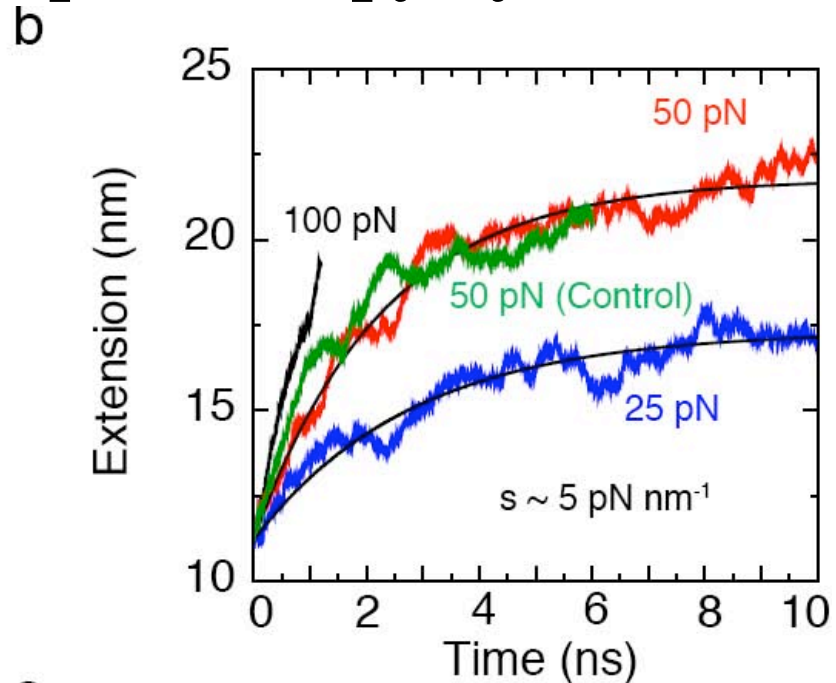
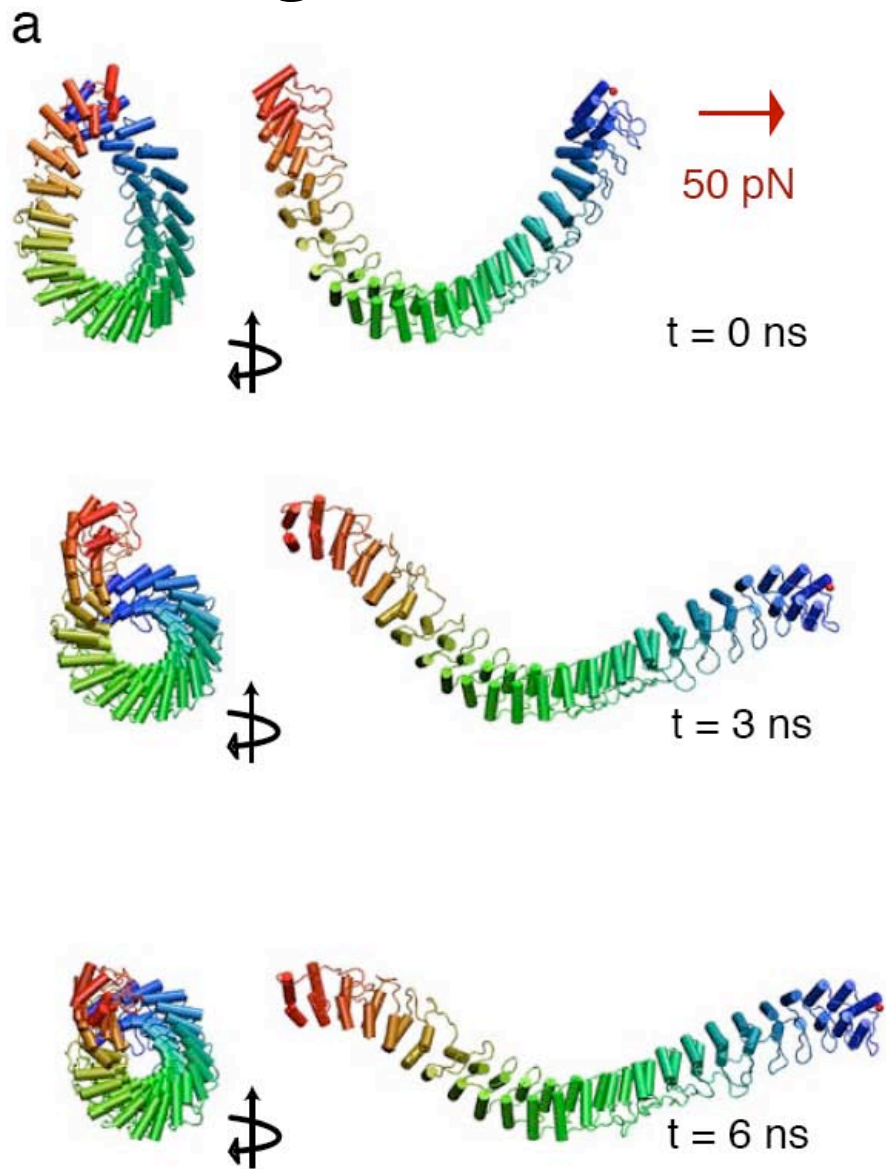


**Inner Ear Mechanism**

M. Sotomayor, D. Corey, and K. Schulten *Structure*, 13:669-682, 2005

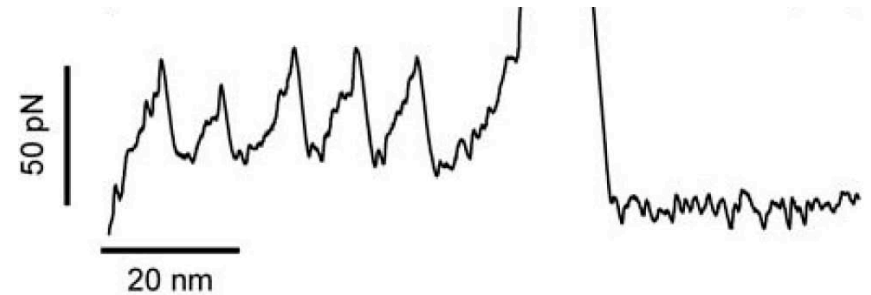
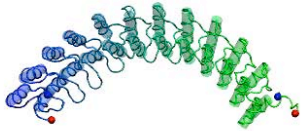
**Hair bundle** (Assad and Corey, from Sensory Transduction, G. L. Fain).

# Hookean Elasticity of 24 repeats of Ankyrin measured by Single Molecule Force Spectroscopy by AFM



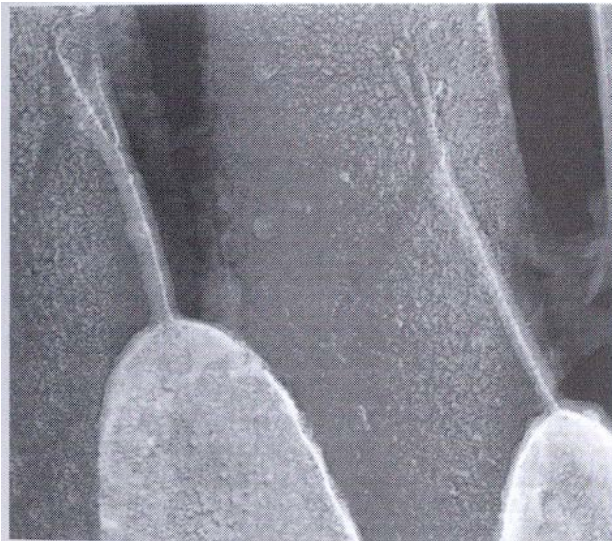


# Ankyrin - Secondary Structure Spring at Large Force



Experiment:

L. Li, S. Wetzel, A. Pluckthun, and J. M. Fernandez, *Biophys. J.* 90, L30–L32, 2006.



**Tip Links** (Kachar et al., 2000; Corey Lab)

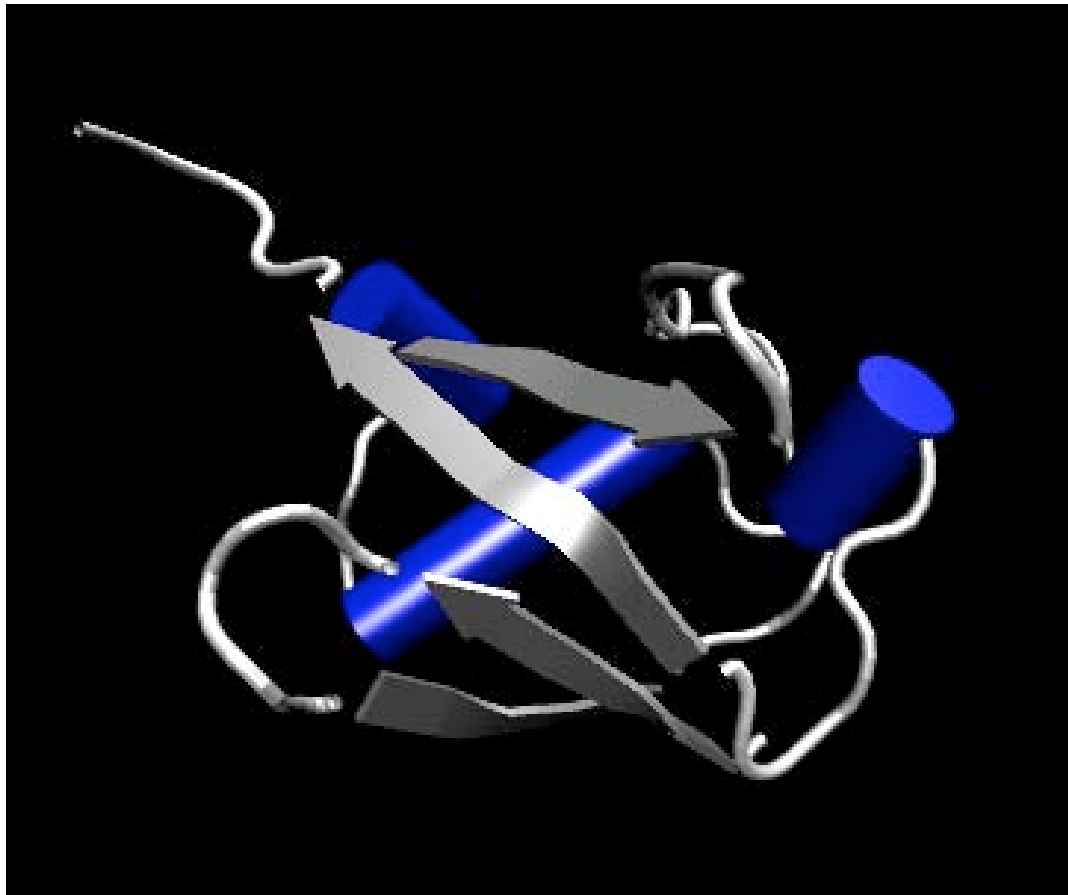


**Inner Ear Mechanism**

M. Sotomayor, D. Corey, and K. Schulten *Structure*, 13:669-682, 2005

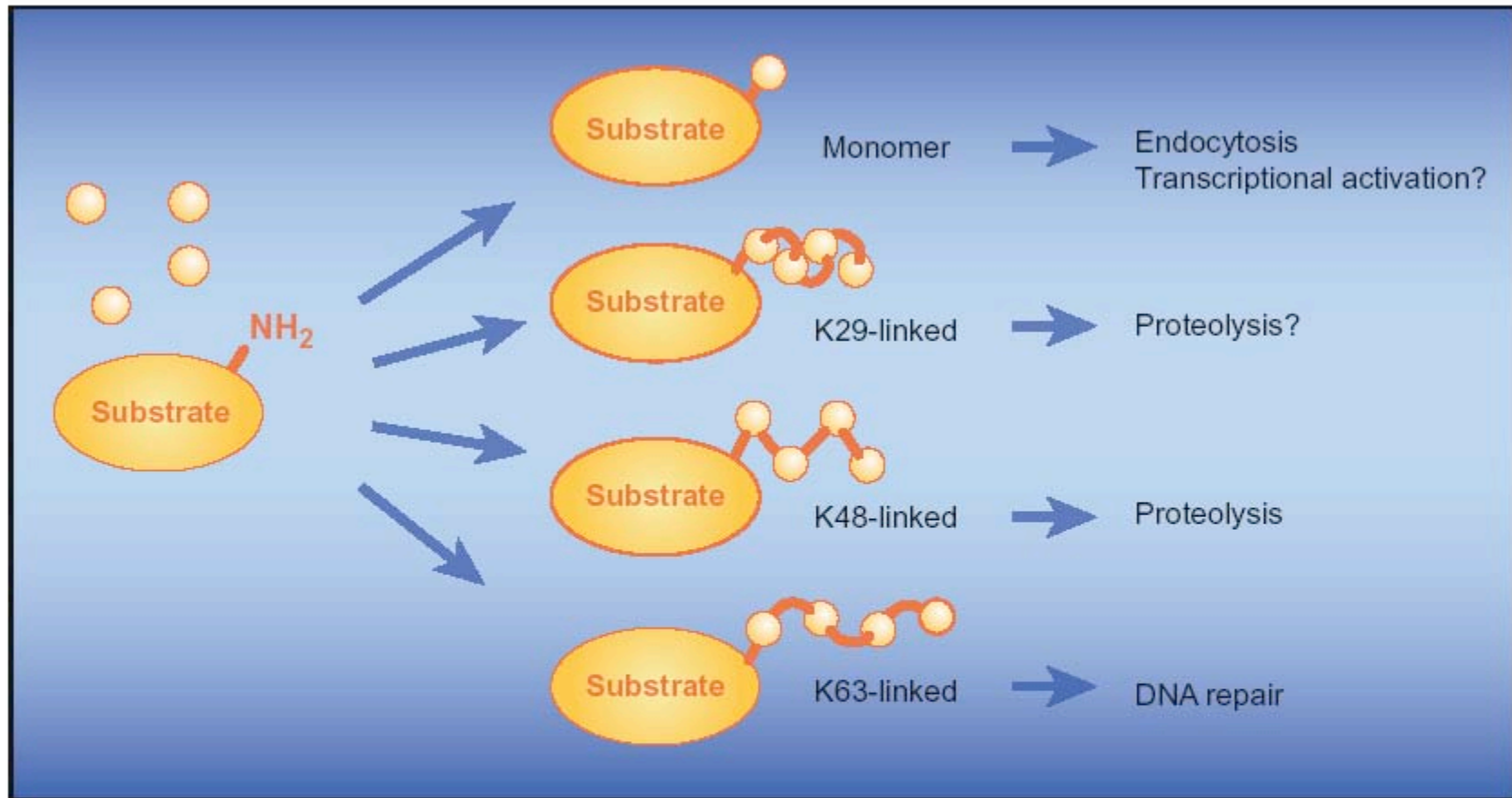
**Hair bundle** (Assad and Corey, from *Sensory Transduction*, G. L. Fain).

# Ubiquitin



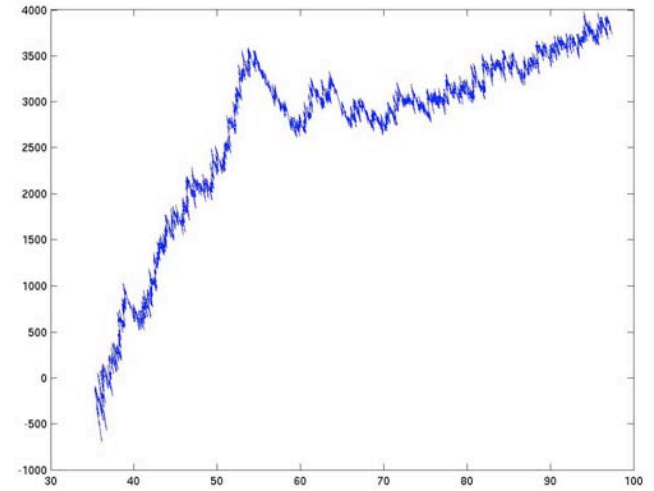
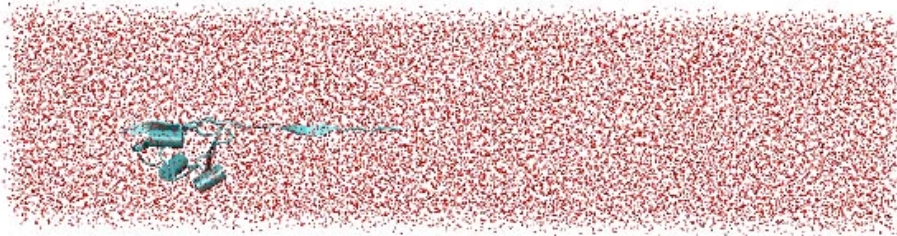
Fatemeh Araghi, Timothy Isgro, Marcos Sotomayor

# Monoubiquitylation versus multi-ubiquitylation



**Multifaceted.** Ubiquitin can attach to its various substrate proteins, either singly or in chains, and that in turn might determine what effect the ubiquitination has. (K29, K48, and K63 refer to the particular lysine amino acid used to link the ubiquitins to each other.)

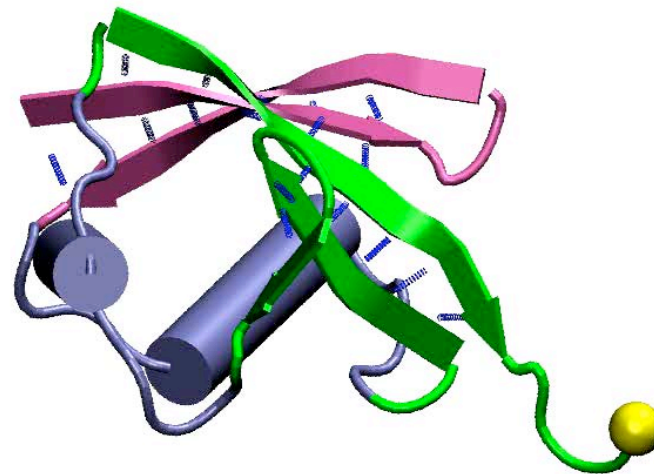
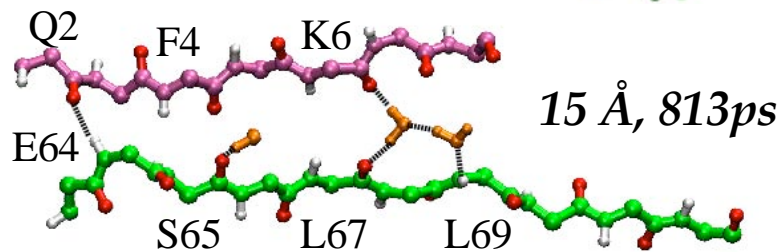
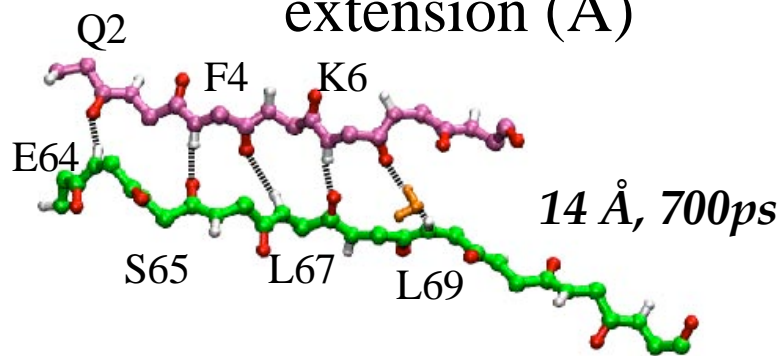
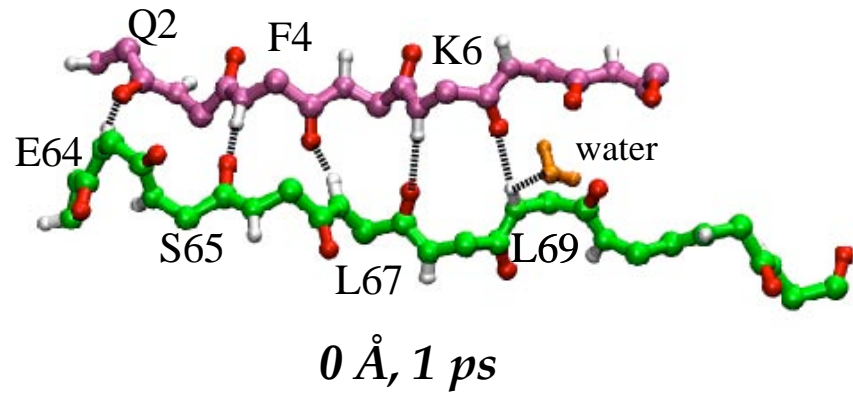
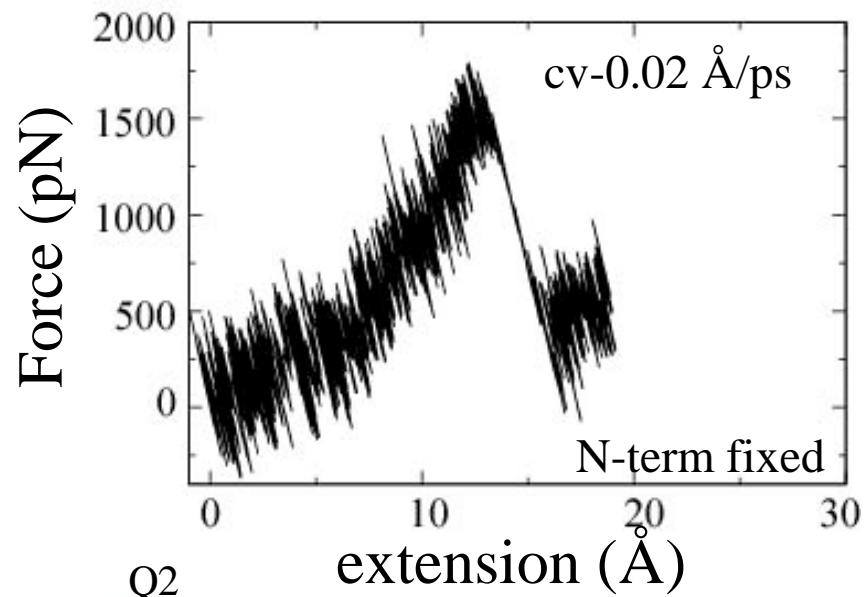
# First SMD Simulation



First peak when the first beta strand is stretched out

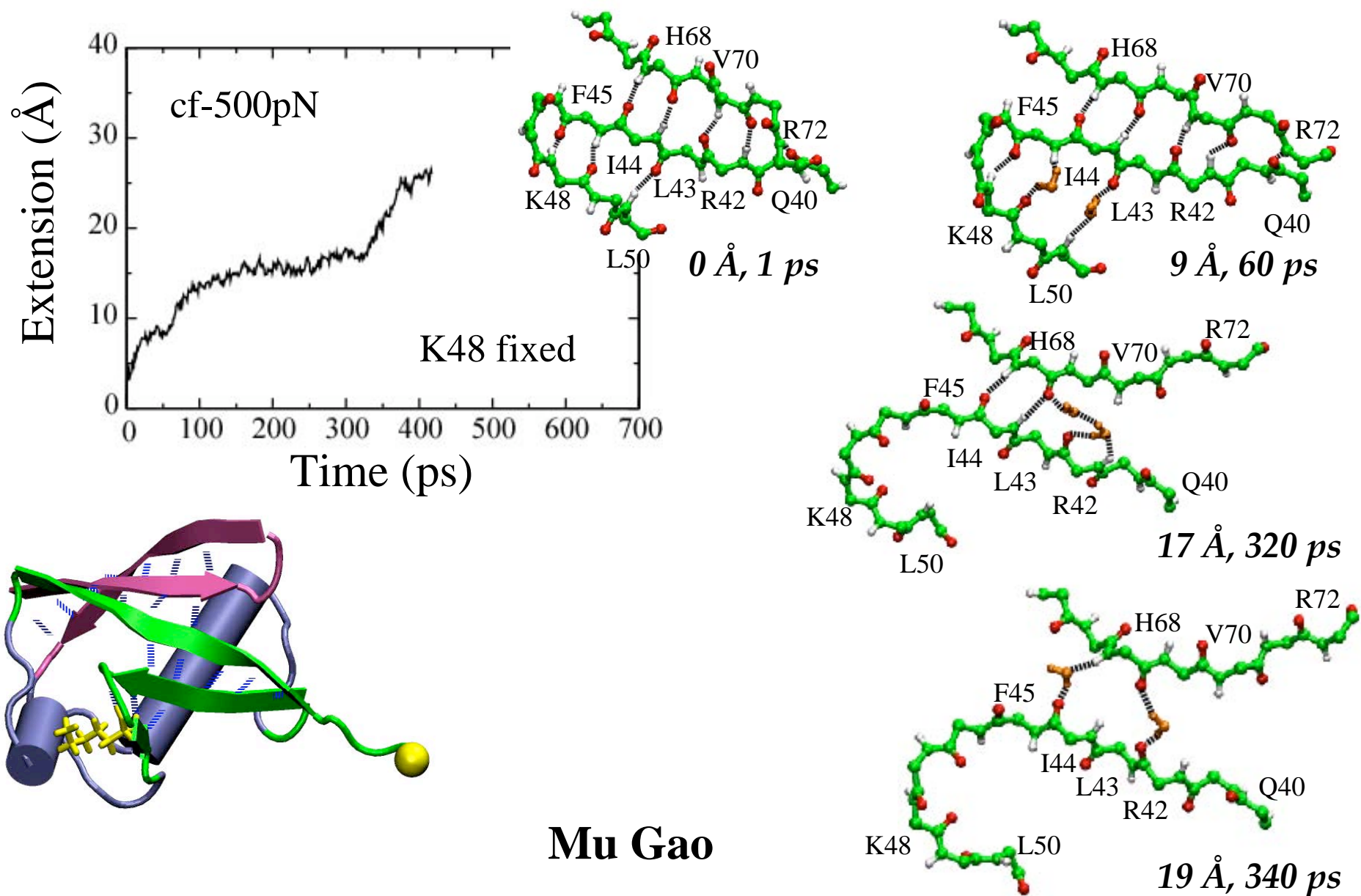
- SMD simulation, with constant velocity
- Box of water 70x240x70 Å ~81K atoms
- smd velocity 0.4 Å/ps
- smd spring constant 7 kcal/mol Å<sup>2</sup>

# Ubiquitin Unfolding I



Mu Gao

# Ubiquitin Unfolding II



# Pulling Dimer

- SMD ( $v=0.4$  A/ps  $k=7$  kcal/mol A<sup>2</sup>) constant P
- Two monomers separate.

