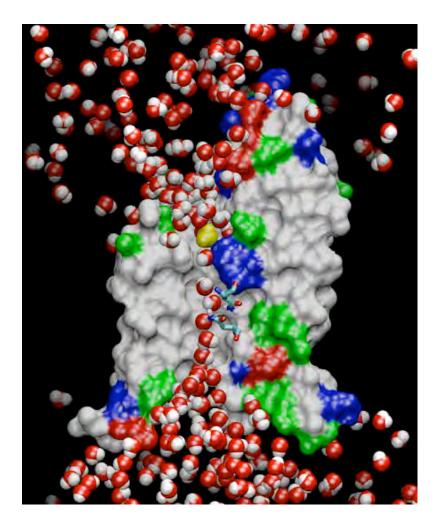
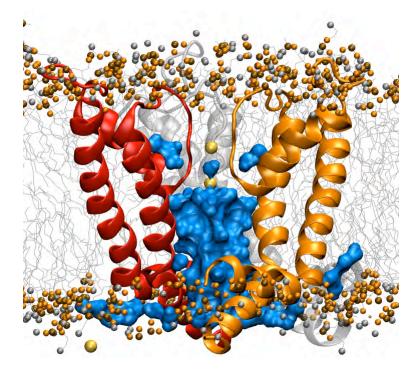
#### Molecular Dynamics Studies of Mechanisms of Permeation, Selectivity, and Gating in Membrane Channels





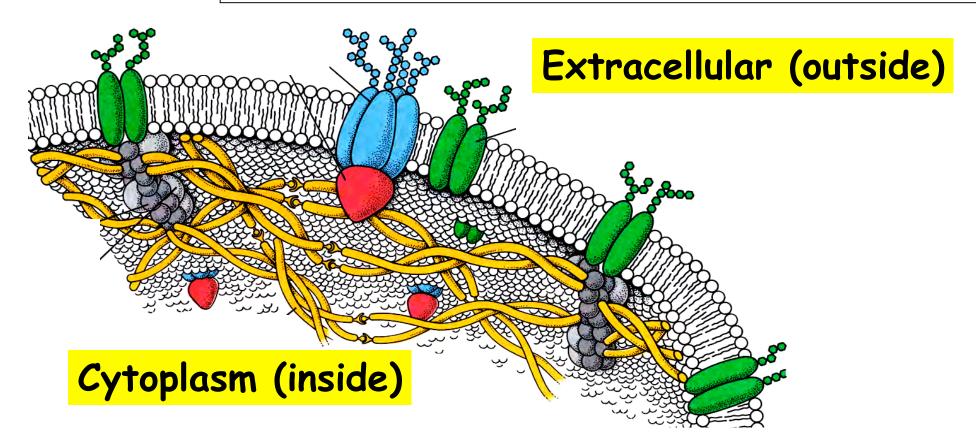
Emad Tajkhorshid Department of Biochemistry, Center for Biophysics and Computational Biology, and Beckman Institute University of Illinois at Urbana-Champagin



# Why Do Living Cells Need Membrane Channels (Proteins)?

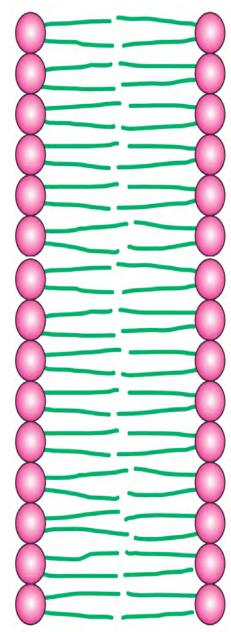
• Living cells also need to exchange materials and information with the outside world

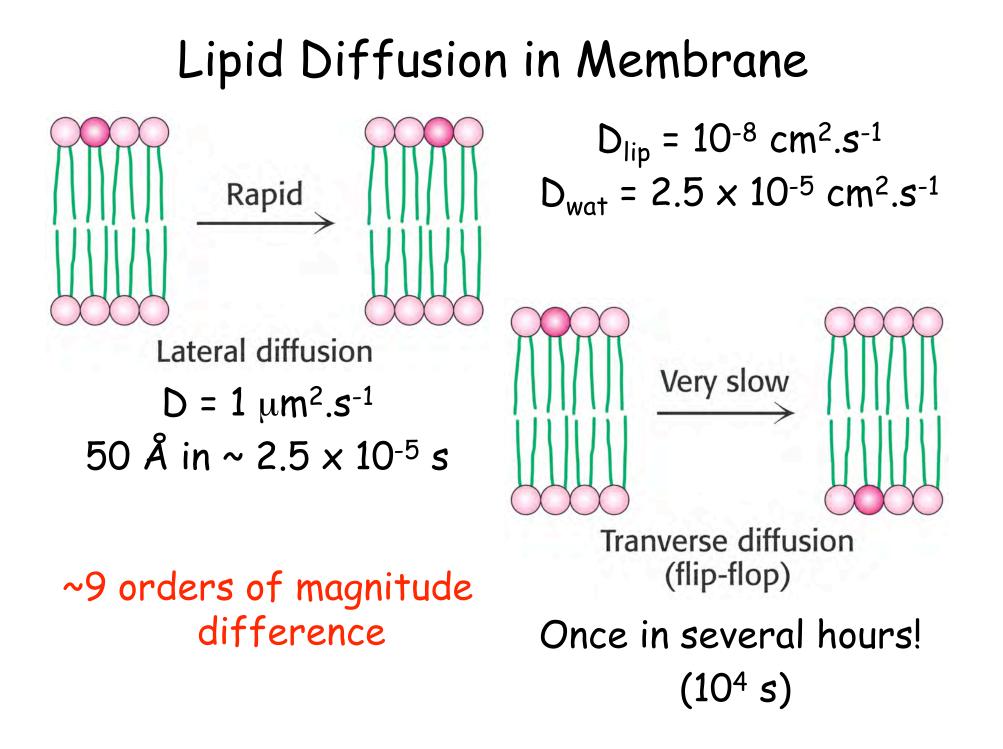
... however, in a highly <u>selective</u> manner.



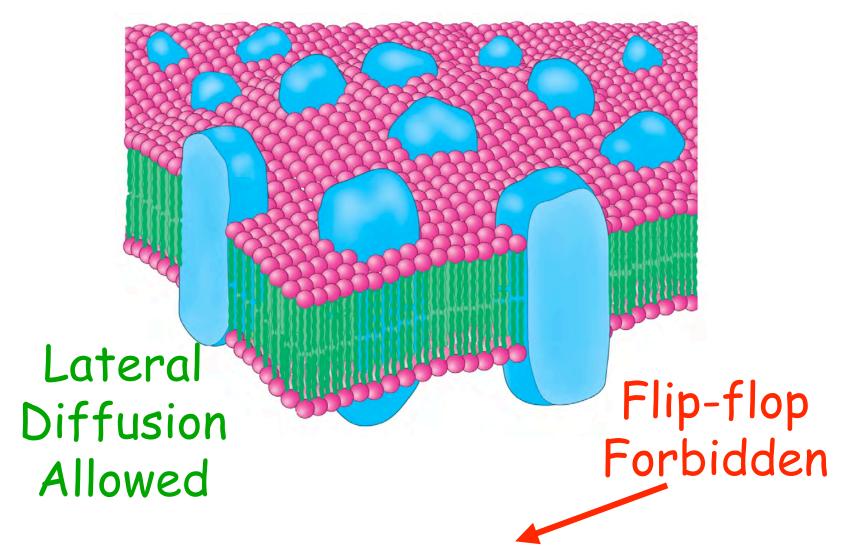
# Lipid Bilayers Are Excellent For Cell Membranes

- Hydrophobic interaction is the driving force
- Self-assembly in water
- Tendency to close on themselves
- Self-sealing (a hole is unfavorable)
- Extensive: up to millimeters





### Fluid Mosaic Model of Membrane

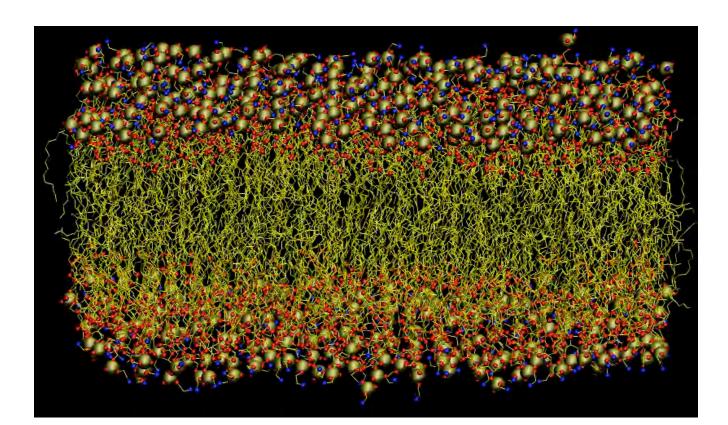


Ensuring the conservation of membrane asymmetric structure

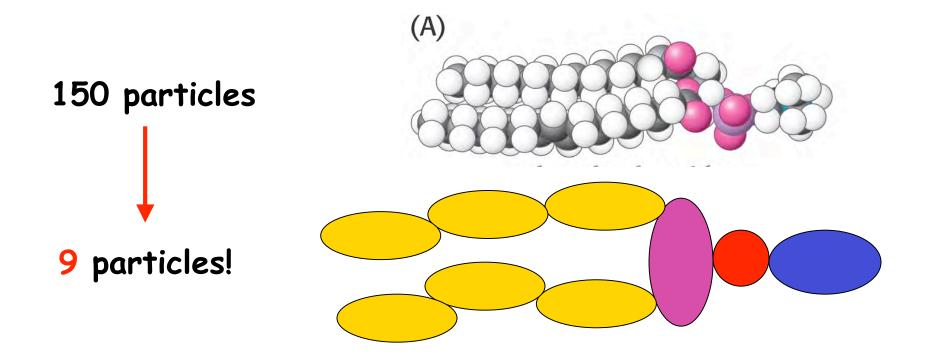
# Technical difficulties in Simulations of Biological Membranes

- Time scale
- Heterogeneity of biological membranes

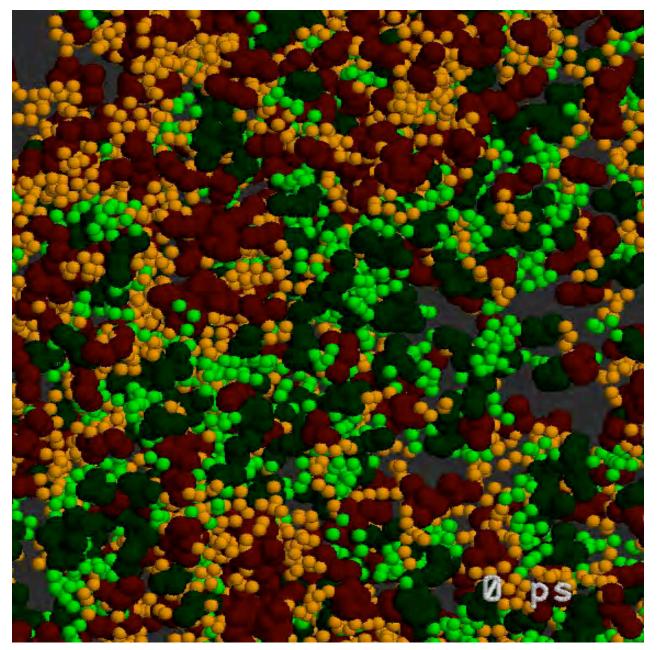
60 x 60 Å Pure POPE 5 ns ~100,000 atoms







Also, increasing the time step by orders of magnitude.

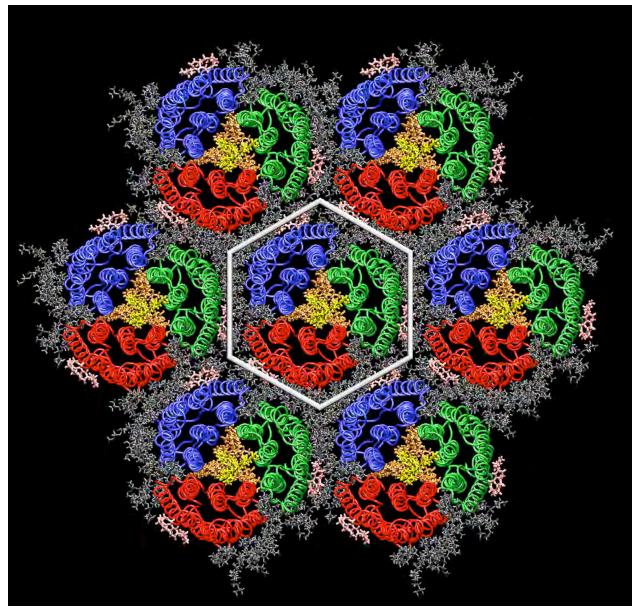


by: J. Siewert-Jan Marrink and Alan E. Mark, University of Groningen, The Netherlands

# Protein/Lipid ratio

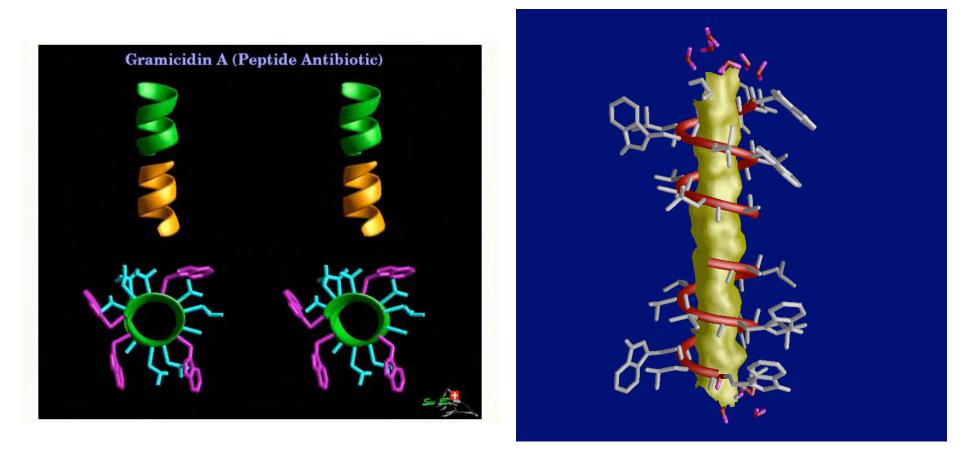
- Pure lipid: insulation (neuronal cells)
- Other membranes: on average 50%
- Energy transduction membranes (75%)
   Membranes of mitocondria and chloroplast
   Purple membrane of halobacteria
- Different functions = different protein composition

# Protein / Lipid Composition

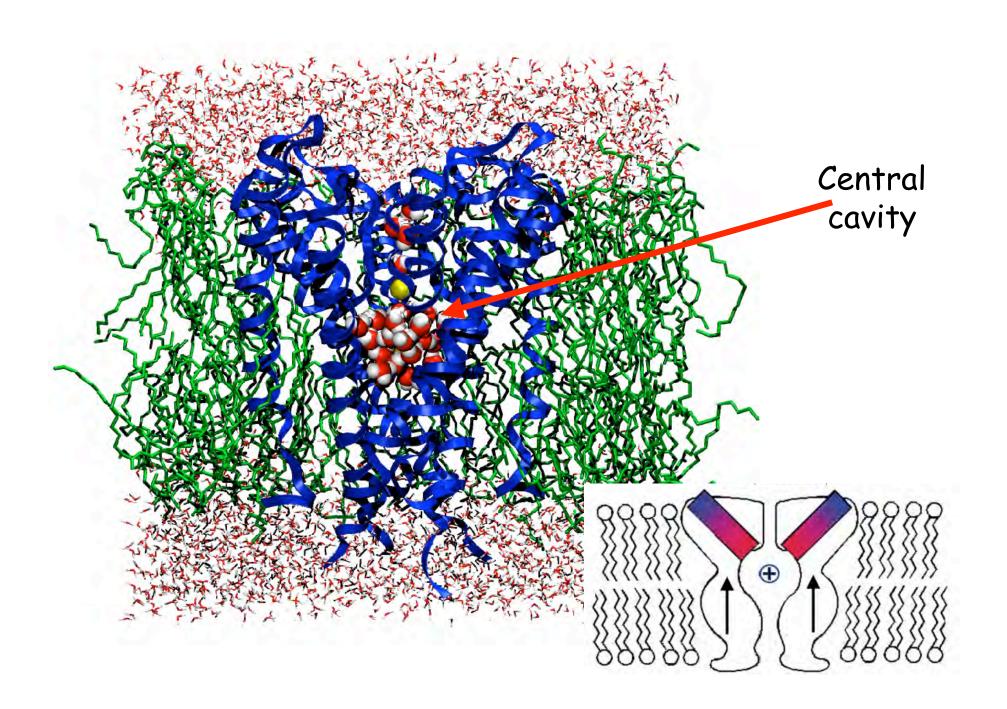


#### The purple membrane of halobacteria

#### Gramicidin A an ion leak inside the membrane



Through dissipating the electrochecmical potential of membrane, gramicidin A acts as an antibiotic.

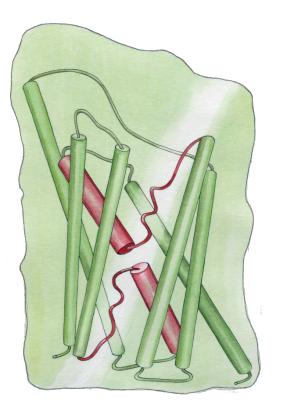


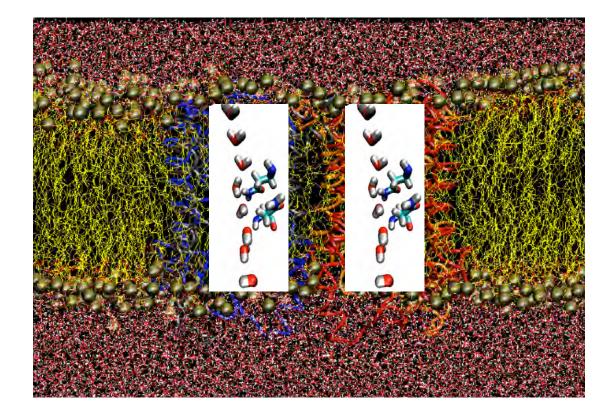
# Analysis of Molecular Dynamics Simulations of Biomolecules

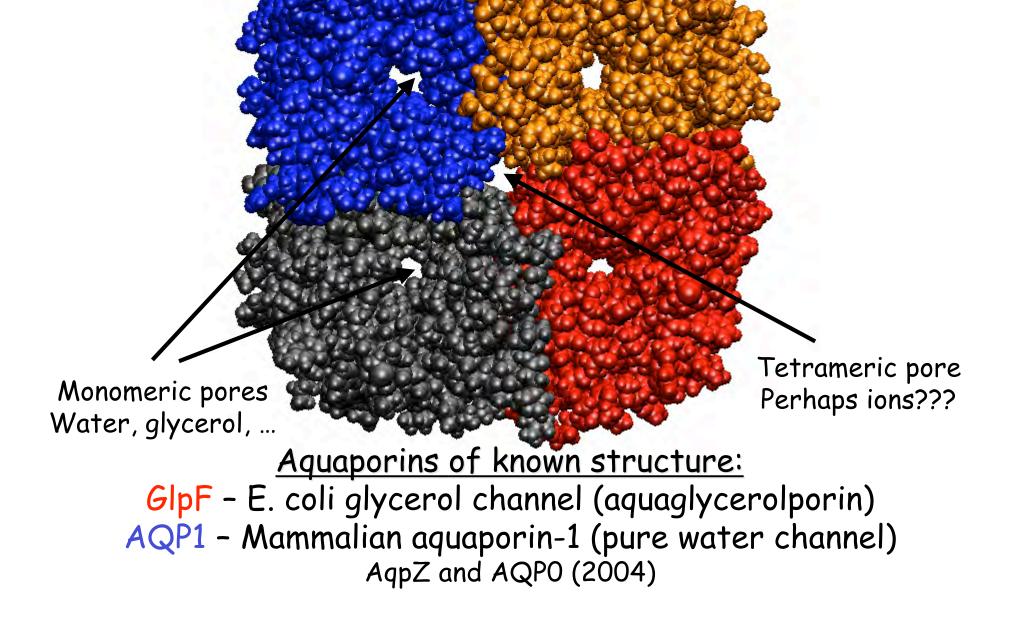
- A very complicated arrangement of hundreds of groups interacting with each other
- Where to start to look at?
- What to analyze?
- How much can we learn from simulations?

# It is very important to get acquainted with your system

# Aquaporins Membrane water channels

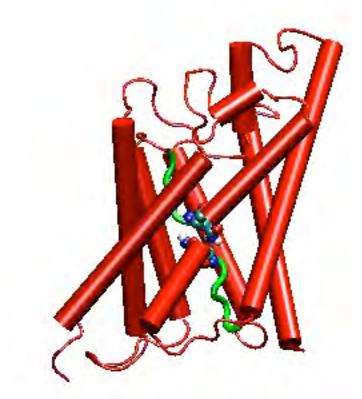


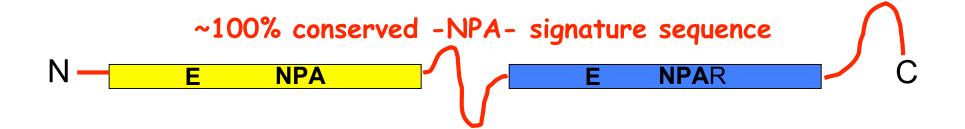




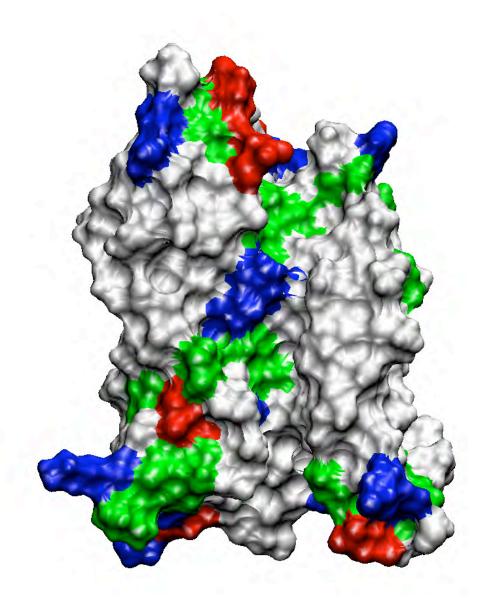
# Functionally Important Features

- Tetrameric architecture
- Amphipatic channel interior
- Water and glycerol transport
- Protons, and other ions are excluded
- Conserved asparagine-prolinealanine residues; NPA motif
- Characteristic half-membrane
   spanning structure





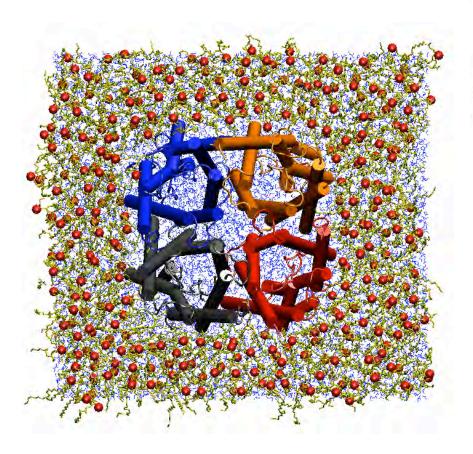
#### A Semi-hydrophobic channel

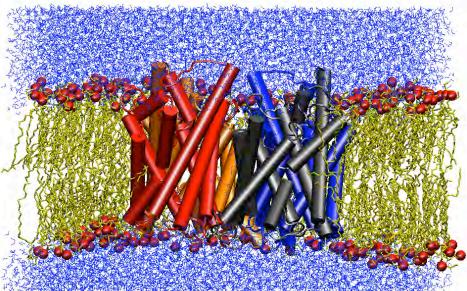


# Molecular Dynamics Simulations

Protein: ~ Lipids (POPE): ~ Water: ~ Total: ~

- 15,000 atoms
  40,000 atoms
  51,000 atoms
  - ~ 106,000 atoms

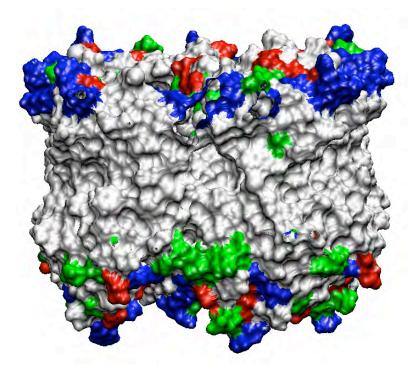


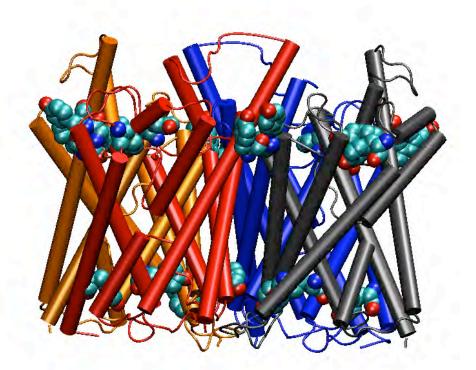


NAMD, CHARMM27, PME NpT ensemble at 310 K 1ns equilibration, 4ns production 10 days /ns - 32-proc Linux cluster 3.5 days/ns - 128 O2000 CPUs

0.35 days/ns - 512 LeMieux CPUs

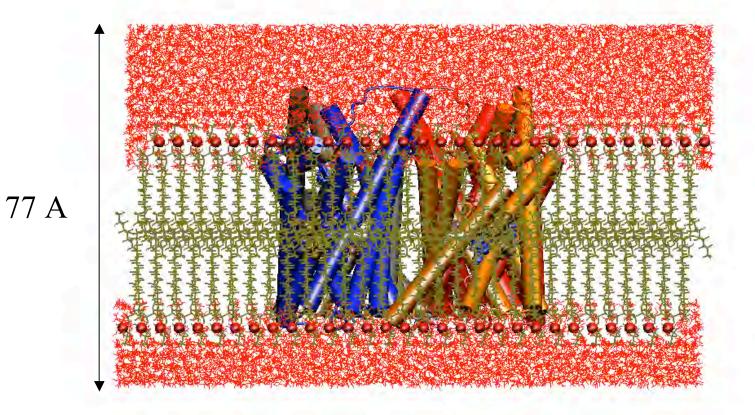
# Protein Embedding in Membrane

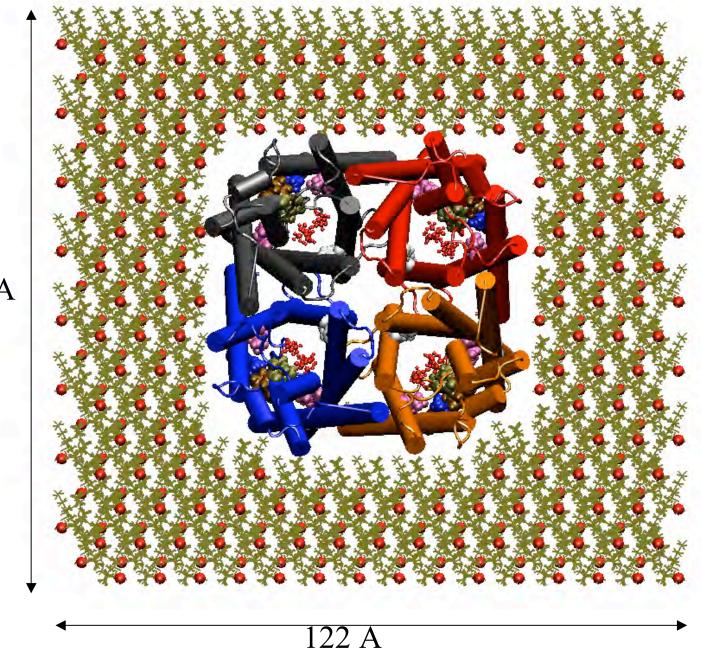




Hydrophobic surface of the protein Ring of Tyr and Trp

# Embedding GlpF in Membrane



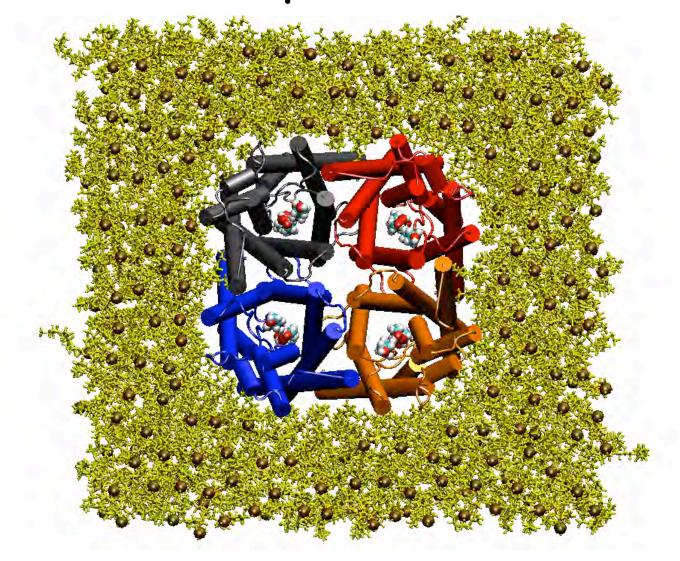


112 A

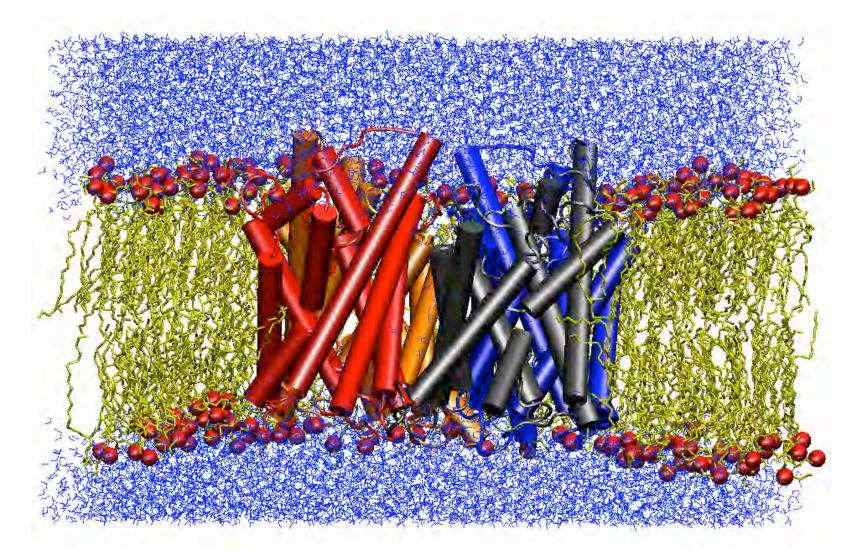
#### A Recipe for Membrane Protein Simulations

- Insert your protein into a hydrated lipid bilayer.
- Fix the protein; minimize the rest and run a short "constantpressure" MD to bring lipids closer to the protein and fill the gap between the protein and lipids.
- Watch water molecules; if necessary apply constraints to prevent them from penetrating into the open gaps between lipids and the protein.
- Monitor the volume of your simulation box until it is almost constant. Do not run the system for too long during this phase.
- Now release the protein, minimize the whole system, and start an NpT simulation of the whole system.
- If desired, you may switch to an NVT simulation, when the system reaches a stable volume.

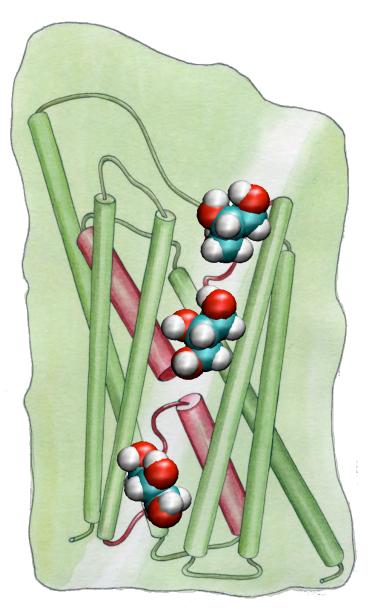
# Lipid-Protein Packing During the Initial NpT Simulation

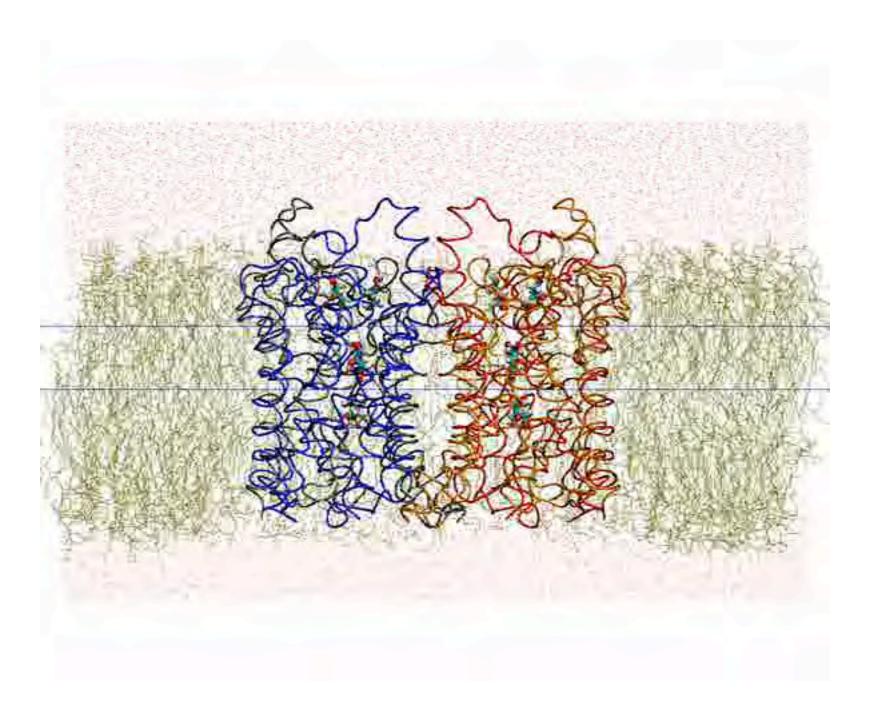


# Adjustment of Membrane Thickness to the Protein Hydrophobic Surface

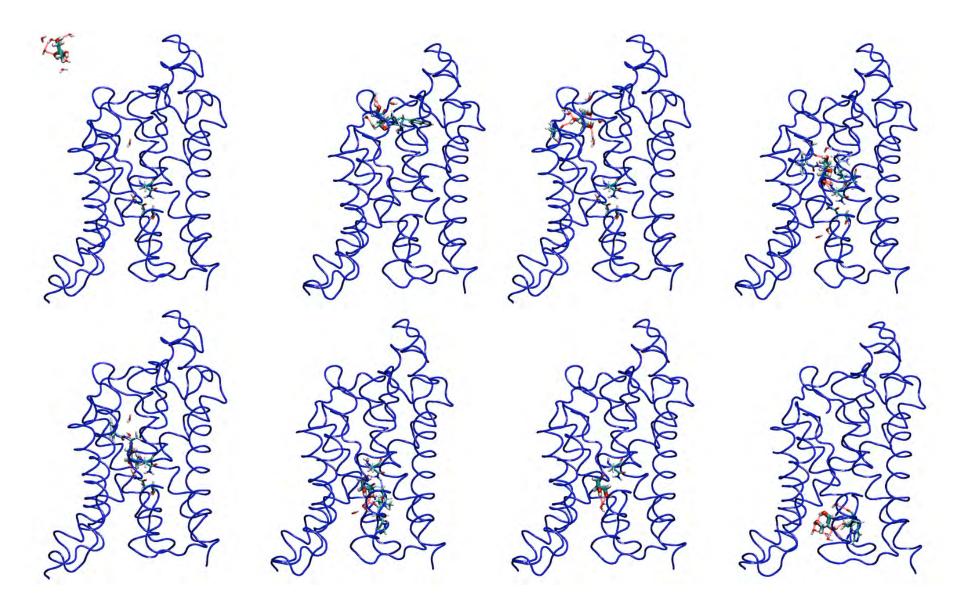


# **Glycerol-Saturated GlpF**





# Description of full conduction pathway



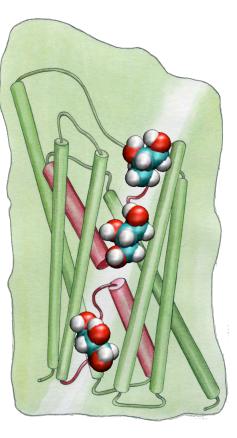
#### Complete description of the conduction pathway



# Channel Hydrogen Bonding Sites

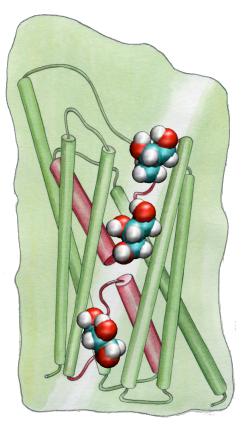
```
{set frame 0}{frame < 100}{incr frame}{
    animate goto $frame
    set donor [atomselect top
    "name 0 N and within 2 of
    (resname GCL and name HO)"]
    lappend [$donor get index] list1
    set acceptor [atomselect top
    "resname GCL and name 0 and
    within 2 of (protein and name HN HO)"]
    lappend [$acceptor get index] list2</pre>
```

. . .



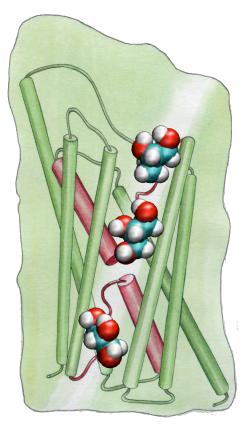
# Channel Hydrogen Bonding Sites

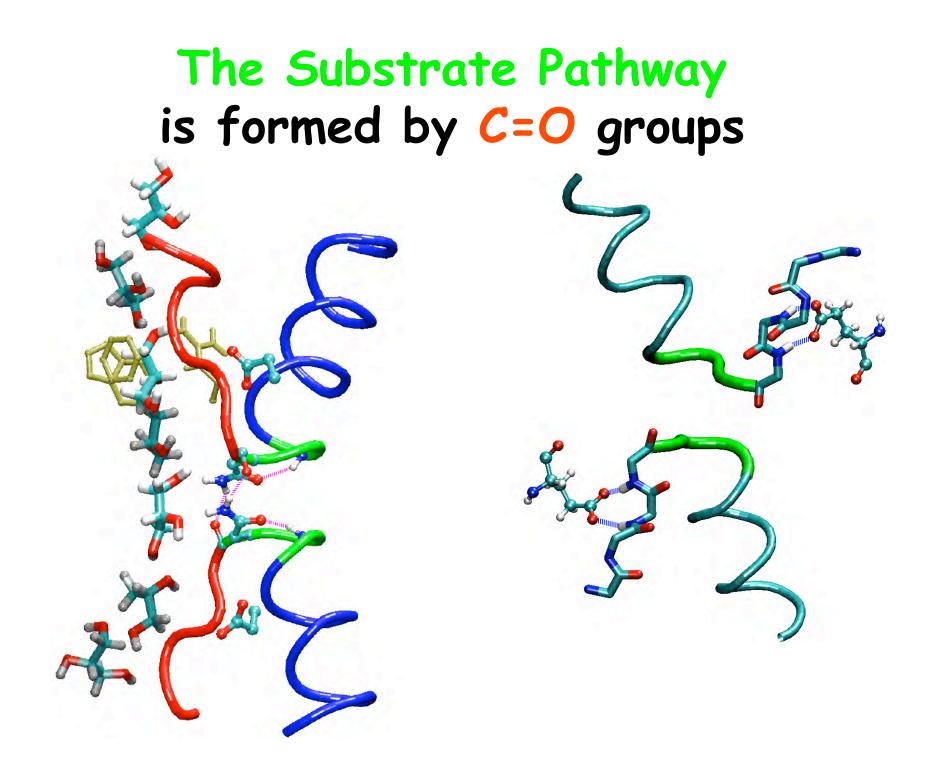
GLN	41	OE1 NE2	LEU	<b>197</b>	0
TRP	<b>48</b>	O NE1	THR	<b>198</b>	0
GLY	64	0	GLY	199	0
ALA	65	0	PHE	200	0
HIS	66	O ND1	ALA	201	0
LEU	67	0	ASN	203	ND2
ASN	68	ND2			
ASP	130	OD1	LYS	33	HZ1 HZ3
GLY	133	0	GLN	41	<b>HE21</b>
SER	136	0	TRP	<b>48</b>	HE1
TYR	138	0	HIS	66	HD1
PRO	139	ΟΝ	<u>ASN</u>	<b>68</b>	HD22
ASN	140	OD1 ND2	TYR	138	HN
HIS	142	ND1	ASN	140	HN HD21 HD22
THR	167	<b>OG1</b>	HIS	142	HD1
GLY	195	0	GLY	199	HN
PRO	196	0	<u>ASN</u>	203	HN HD21HD22
			<u>ARG</u>	206	<b>HE HH21HH22</b>



# Channel Hydrogen Bonding Sites

GLN	41	OE1 NE2	LEU	<b>197</b>	0
TRP	<b>48</b>	O NE1	THR	198	0
GLY	64	0	GLY	199	0
ALA	65	0	PHE	200	0
HIS	66	<b>O ND1</b>	ALA	201	0
LEU	67	0	ASN	203	ND2
ASN	68	ND2			
ASP	130	OD1	LYS	33	HZ1 HZ3
GLY	133	0	GLN	41	HE21
SER	136	0	TRP	<b>48</b>	HE1
TYR	138	0	HIS	66	HD1
PRO	139	<b>O</b> N	<u>ASN</u>	68	HD22
ASN	140	OD1 ND2	TYR	138	HN
HIS	142	ND1	ASN	140	HN HD21 HD22
THR	167	<b>OG1</b>	HIS	142	HD1
GLY	195	0	GLY	199	HN
PRO	196	0	<u>ASN</u>	203	HN HD21HD22
			<u>ARG</u>	206	<b>HE HH21HH22</b>





# The Substrate Pathway is formed by C=O groups

**NPA**R

Non-helical motifs are stabilized by two glutamate residues.

NPA

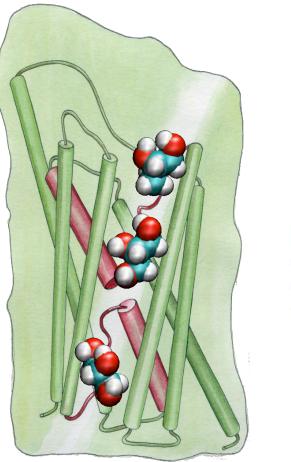
Ν

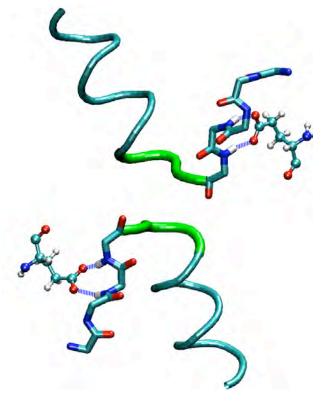
#### Conservation of Glutamate Residue in Human Aquaporins

AQP0HUMAN---LNTLHPAVSVGQATIVEIVELVLTLOFVLCIFATYDE-RRNGQLGAQP1HUMAN---RNDLADGVNSGQGLGIEIIGTLQLVLCVLATTDR-RRRDLGGAQP2HUMAN---VNALSNSTTAGQAVIVELVELVLTLQLVLCIFASTDE-RRGENPGAQP3HUMANGIFATYPSCHLDMINGFPDOFIGTASLIVCVLAIVDPYNNPVPRGAQP4HUMAN---VTMVHGNLTAGHGLIVELITFQLVFTIFASCDS-KRTDVTGAQP5HUMAN---VNALNNNTTQGQAMVELAQP6HUMAN---INVVRNSVSTGQAVAVELITFQLVFTIFASCDS-RRTSPVGAQP6HUMAN---INVVRNSVSTGQAVAVELITFQLVCFASTDS-RQTS-GAQP7HUMAN---INVVRNSVSTGQAVAVELITTCLALAVCMGAIN--EKTKGPAQP8HUMAN-AFVTVQEQGQVAGALAEIILTTLLALAVCMGAIN--EKTKGPAQP9HUMANHIFATYPAPYLSLANAFADQV/ATMILLIIVFAIFDSRNLGAPRGGLPFGLPFECOLIGTFSTYPNPHINFVQAFAVEM/ITAILMGLILALTDDGNGVPRGPruler...180...190...200...210

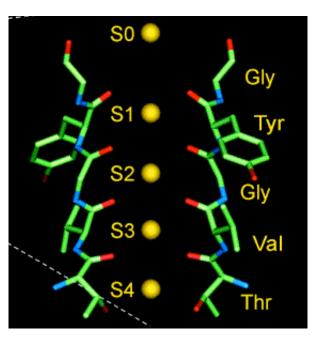
# Glycerol - water competition for hydrogen bonding sites

# Revealing the Functional Role of Reentrant Loops



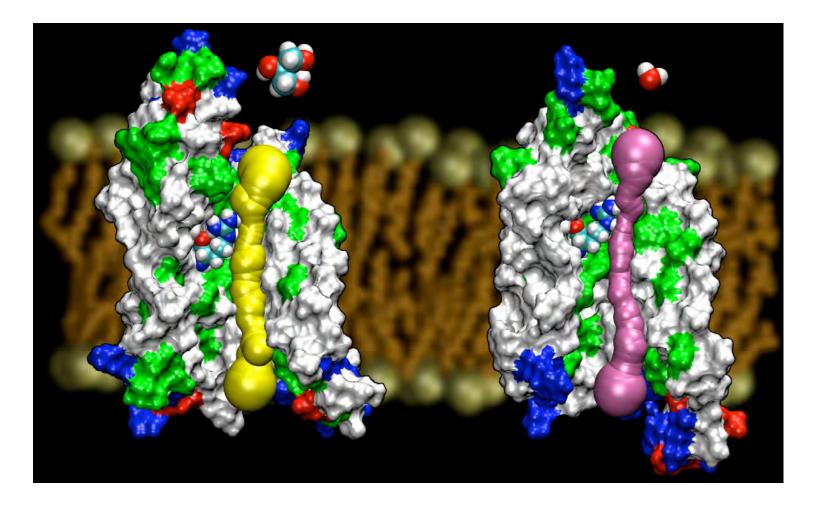


#### Potassium channel



# AqpZ vs. GlpF

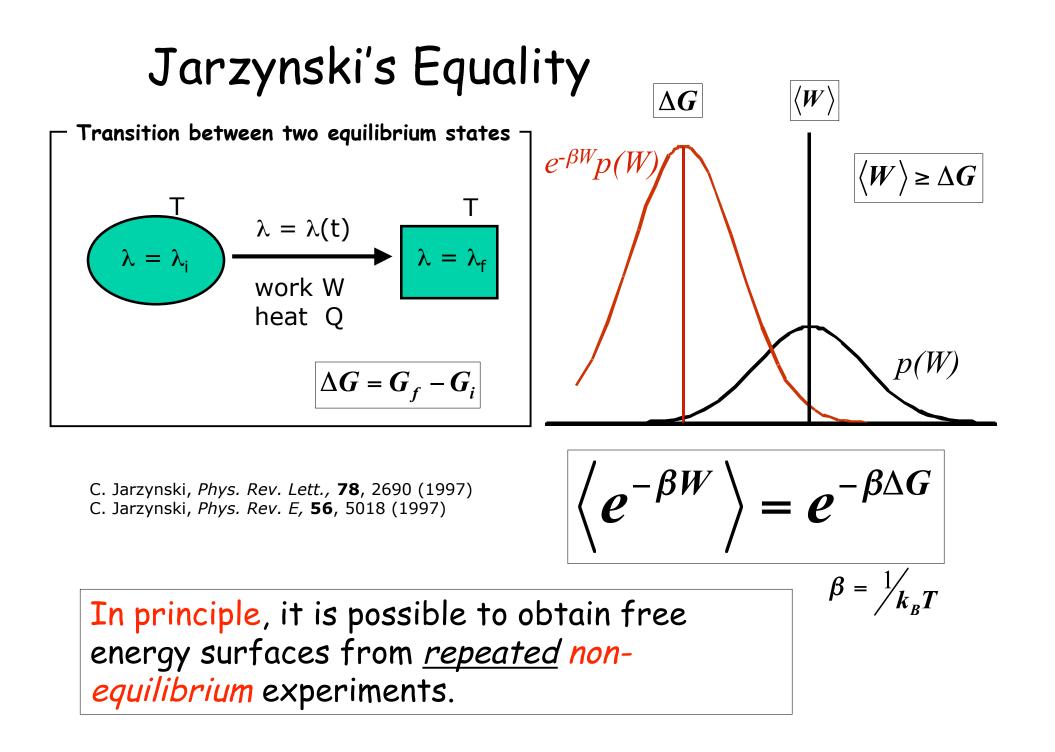
- Both from *E. coli*
- AqpZ is a pure water channel
- GlpF is a glycerol channel
- We have high resolution structures for both channels



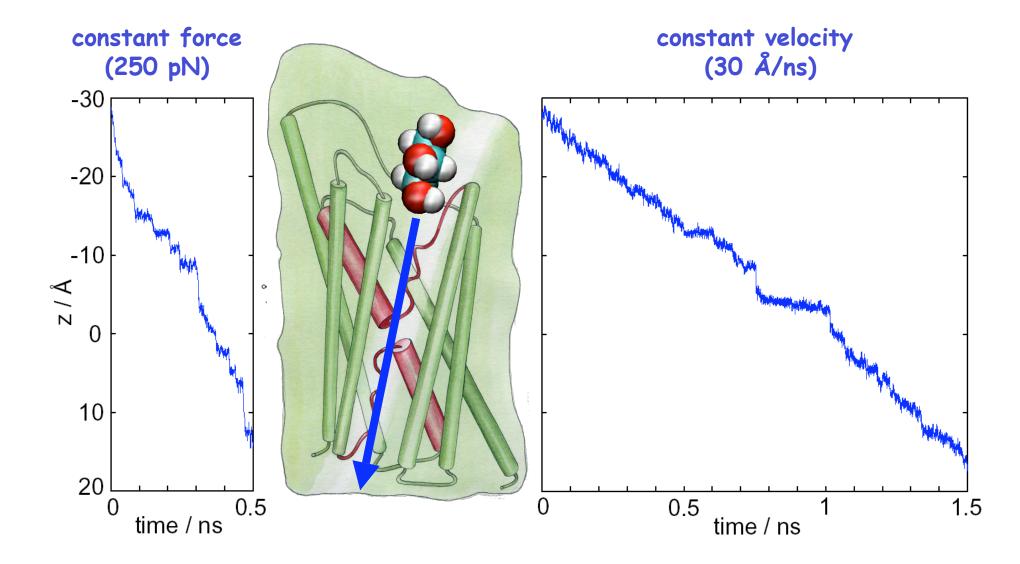
# Steered Molecular Dynamics is a non-equilibrium method by nature

- A wide variety of events that are inaccessible to conventional molecular dynamics simulations can be probed.
- The system will be driven, however, away from equilibrium, resulting in problems in describing the energy landscape associated with the event of interest.

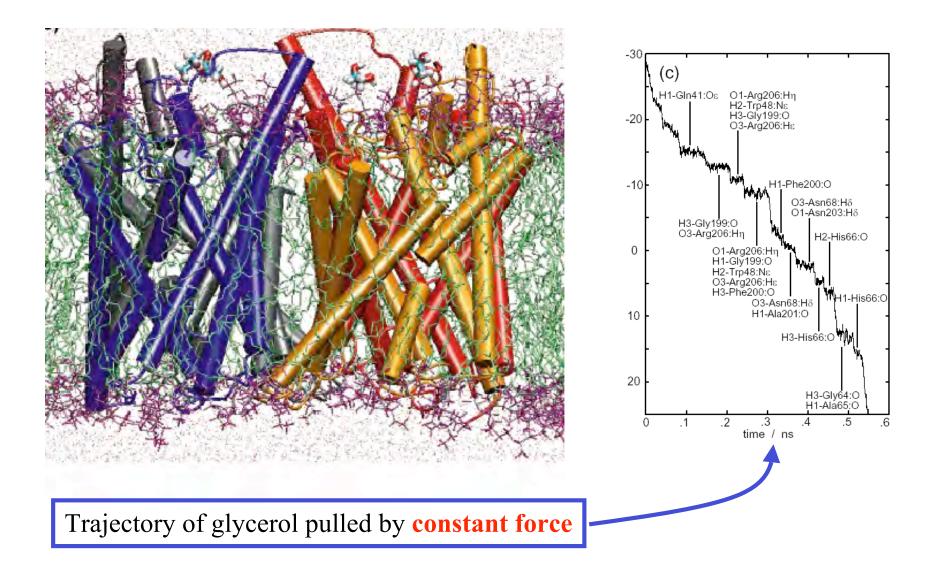
## Second law of thermodynamics $\longrightarrow W \geq \Delta G$



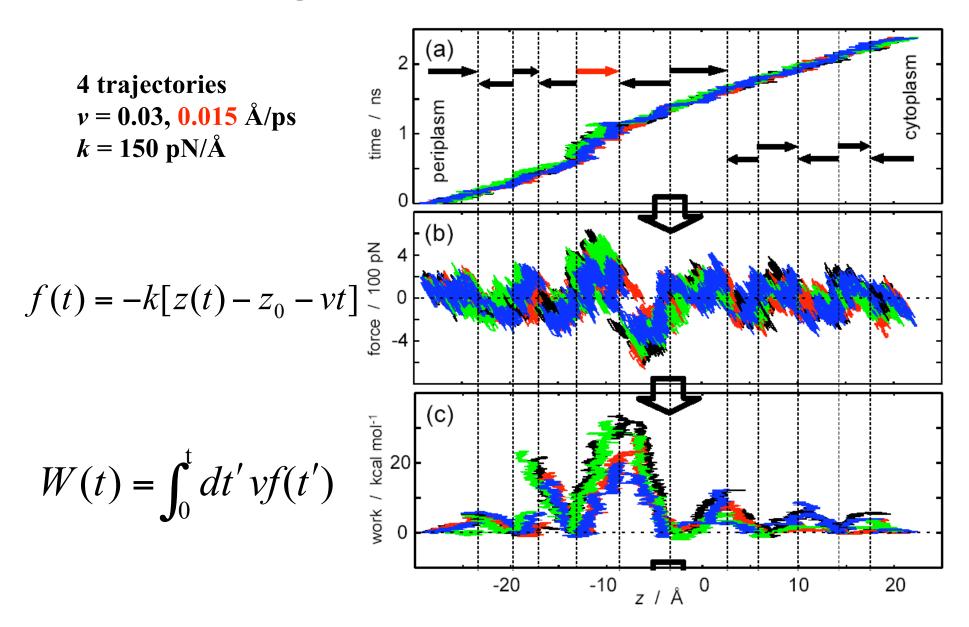
## **Steered Molecular Dynamics**



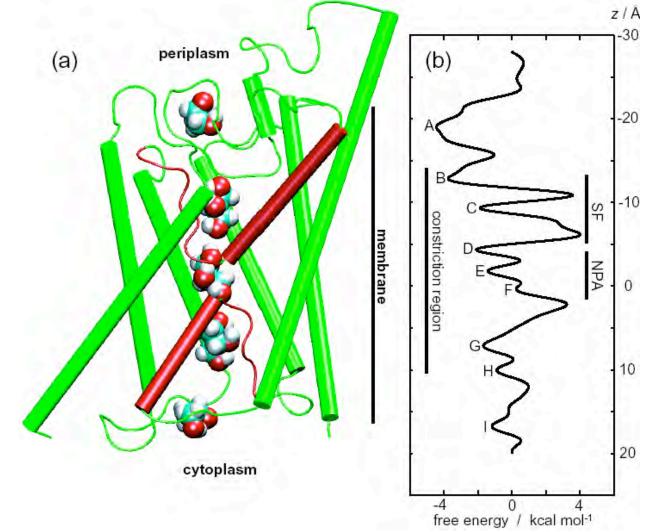
### SMD Simulation of Glycerol Passage



#### Constructing the Potential of Mean Force



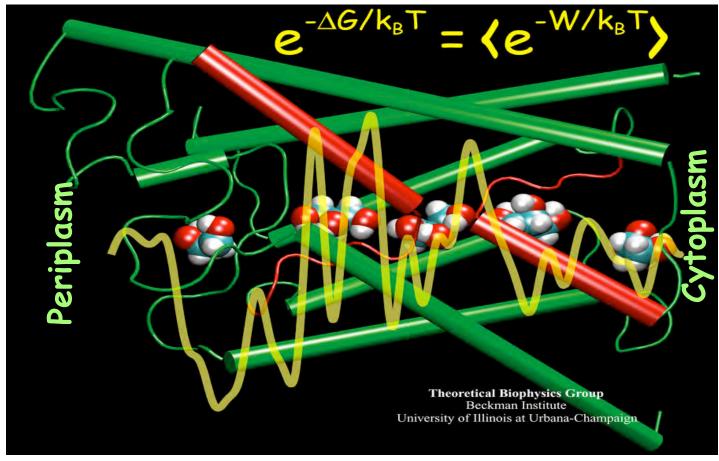
#### Features of the Potential of Mean Force



- Captures major features of the channel
- The largest barrier ~ 7.3 kcal/mol; exp.: 9.6±1.5 kcal/mol

Jensen et al., PNAS, 99:6731-6736, 2002.

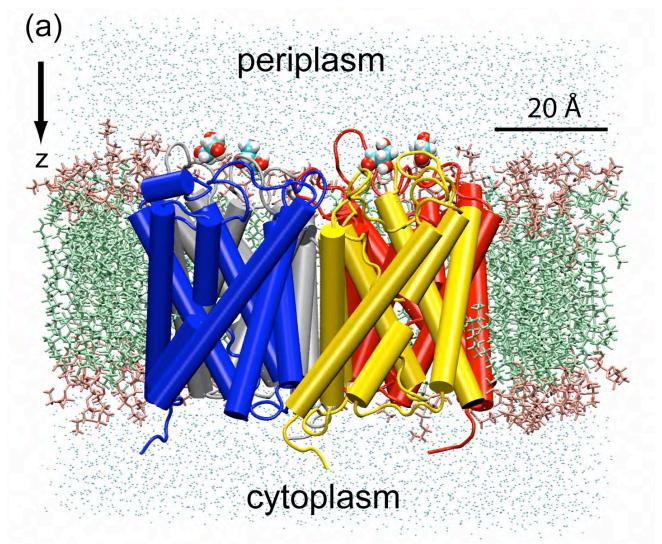
### Features of the Potential of Mean Force



#### Asymmetric Profile in the Vestibules

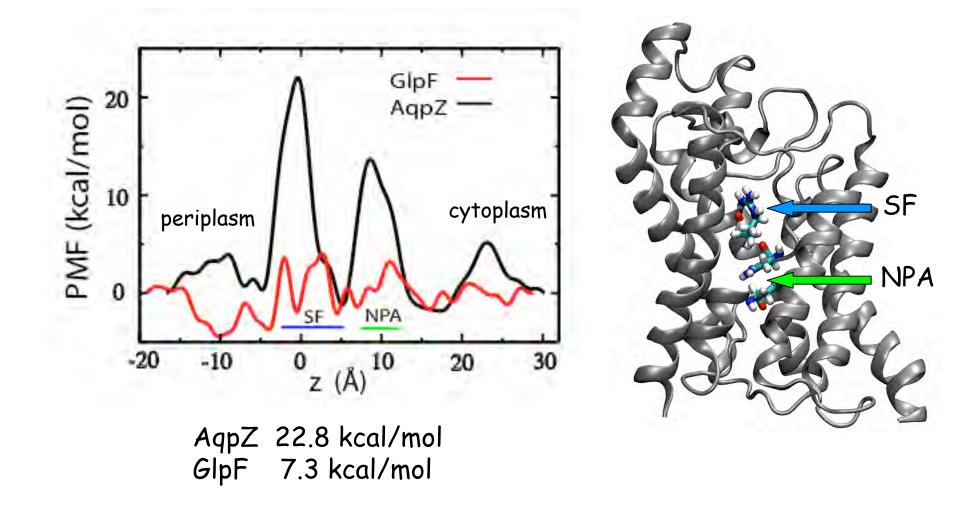
Jensen et al., PNAS, 99:6731-6736, 2002.

# Artificial induction of glycerol conduction through AqpZ



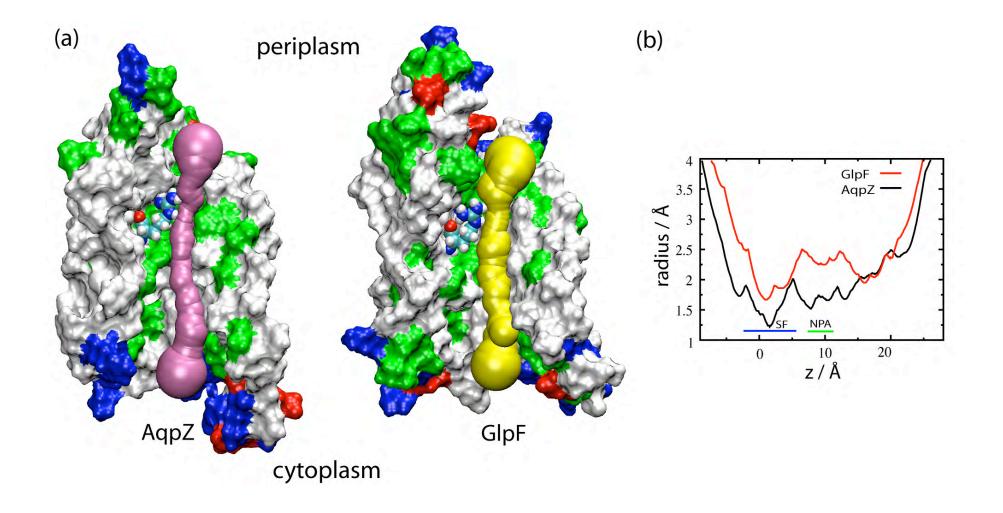
Y. Wang, K. Schulten, and E. Tajkhorshid Structure 13, 1107 (2005)

## Three fold higher barriers



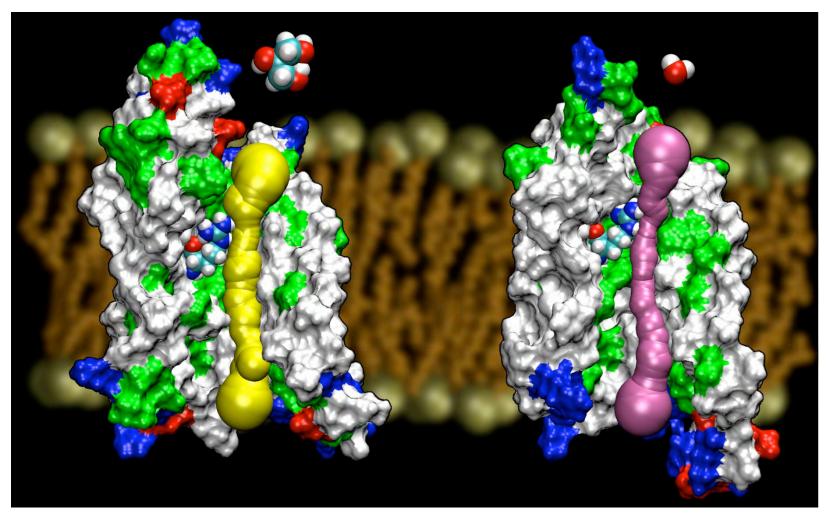
Y. Wang, K. Schulten, and E. Tajkhorshid Structure 13, 1107 (2005)

### Could it be simply the size?



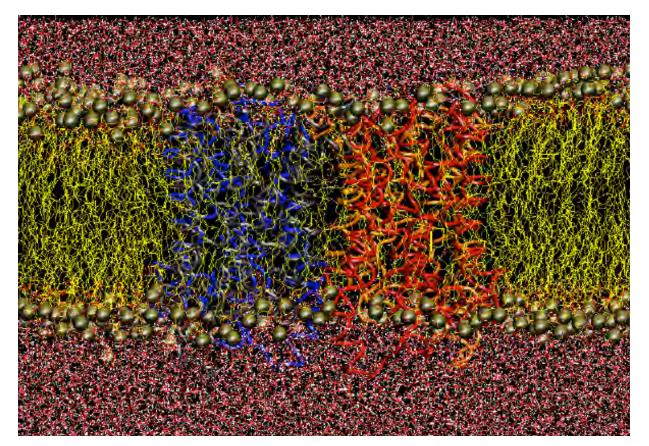
Y. Wang, K. Schulten, and E. Tajkhorshid Structure 13, 1107 (2005)

# It is probably just the size that matters!



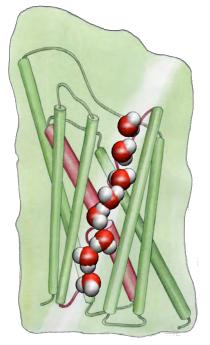
Y. Wang, K. Schulten, and E. Tajkhorshid Structure 13, 1107 (2005)

## Water permeation

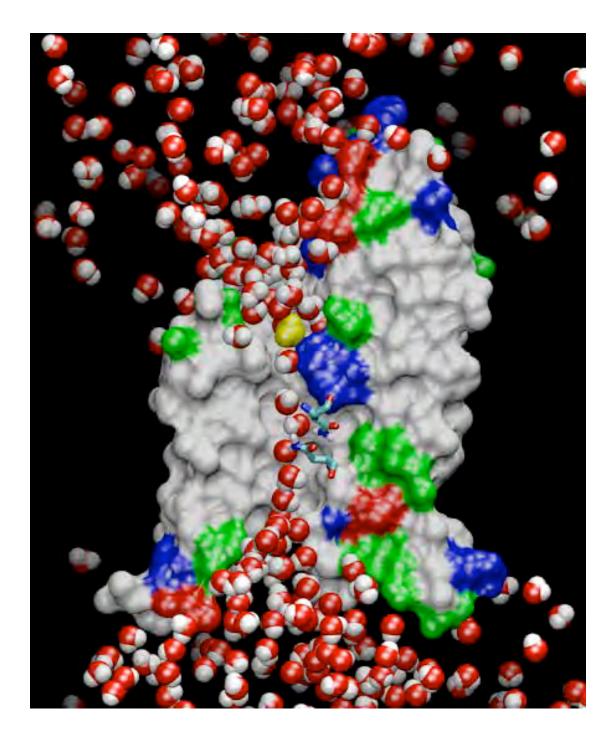


18 water conducted In 4 monomers in 4 ns 1.125 water/monomer/ns Exp. = ~ 1-2 /ns

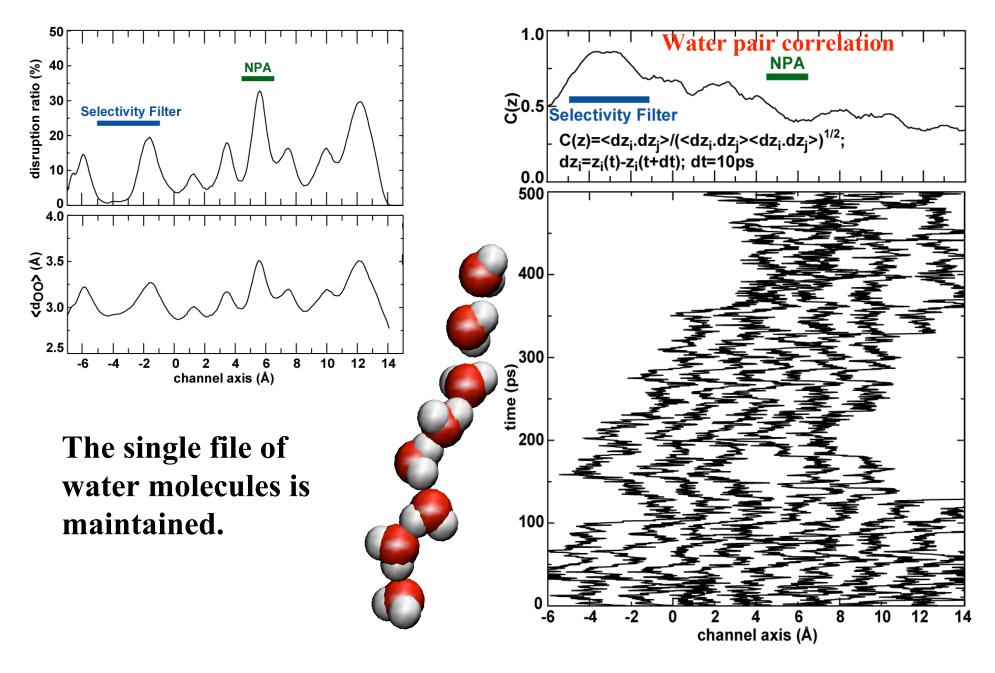
#### 5 nanosecond Simulation



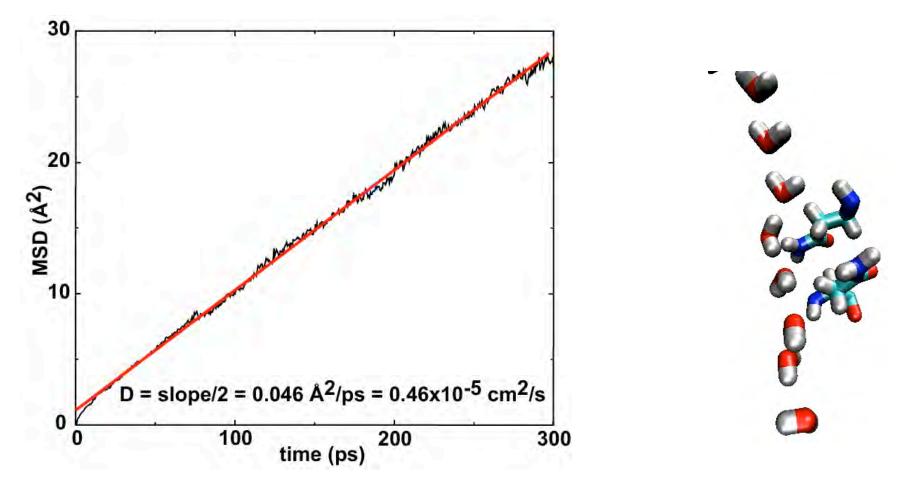
7-8 water molecules in each channel



#### **Correlated Motion of Water in the Channel**

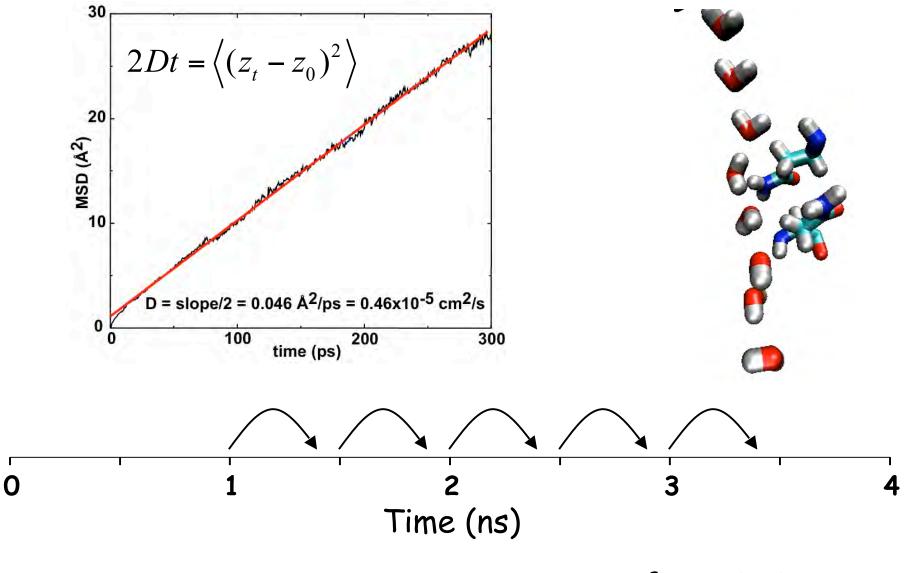


## Diffusion of Water in the channel



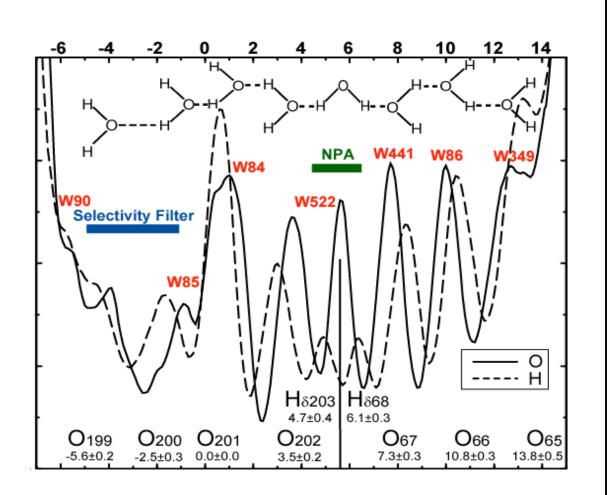
One dimensional diffusion:  $2Dt = \langle (z_t - z_0)^2 \rangle$ Experimental value for AQP1: 0.4-0.8 e-5

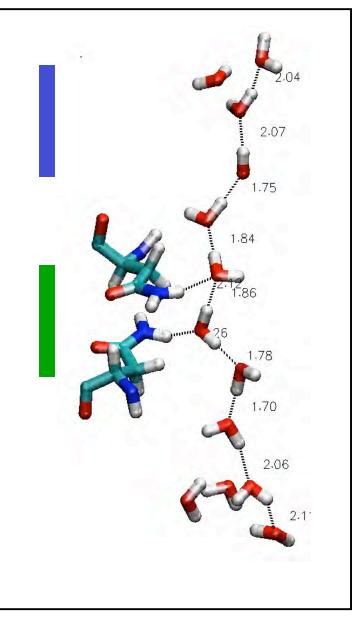
## Diffusion of Water in the channel



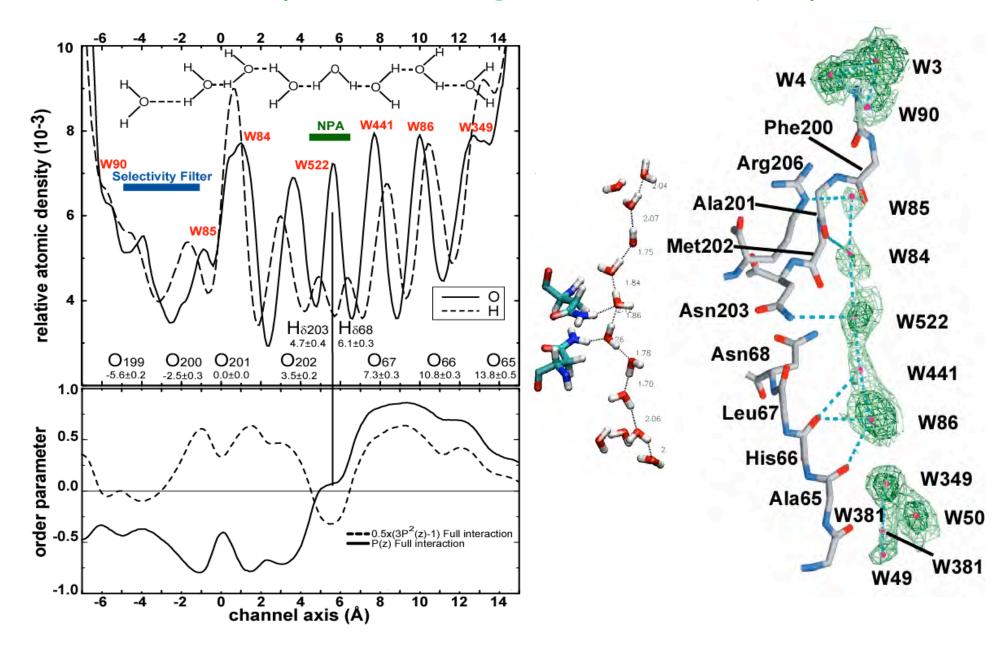
#### Improvement of statistics

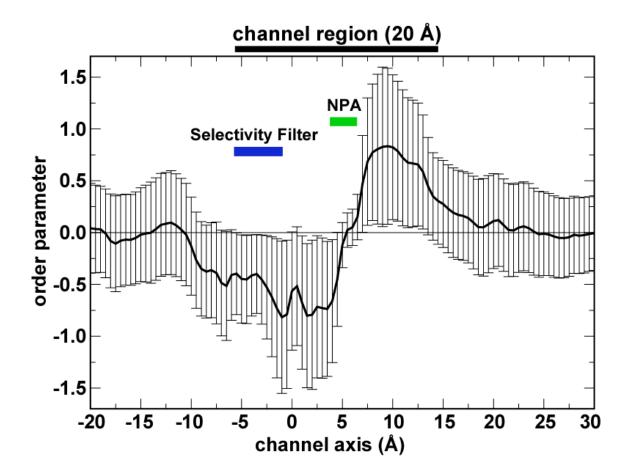
#### Water Bipolar Configuration in Aquaporins





#### Water Bipolar Configuration in Aquaporins



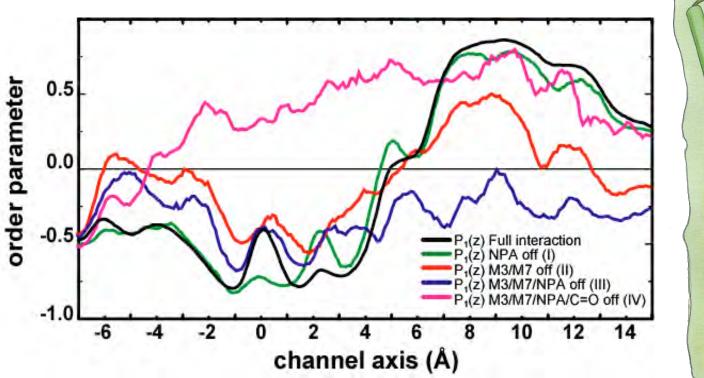


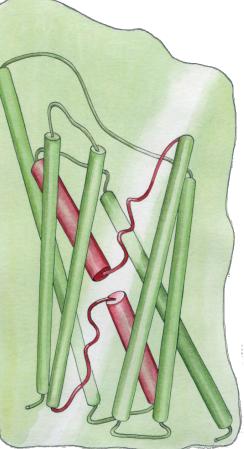
#### REMEMBER:

One of the most useful advantages of simulations over experiments is that you can modify the system as you wish: You can do modifications that are not even possible at all in reality!

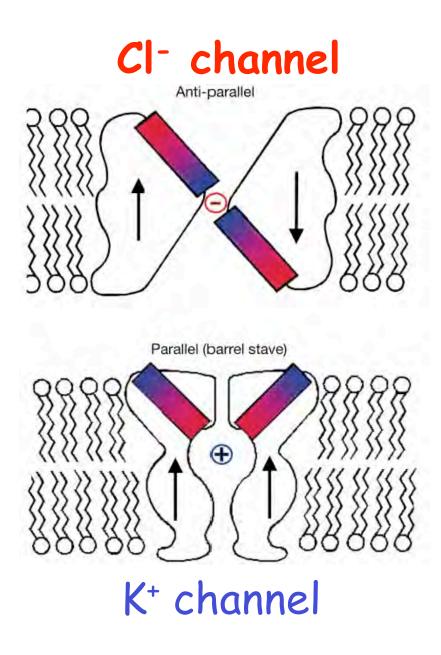
This is a powerful technique to test hypotheses developed during your simulations. Use it!

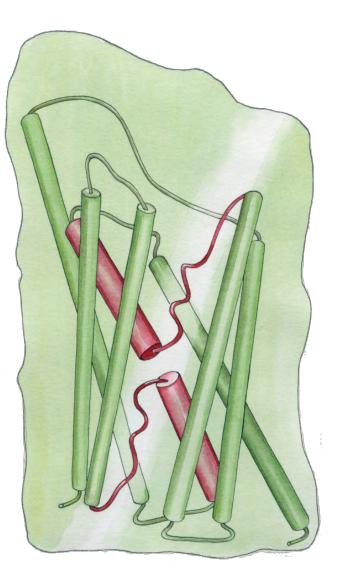
## Electrostatic Stabilization of Water Bipolar Arrangement



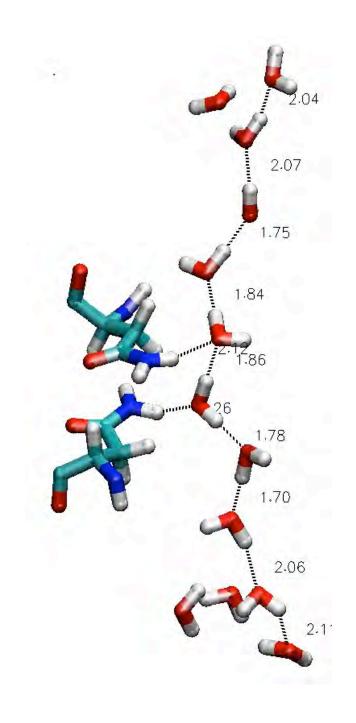


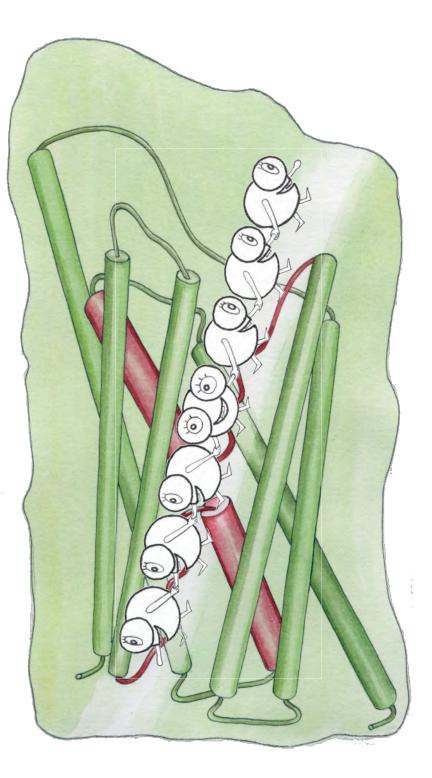
# Proton transfer through water H⁺ H⁺ H⁺ H⁺ H⁺ θ H<sup>+</sup> H<sup>+</sup> H<sup>+</sup> H-H-Η



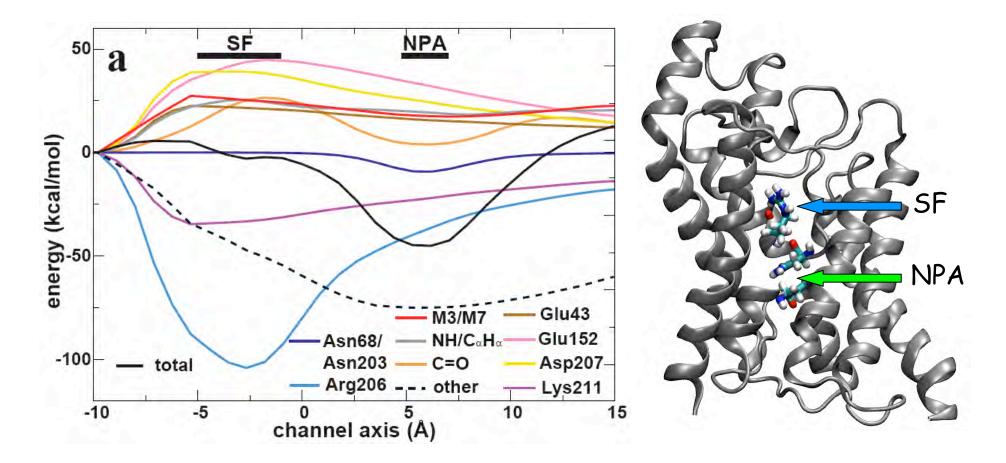


Aquaporins





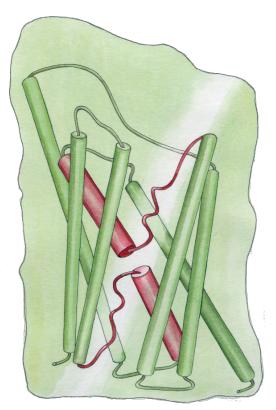
### A Complex Electrostatic Interaction



"Surprising and clearly not a hydrophobic channel"

M. Jensen, E. Tajkhorshid, K. Schulten, Biophys. J. 85, 2884 (2003)

# A Repulsive Electrostatic Force at the Center of the Channel



QM/MM MD of the behavior of an excessive proton

