

# Force Fields for Classical Molecular Dynamics simulations of Biomolecules

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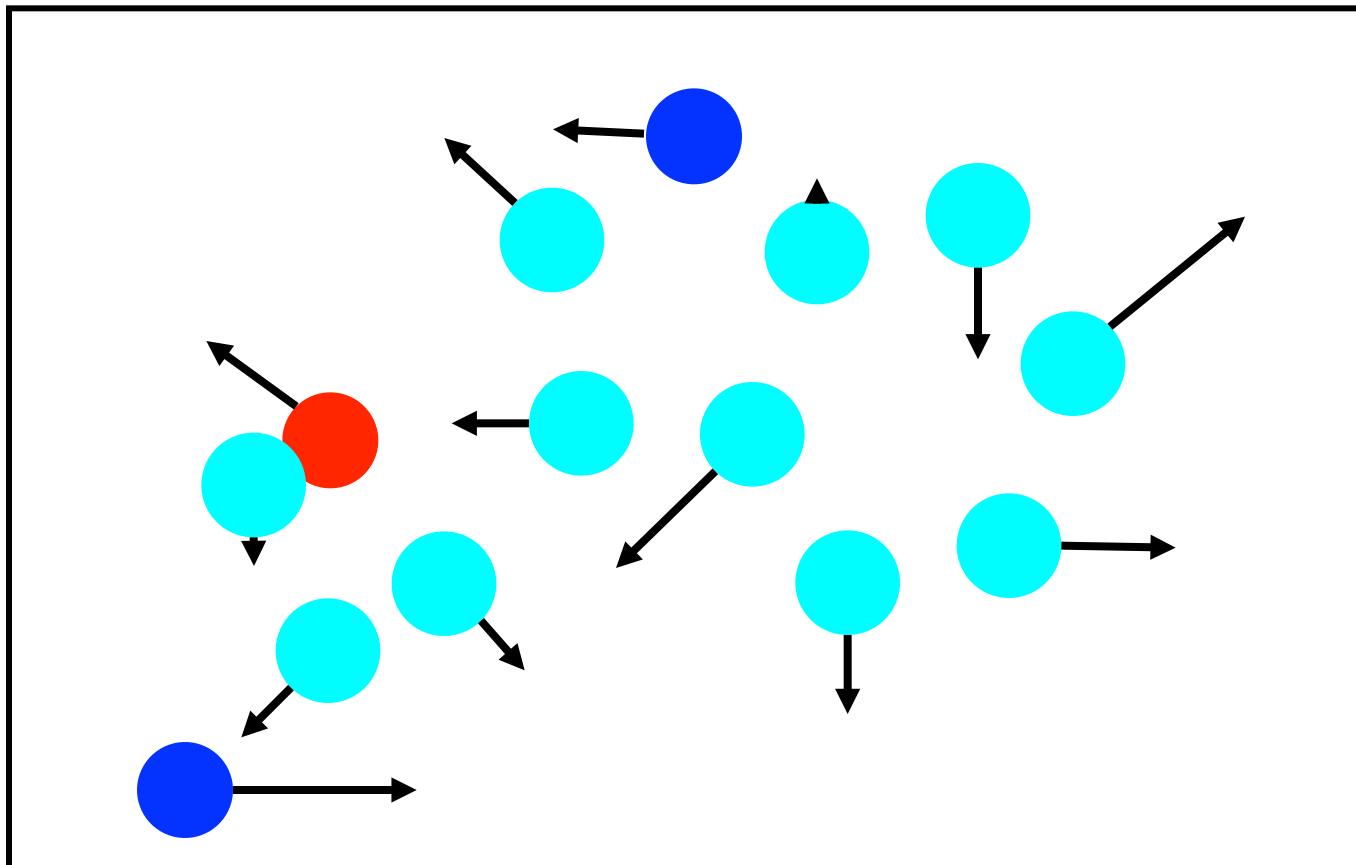
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# Classical Force Field Parameters

- Topology and structure files
- Parameter files
- Where do all the numbers needed by an MD code come from?
- Where to find these numbers and how to change them if needed.
- How to make topology files for ligands, cofactors, special amino acids, ...
- How to develop / put together missing parameters.

# Classical Molecular Dynamics

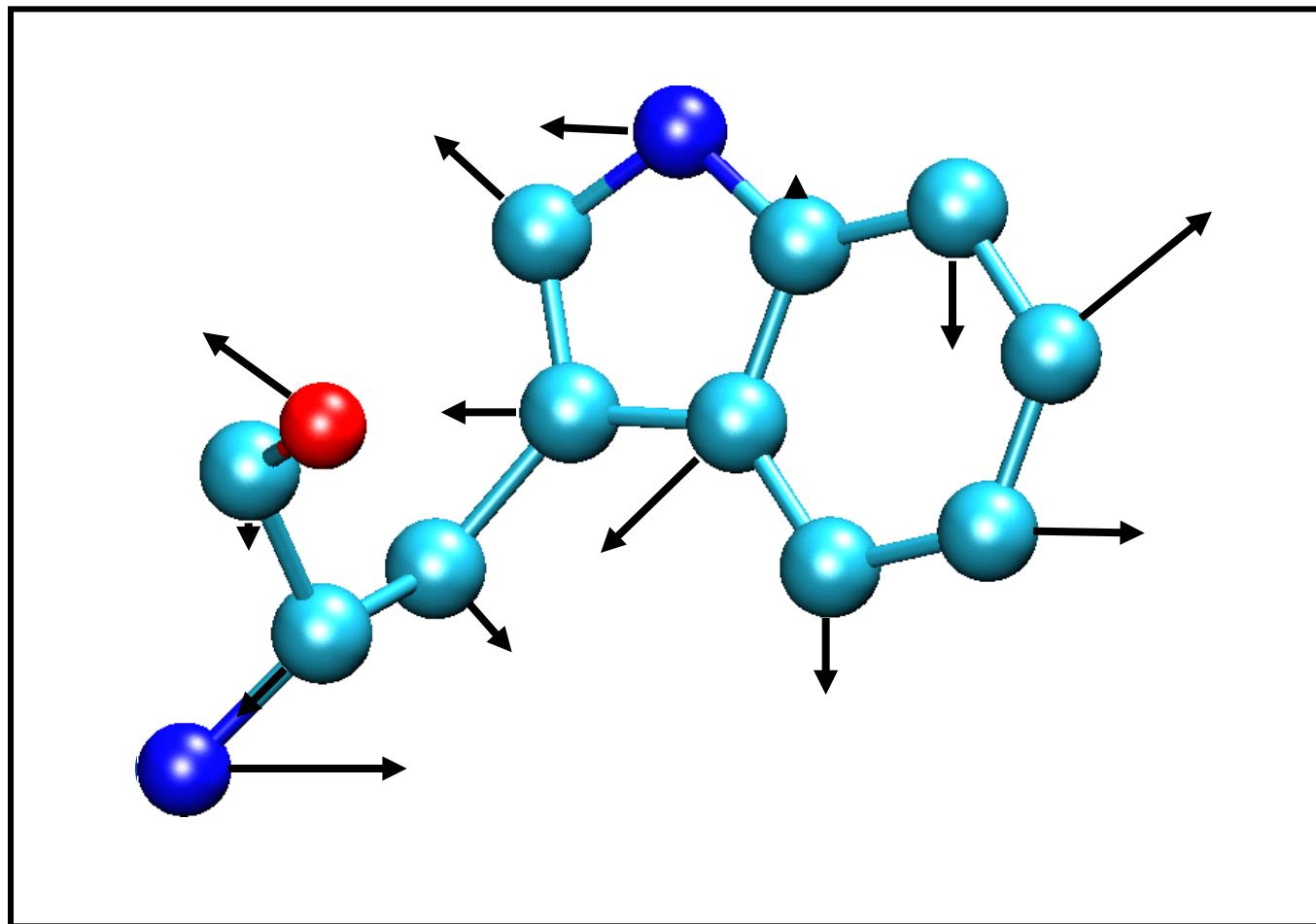


$$U(r) = \frac{1}{4\pi\epsilon_0} \frac{q_i q_j}{r_{ij}}$$

Coulomb interaction

$$U(r) = \epsilon_{ij} \left[ \left( \frac{R_{min,ij}}{r_{ij}} \right)^{12} - \left( \frac{R_{min,ij}}{r_{ij}} \right)^6 \right]$$

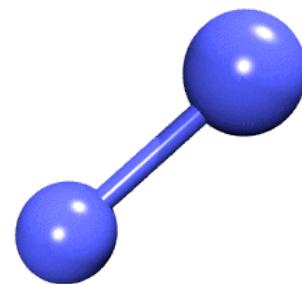
# Classical Molecular Dynamics



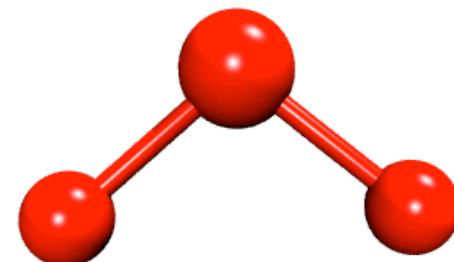
Bond definitions, atom types, atom names, parameters, ....

# Energy Terms Described in

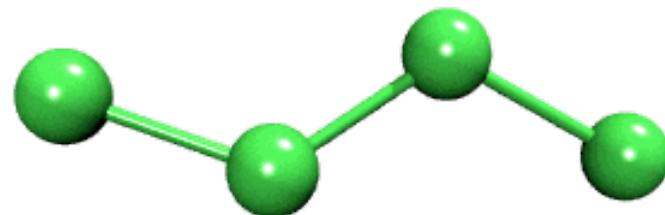
Bond



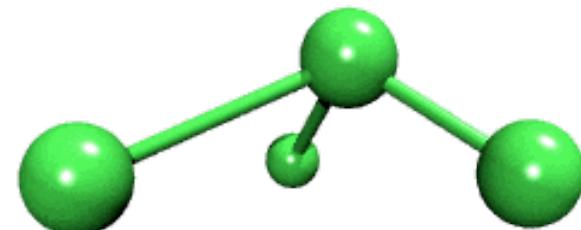
Angle



Dihedral



Improper



# The Potential Energy Function

$$U(\vec{R}) = \underbrace{\sum_{bonds} k_i^{bond} (r_i - r_0)^2}_{U_{bond}} + \underbrace{\sum_{angles} k_i^{angle} (\theta_i - \theta_0)^2}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dihed} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{dihedral}} + \underbrace{\sum_i \sum_{j \neq i} 4\epsilon_{ij} \left[ \left( \frac{\sigma_{ij}}{r_{ij}} \right)^{12} - \left( \frac{\sigma_{ij}}{r_{ij}} \right)^6 \right] + \sum_i \sum_{j \neq i} \frac{q_i q_j}{\epsilon r_{ij}}}_{U_{nonbond}}$$

$U_{bond}$  = oscillations about the equilibrium bond length

$U_{angle}$  = oscillations of 3 atoms about an equilibrium bond angle

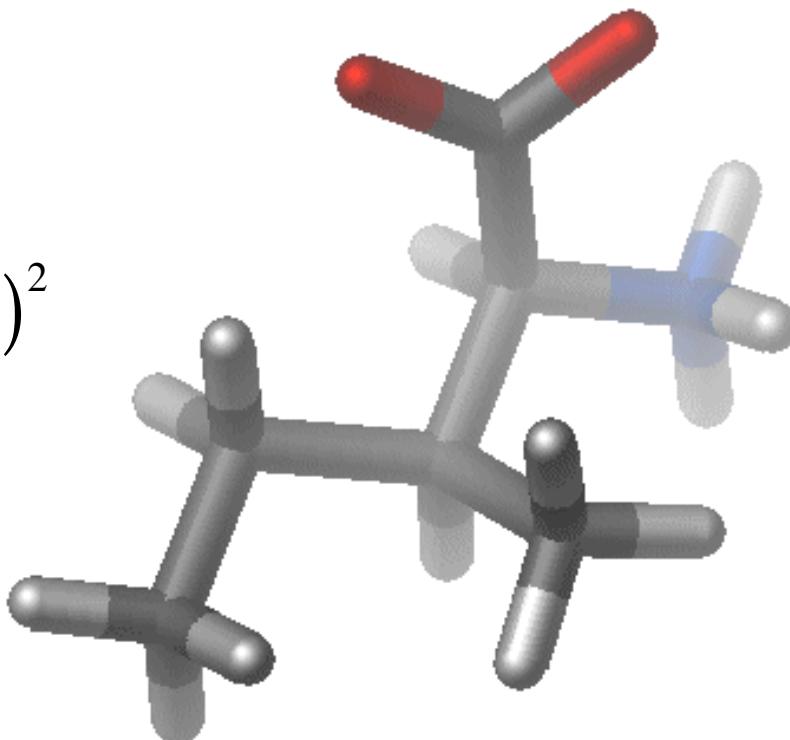
$U_{dihedral}$  = torsional rotation of 4 atoms about a central bond

$U_{nonbond}$  = non-bonded energy terms (electrostatics and Lenard-Jones)

# Interactions between bonded atoms

$$V_{angle} = K_\theta (\theta - \theta_o)^2$$

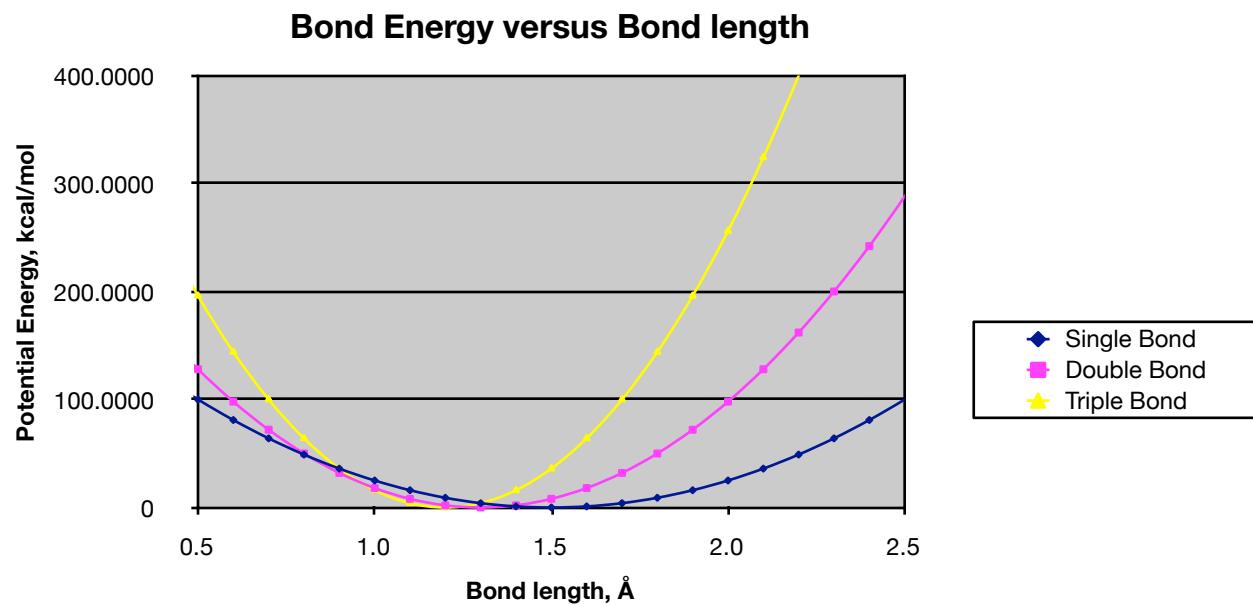
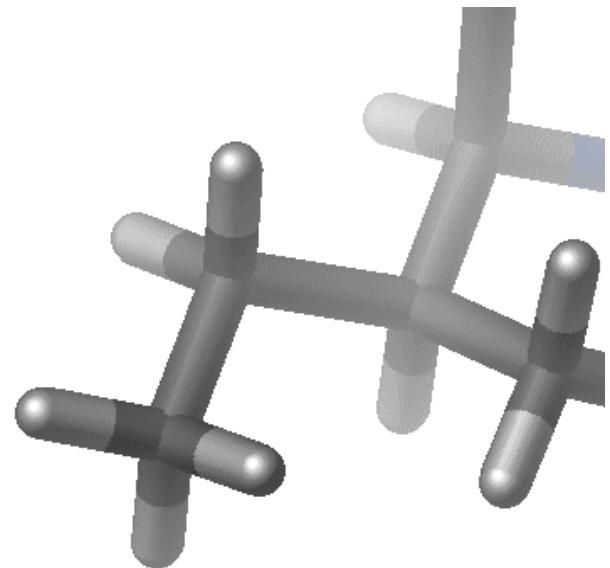
$$V_{bond} = K_b (b - b_o)^2$$



$$V_{dihedral} = K_\phi (1 + \cos(n\phi - \delta))$$

$$V_{bond} = K_b(b - b_o)^2$$

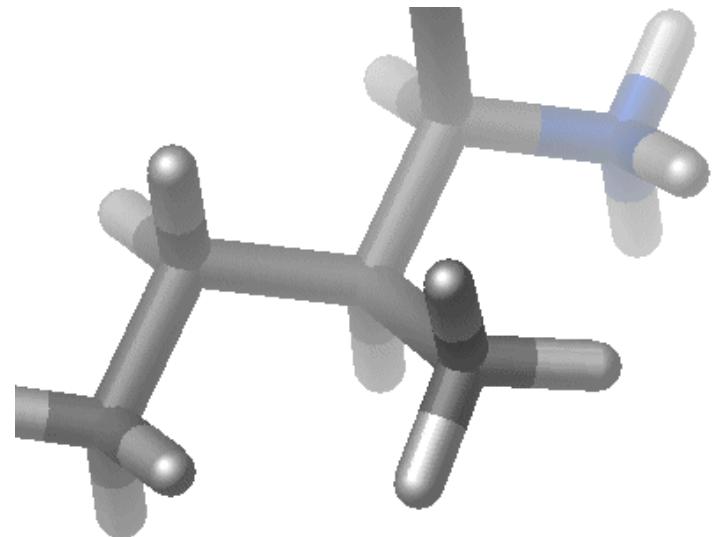
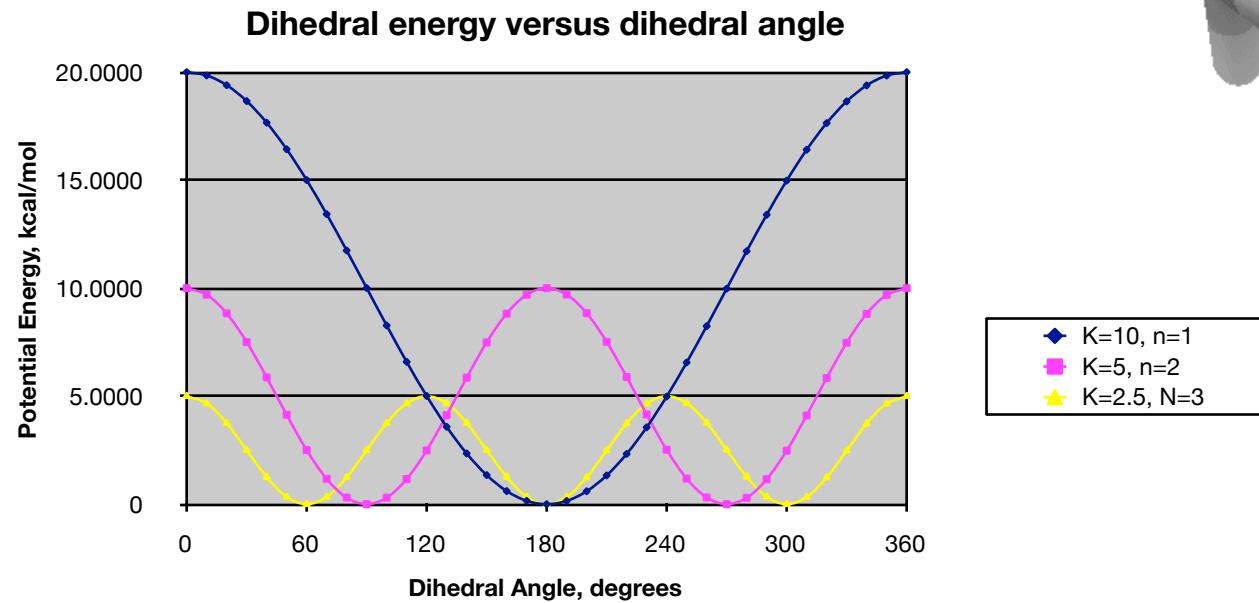
Chemical type	$K_{bond}$	$b_o$
C-C	100 kcal/mole/Å <sup>-2</sup>	1.5 Å
C=C	200 kcal/mole/Å <sup>-2</sup>	1.3 Å
C≡C	400 kcal/mole/Å <sup>-2</sup>	1.2 Å



*Bond angles* and *improper* terms have similar quadratic forms, but with softer spring constants. The force constants can be obtained from vibrational analysis of the molecule (experimentally or theoretically).

# Dihedral Potential

$$V_{dihedral} = K_\phi (1 + \cos(n\phi - \delta))$$



$$\delta = 0^\circ$$

# Nonbonded Parameters

$$\sum_{\text{non-bonded}} \frac{q_i q_j}{4\pi D r_{ij}} + \epsilon_{ij} \left[ \left( \frac{R_{min,ij}}{r_{ij}} \right)^{12} - \left( \frac{R_{min,ij}}{r_{ij}} \right)^6 \right]$$

$q_i$ : partial atomic charge

$D$ : dielectric constant

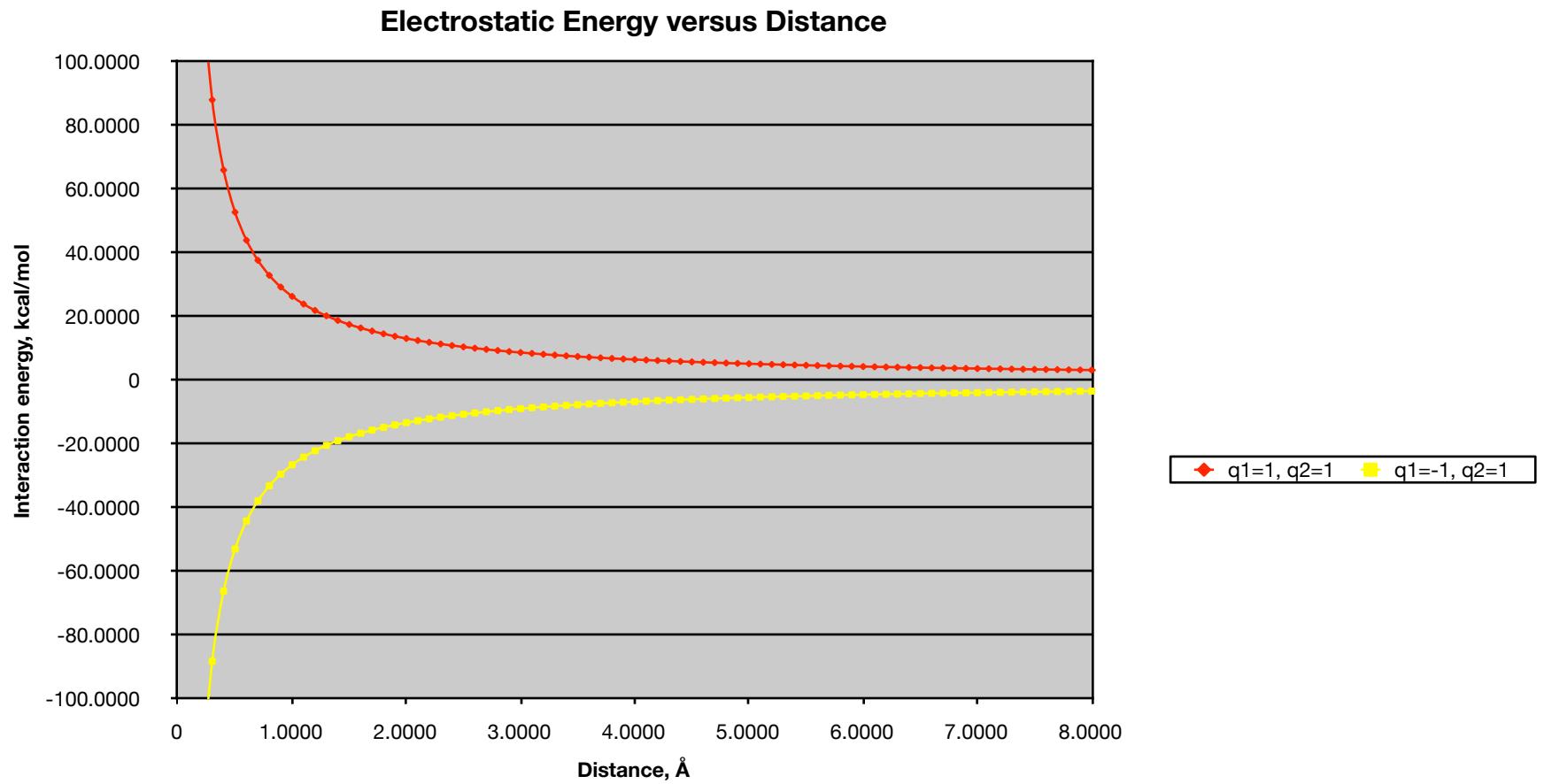
$\epsilon$ : Lennard-Jones (LJ, vdW) well-depth

$R_{min}$ : LJ radius ( $R_{min}/2$  in CHARMM)

Combining rules (CHARMM, Amber)

$$R_{min,i,j} = R_{min,i} + R_{min,j}$$

$$\epsilon_{i,j} = \text{SQRT}(\epsilon_i * \epsilon_j)$$



Note that the effect is long range.

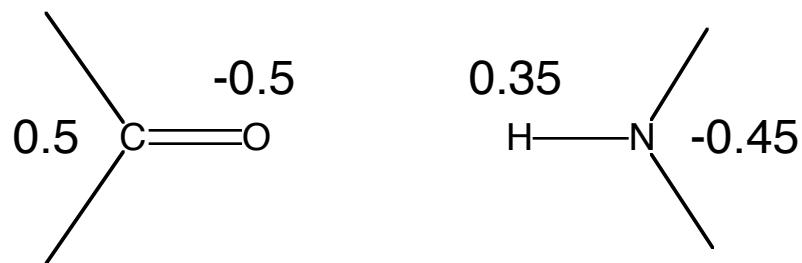
From MacKerell

# Charge Fitting Strategy

CHARMM- Mulliken\*

AMBER(ESP/RESP)

Partial atomic charges



\*Modifications based on interactions with TIP3 water

# CHARMM Potential Function

$$U(\vec{R}) = \underbrace{\sum_{bonds} k_i^{bond} (r_i - r_0)^2}_{U_{bond}} + \underbrace{\sum_{angles} k_i^{angle} (\theta_i - \theta_0)^2}_{U_{angle}} + \underbrace{\sum_{dihedrals} k_i^{dih} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{dihedral}} + \underbrace{\sum_i \sum_{j \neq i} 4\epsilon_{ij} \left[ \left( \frac{\sigma_{ij}}{r_{ij}} \right)^{12} - \left( \frac{\sigma_{ij}}{r_{ij}} \right)^6 \right]}_{U_{nonbond}} + \underbrace{\sum_i \sum_{j \neq i} \frac{q_i q_j}{\epsilon r_{ij}}}_{U_{electrostatic}}$$

The diagram illustrates the components of the CHARMM potential function. The potential energy  $U(\vec{R})$  is the sum of several terms:

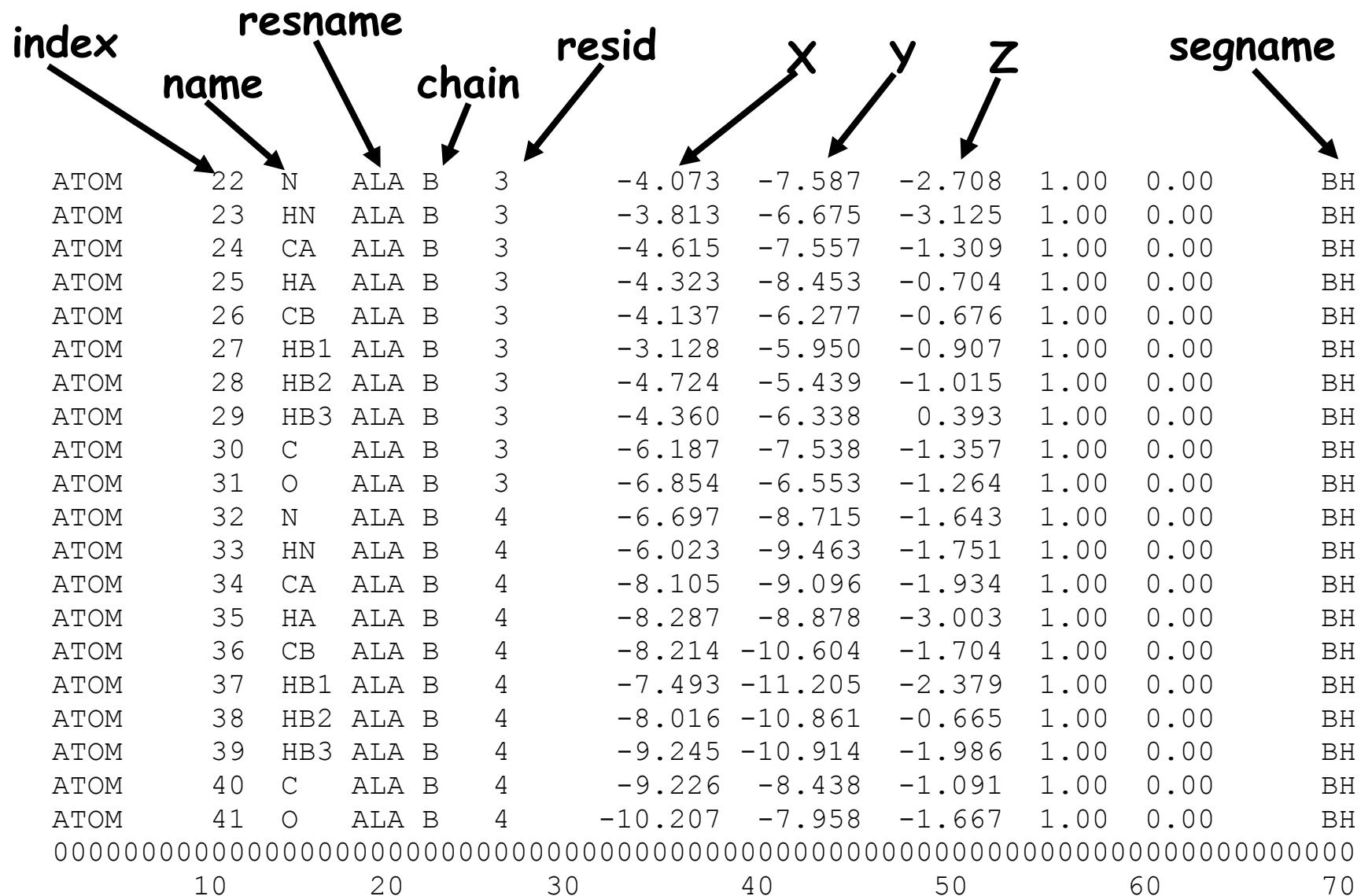
- PDB file** provides **geometry** (bonds, angles, dihedrals).
- Topology** (from PSF file) provides **parameters** for the nonbonded interaction term.
- Parameter file** provides parameters for the nonbonded interaction term.

Blue arrows point from the PDB file and Topology to their respective terms in the equation. Red arrows point from the Parameter file to the nonbonded interaction term.

# File Format/Structure

- The structure of a pdb file
- The structure of a psf file
- The topology file
- The parameter file
- Connection to potential energy terms

# Structure of a PDB file



>>> It is an ascii, fixed-format file <<<

“No connectivity information”

# Looking at File Structures

- PDB file
- Topology file
- PSF file
- Parameter file

# Parameter Optimization Strategies

**Check if it has been parameterized by somebody else**

Literature

Google

**Minimal optimization**

By analogy (direct transfer of known parameters)

Quick, starting point

**Maximal optimization**

Time-consuming

Requires appropriate experimental and target data

**Choice based on goal of the calculations**

Minimal

database screening

NMR/X-ray structure determination

Maximal

free energy calculations, mechanistic studies,  
subtle environmental effects

# Getting Started

- Identify previously parameterized compounds
- Access topology information – assign atom types, connectivity, and charges – **annotate changes**

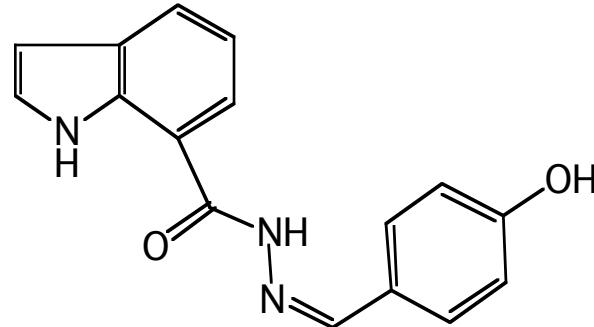
## CHARMM topology (parameter files)

top\_all22\_model.inp (par\_all22\_prot.inp)  
top\_all22\_prot.inp (par\_all22\_prot.inp)  
top\_all22\_sugar.inp (par\_all22\_sugar.inp)  
top\_all27\_lipid.rtf (par\_all27\_lipid.prm)  
top\_all27\_na.rtf (par\_all27\_na.prm)  
top\_all27\_na\_lipid.rtf (par\_all27\_na\_lipid.prm)  
top\_all27\_prot\_lipid.rtf (par\_all27\_prot\_lipid.prm)  
top\_all27\_prot\_na.rtf (par\_all27\_prot\_na.prm)  
toph19.inp (param19.inp)

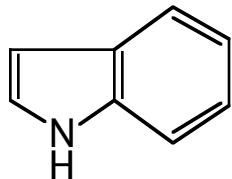
NA and lipid force fields have new LJ parameters for the alkanes, representing increased optimization of the protein alkane parameters. Tests have shown that these are compatible (e.g. in protein-nucleic acid simulations). For new systems it is suggested that the new LJ parameters be used. Note that only the LJ parameters were changed; the internal parameters are identical.

[www.pharmacy.umaryland.edu/faculty/amackere/force\\_fields.htm](http://www.pharmacy.umaryland.edu/faculty/amackere/force_fields.htm)

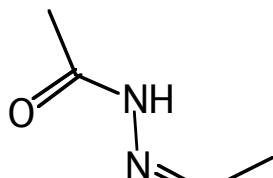
# Break Desired Compound into 3 Smaller Ones



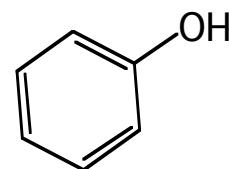
A



B



C



When creating a covalent link between model compounds move the charge on the deleted H into the carbon to maintain integer charge  
(i.e. methyl ( $q_C=-0.27$ ,  $q_H=0.09$ ) to methylene ( $q_C=-0.18$ ,  $q_H=0.09$ ))

## From top\_all22\_model.inp

```
RESI PHEN      0.00 ! phenol, adm jr.  
GROUP  
ATOM CG  CA   -0.115 !  
ATOM HG  HP    0.115 !          HD1  HE1  
GROUP          !          |  |  
ATOM CD1 CA   -0.115 !          CD1--CE1  
ATOM HD1 HP   0.115 !          //  \\  
GROUP          !          HG--CG      CZ--OH  
ATOM CD2 CA   -0.115 !          \    /    \  
ATOM HD2 HP   0.115 !          CD2==CE2    HH  
GROUP          !          |  |  
ATOM CE1 CA   -0.115 !          HD2  HE2  
ATOM HE1 HP   0.115  
GROUP  
ATOM CE2 CA   -0.115  
ATOM HE2 HP   0.115  
GROUP  
ATOM CZ  CA   0.110  
ATOM OH  OH1  -0.540  
ATOM HH  H    0.430  
BOND CD2 CG CE1 CD1 CZ CE2 CG HG CD1 HD1  
BOND CD2 HD2 CE1 HE1 CE2 HE2 CZ OH OH HH  
DOUBLE CD1 CG CE2 CD2  CZ CE1
```

Top\_all22\_model.inp contains all protein model compounds. Lipid, nucleic acid and carbohydrate model compounds are in the full topology files.

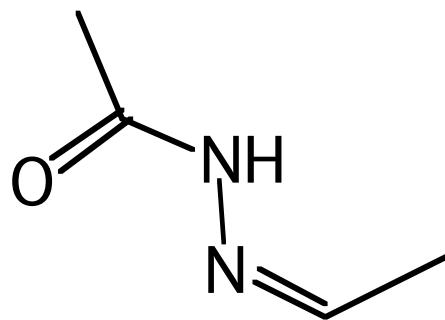
HG will ultimately be deleted. Therefore, move HG (hydrogen) charge into CG, such that the CG charge becomes 0.00 in the final compound.

Use remaining charges/atom types without any changes.

Do the same with indole

# Creation of topology for central model compound

```
RESI Mod1 ! Model compound 1
Group
ATOM C1 CT3 -0.27
ATOM H11 HA3 0.09
ATOM H12 HA3 0.09
ATOM H13 HA3 0.09
GROUP
ATOM C2 C 0.51
ATOM O2 O -0.51
GROUP
ATOM N3 NH1 -0.47
ATOM H3 H 0.31
ATOM N4 NR1 0.16 ! new atom
ATOM C5 CEL1 -0.15
ATOM H51 HEL1 0.15
ATOM C6 CT3 -0.27
ATOM H61 HA 0.09
ATOM H62 HA 0.09
ATOM H63 HA 0.09
BOND C1 H11 C1 H12 C1 H13 C1 C2 C2 O2 C2 N3 N3 H3
BOND N3 N4 C5 H51 C5 C6 C6 H61 C6 H62 C6 H63
DOUBLE N4 C5 (DOUBLE only required for MMFF)
```



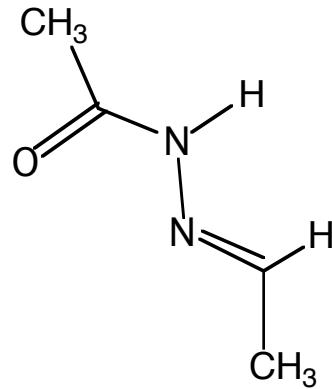
Start with alanine dipeptide.  
Note use of new aliphatic LJ parameters and, importantly, atom types.

NR1 from histidine unprotonated ring nitrogen.  
Charge (very bad) initially set to yield unit charge for the group.

Note use of large group to allow flexibility in charge optimization.

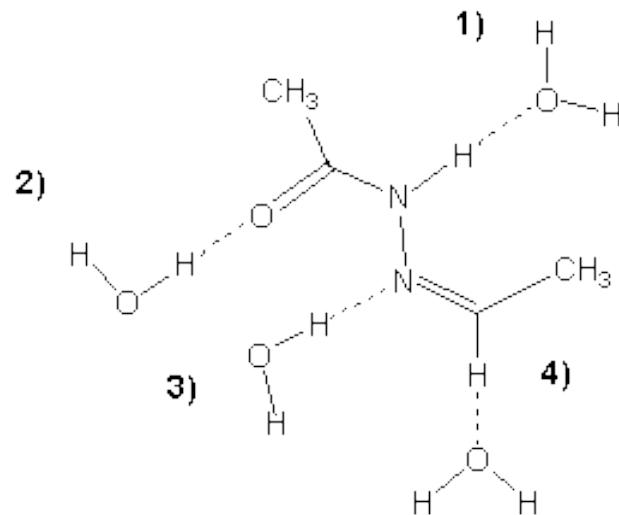
# Partial Charge Assignment

- Most important aspect for ligands
- Different force fields might take different philosophies
  - AMBER: RESP charges at the HF/6-31G level
    - Overestimation of dipole moments
    - Easier to set up
  - CHARMM: Interaction based optimization
    - TIP3P water representing the environment
    - Could be very difficult to set up
- Conformation dependence of partial charges
- Lack of polarization
- Try to be consistent within the force field
- pKa calculations for titratable residues



Starting charges??  
Mulliken population analysis  
Analogy comparison

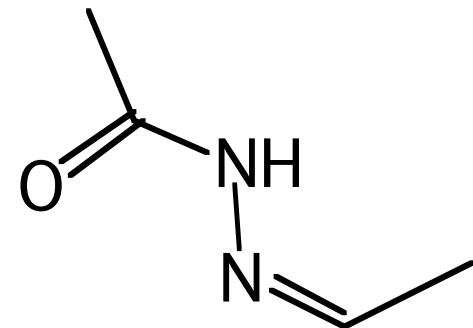
Final charges (methyl, vary  $q_C$  to maintain integer charge,  $q_H = 0.09$ )  
interactions with water (HF/6-31G\*, monohydrates!)



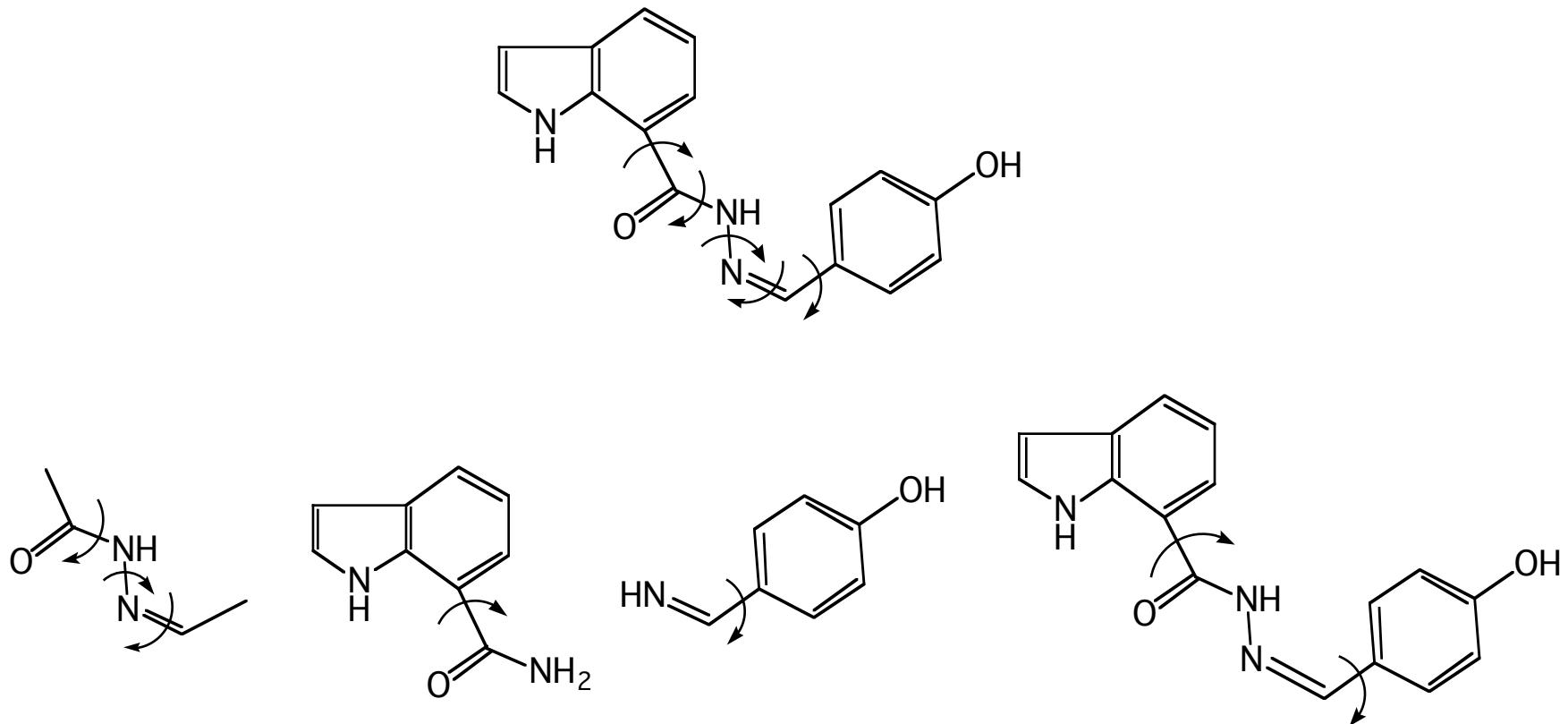
From MacKerell

## Comparison of analogy and optimized charges

Name	Type	Analogy	Optimized
C1	CT3	-0.27	-0.27
H11	HA3	0.09	0.09
H12	HA3	0.09	0.09
H13	HA3	0.09	0.09
C2	C	0.51	0.58
O2	O	-0.51	-0.50
N3	NH1	-0.47	-0.32
H3	H	0.31	0.33
N4	NR1	0.16	-0.31
C5	CEL1	-0.15	-0.25
H51	HEL1	0.15	0.29
C6	CT3	-0.27	-0.09
H61	HA	0.09	0.09
H62	HA	0.09	0.09
H63	HA	0.09	0.09



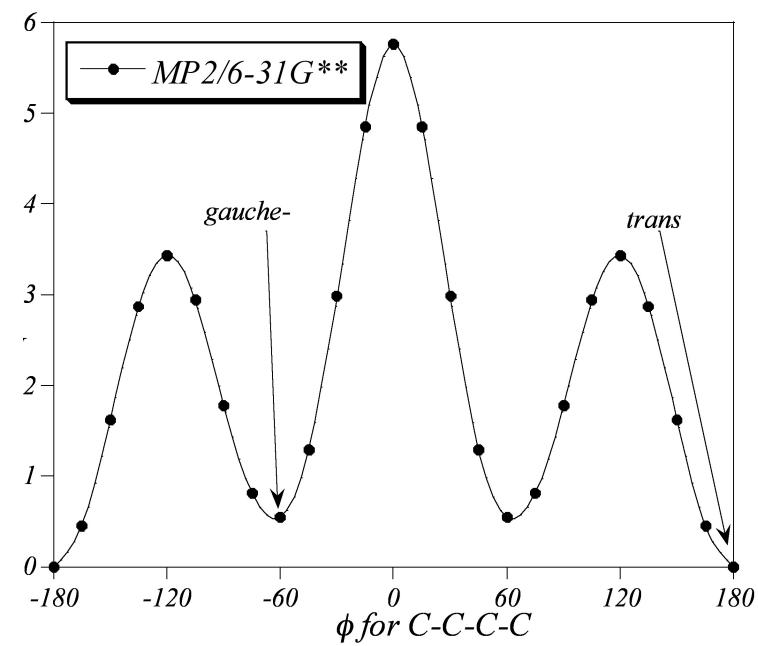
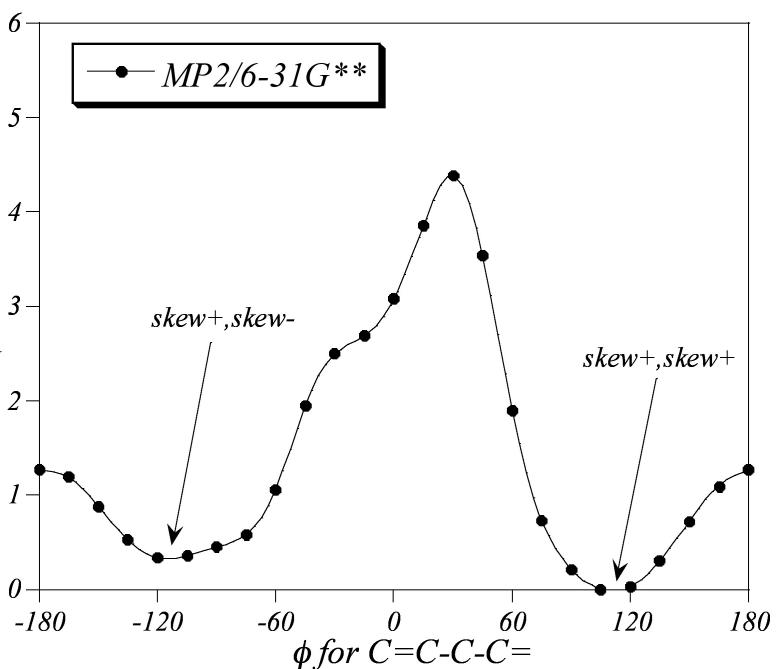
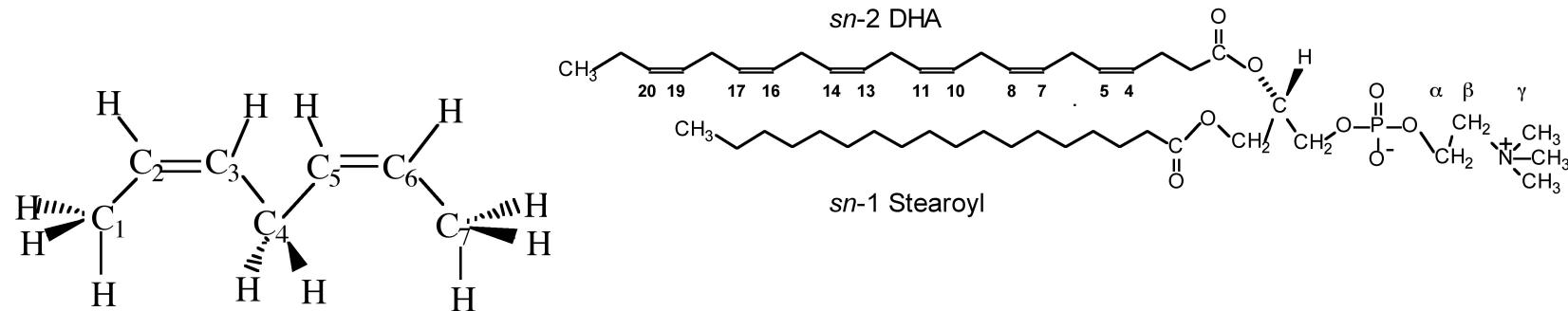
Dihedral optimization based on QM potential energy surfaces (HF/6-31G\* or MP2/6-31G\*).



From MacKerell

# Parameterization of unsaturated lipids

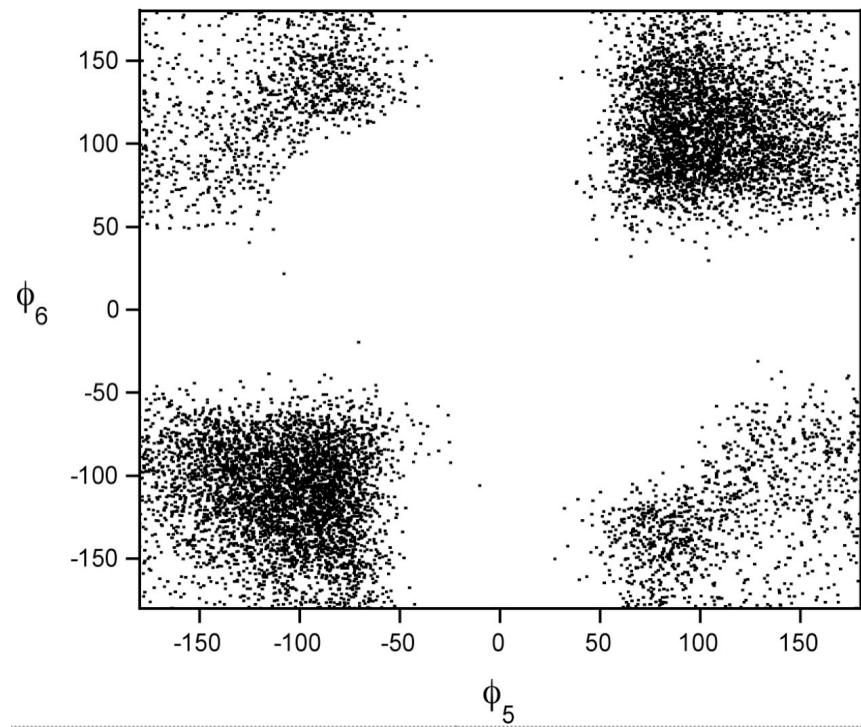
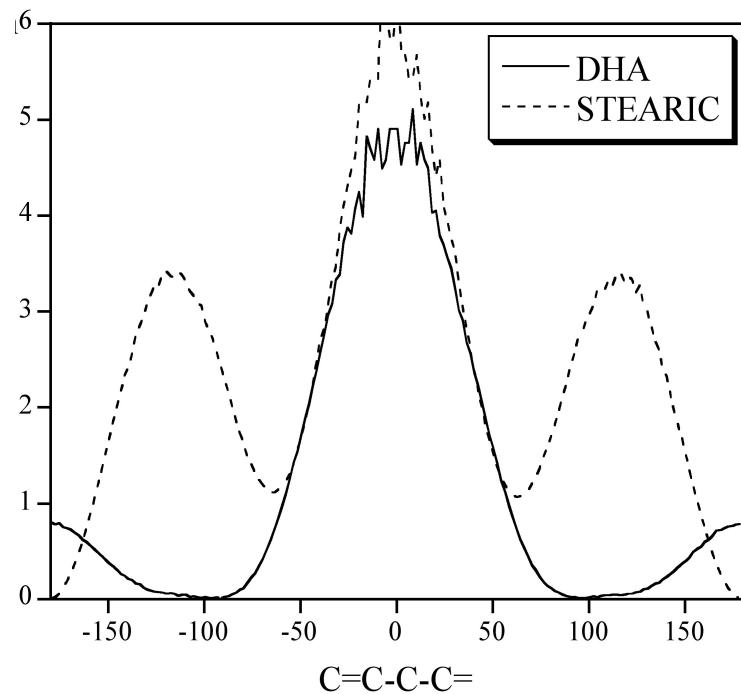
- All C=C bonds are cis, what does rotation about neighboring single bonds look like?



Courtesy of Scott Feller, Wabash College

# DHA conformations from MD

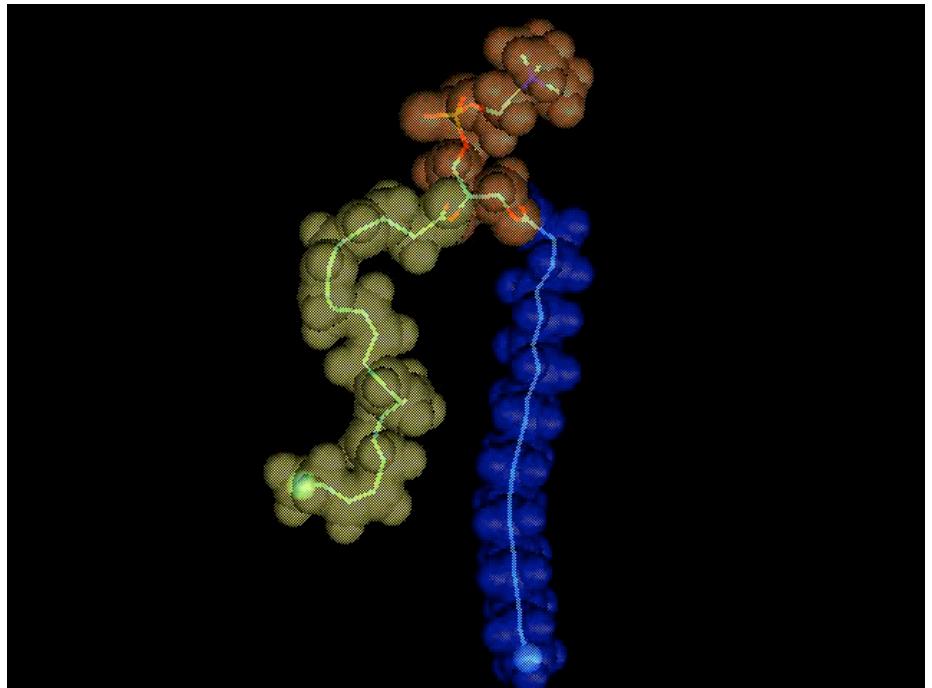
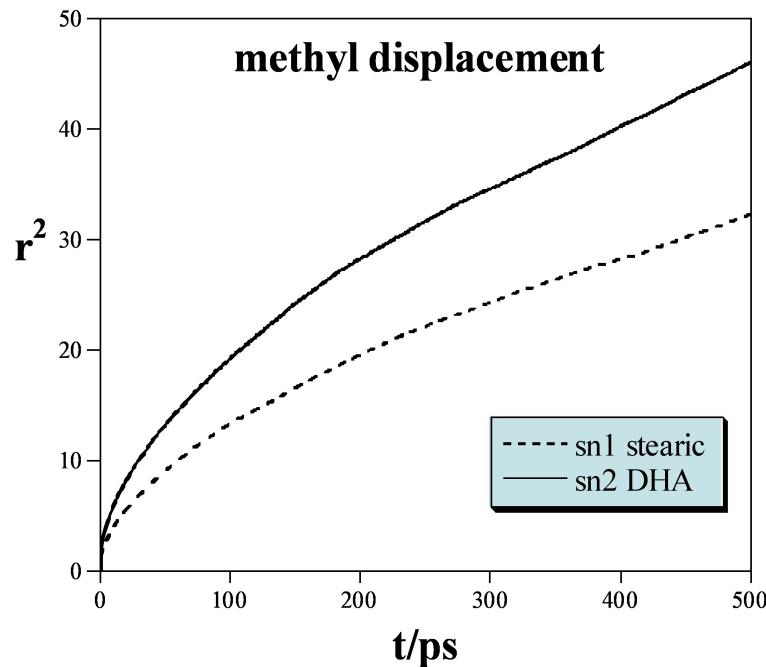
- rotational barriers are extremely small
- many conformers are accessible w/ short lifetimes



Courtesy of Scott Feller, Wabash College

# Dynamics of saturated vs. polyunsaturated lipid chains

- $sn1$  stearic acid = blue
- $sn2$  DHA = yellow
- 500 ps of dynamics

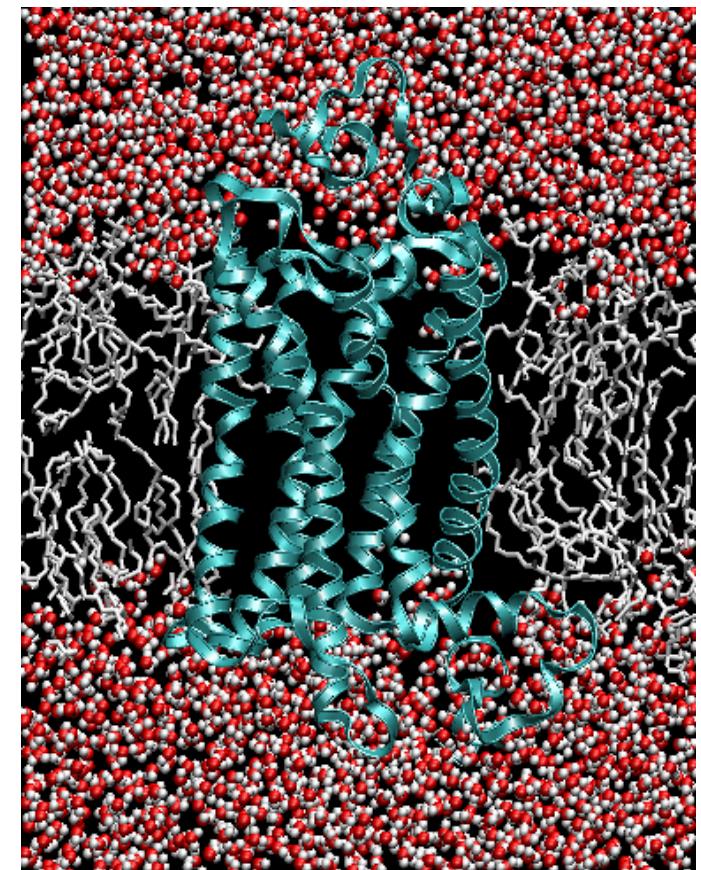
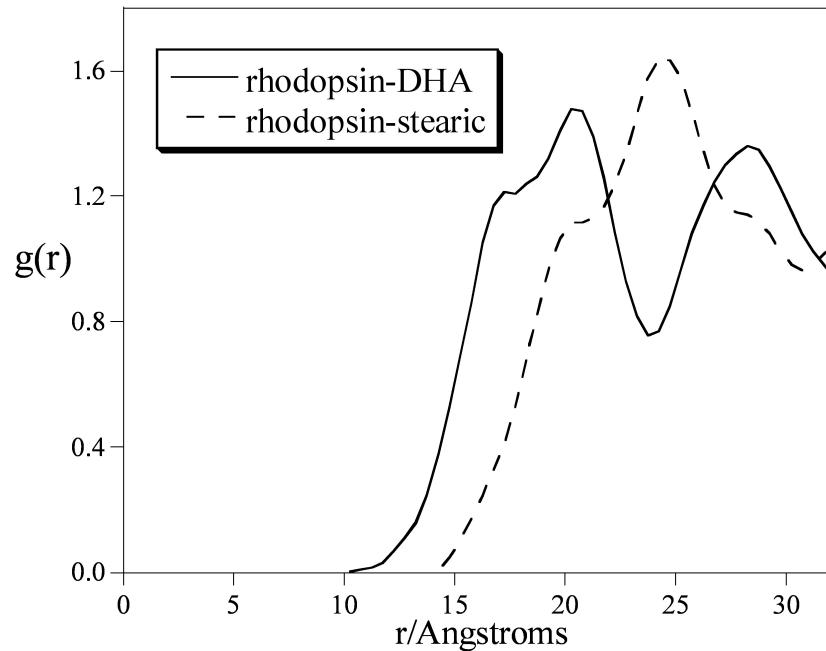


*Movie courtesy of Mauricio Carrillo Tripp*

Courtesy of Scott Feller, Wabash College

# Lipid-protein interactions

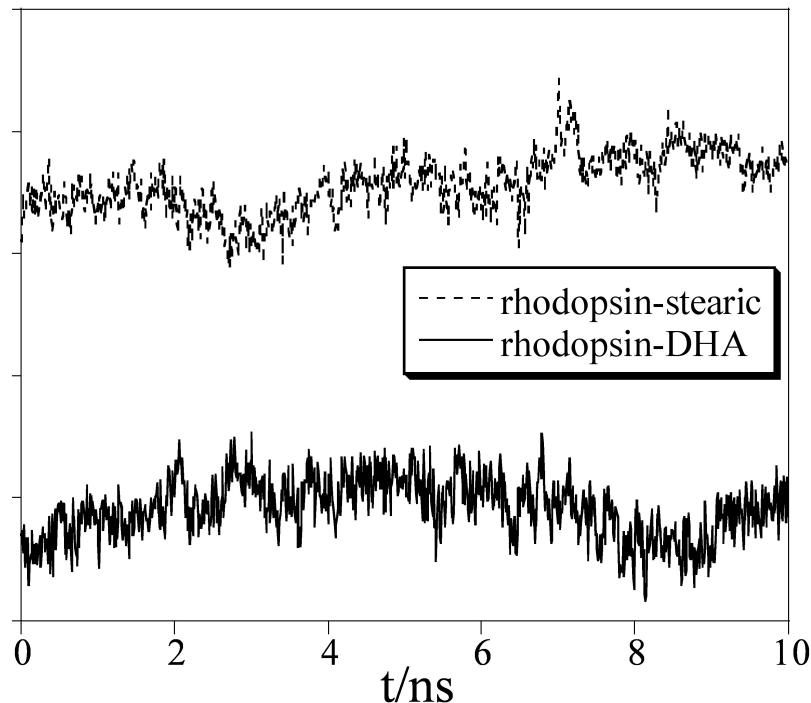
- Radial distribution around protein shows distinct layering of acyl chains



Courtesy of Scott Feller, Wabash College

# Lipid-protein interactions

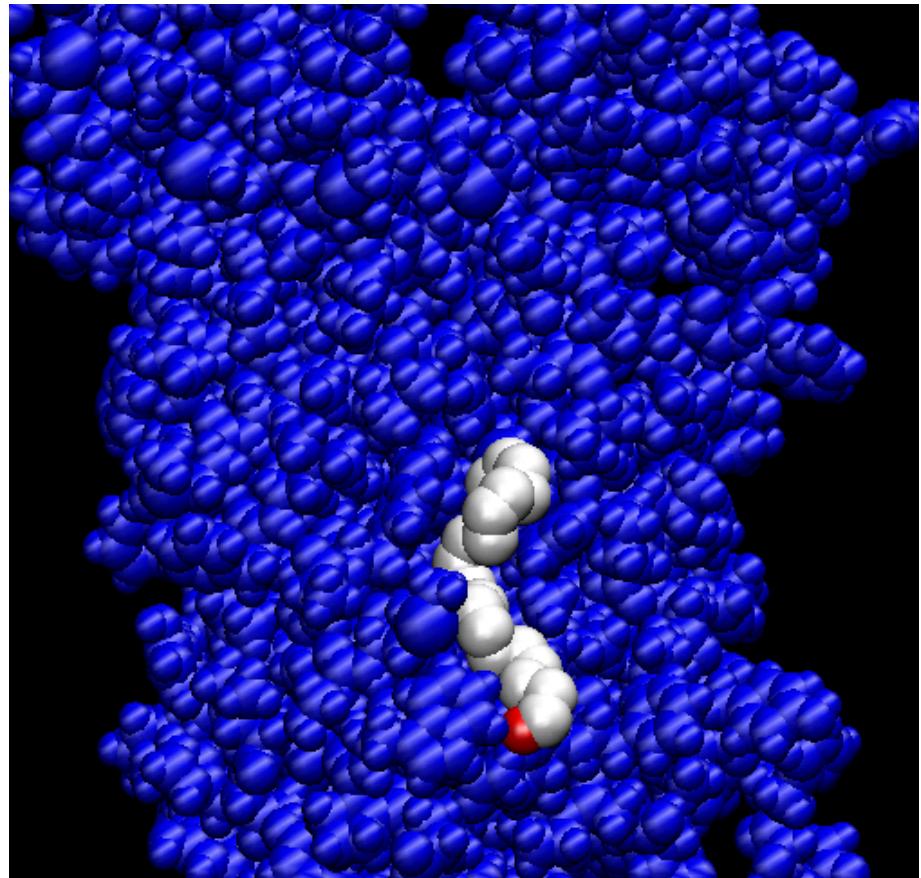
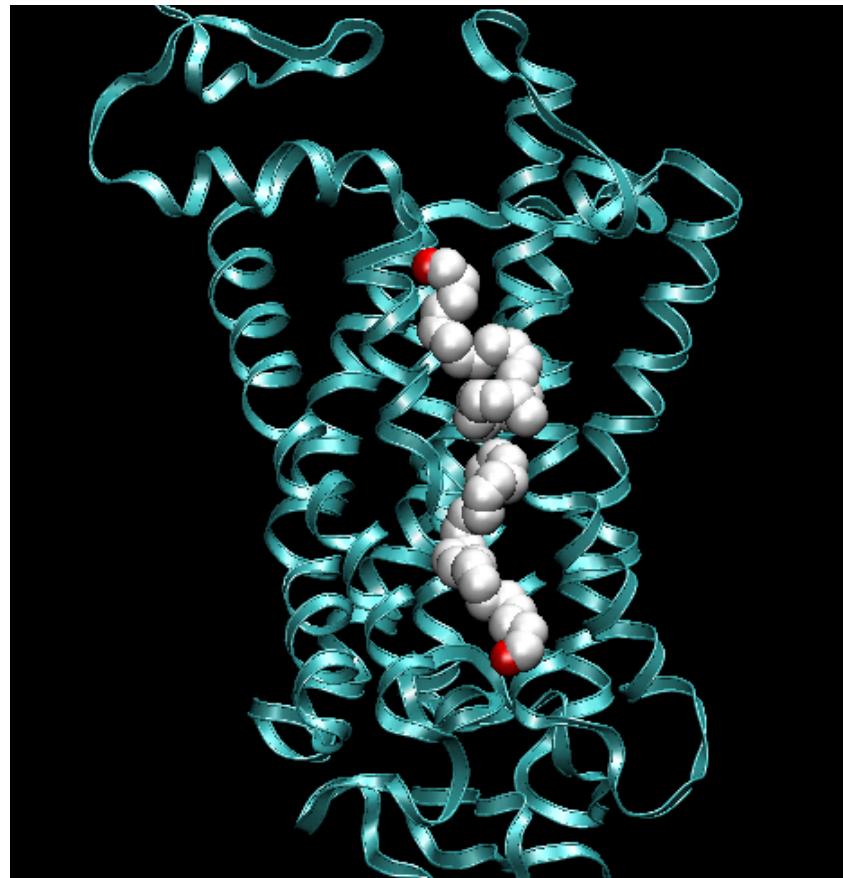
- Decomposition of non-bonded interaction shows rhodopsin is strongly attracted to unsaturated chain
- All hydrophobic residues are stabilized by DHA



<u>resname</u>	$U_{DHA}$	$U_{stearic}$	<u>ratio</u>
PHE	-44.9	-22.6	2.0
ILE	-30.0	-10.1	3.0
VAL	-24.0	-9.6	2.5
LEU	-23.1	-13.0	1.8
MET	-22.8	-9.7	2.4
TYR	-18.6	-10.4	1.8
ALA	-11.4	-3.0	3.8
TRP	-10.3	-2.4	4.2

Courtesy of Scott Feller, Wabash College

# Origin of protein:DHA attraction



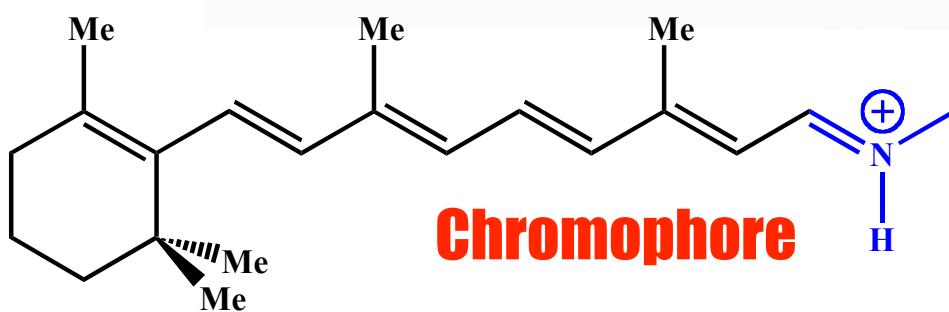
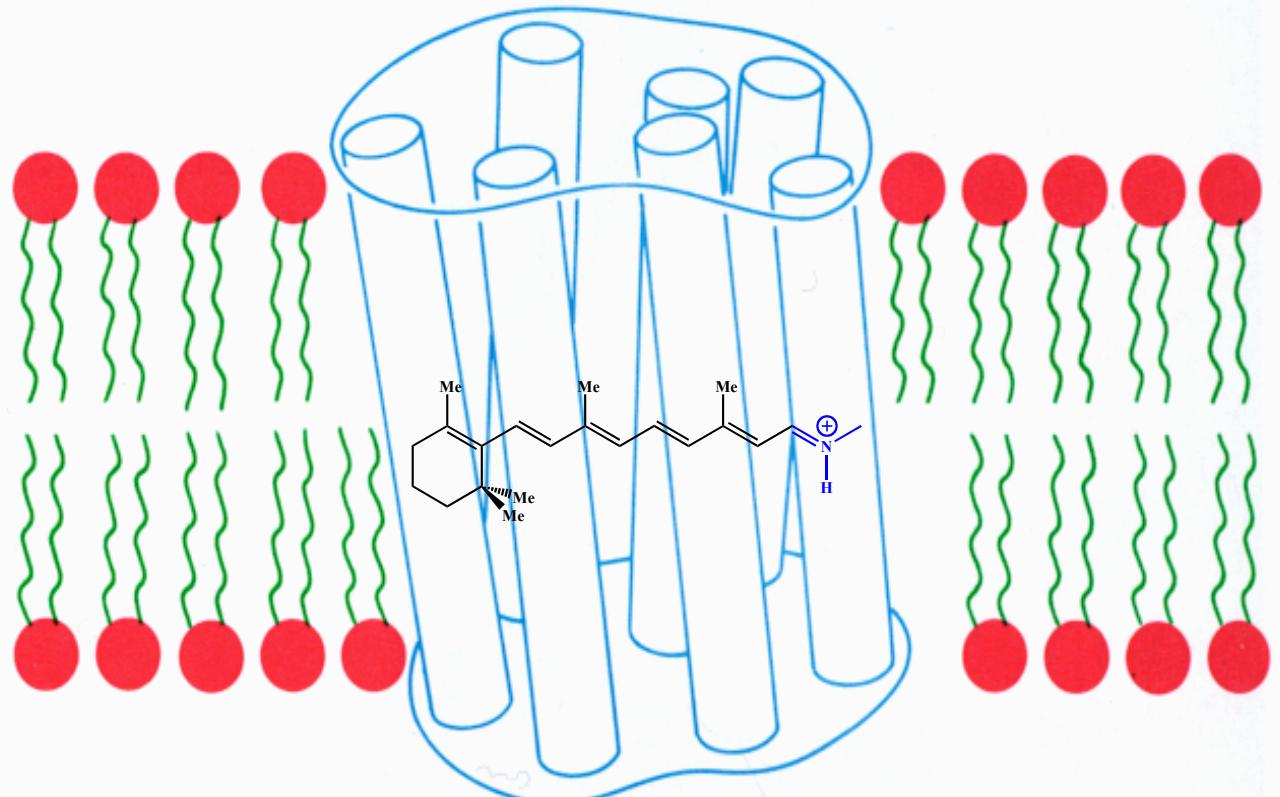
- Flexibility of the DHA chain allows solvation of the rough protein surface to occur with little intra-molecular energy cost

Courtesy of Scott Feller, Wabash College

# Major Recent Developments

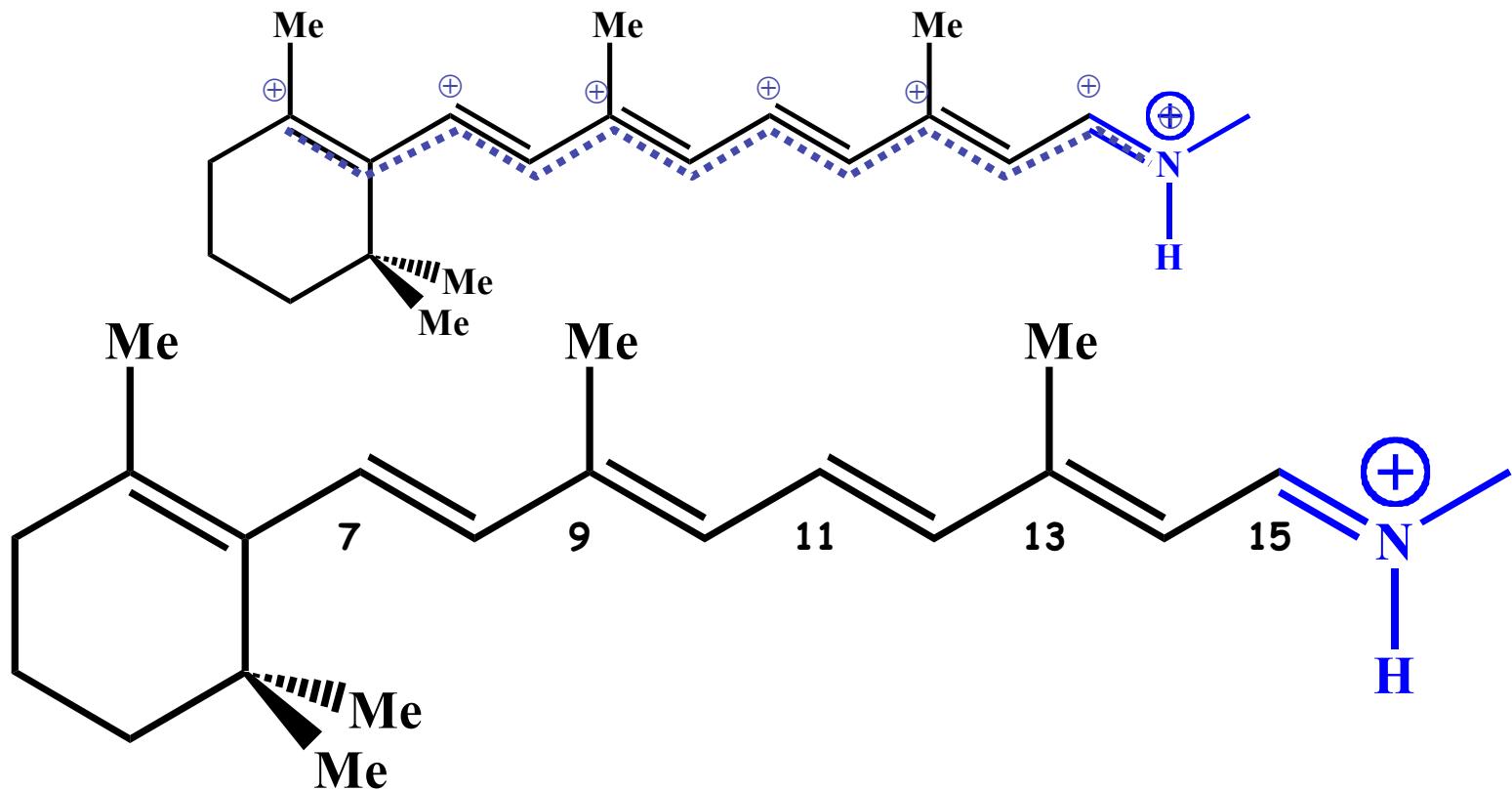
- New set of lipid force field parameters for CHARMM (CHARMM32<sup>+</sup>)
  - Pastor, B. Brooks, MacKerell
- Polarizable force field
  - Roux, MacKerell

# Retinal Proteins -- Rhodopsins

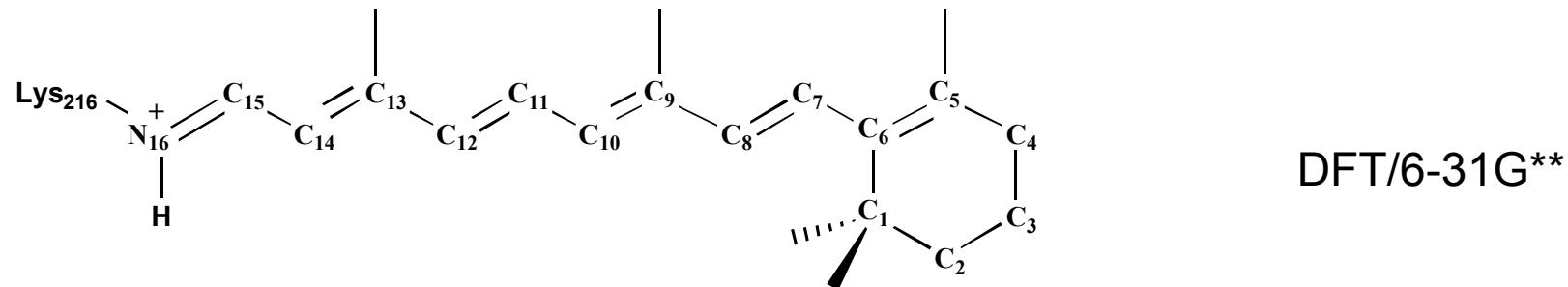


- Covalently linked to a lysine
- Usually protonated *Schiff base*
- all-trans and 11-cis isomers

# Unconventional chemistry



# Isomerization Barriers in retinal



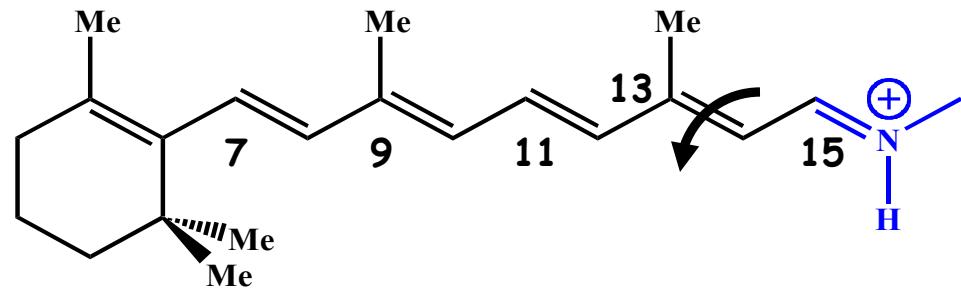
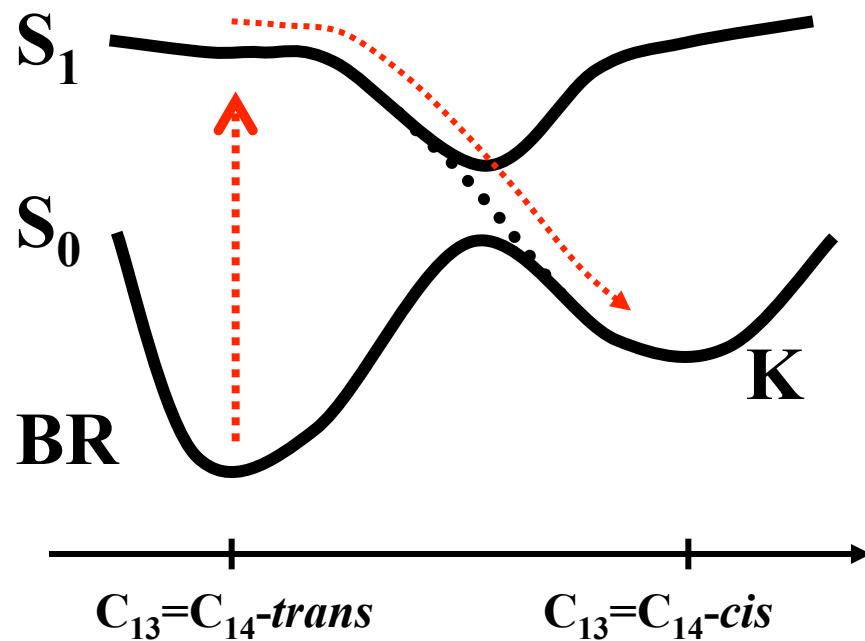
**TABLE 2** The parameter set B used for the torsional potentials of the main polyene chain of the retinal Schiff base

$\phi_i$	$k_i$ (kcal/mol)*	$n_i$	$\delta_i$ (deg)
C <sub>5</sub> =C <sub>6</sub> =C <sub>7</sub> =C <sub>8</sub>	11.24	2.0	180.00
C <sub>6</sub> =C <sub>7</sub> =C <sub>8</sub> =C <sub>9</sub>	39.98	2.0	180.00
C <sub>7</sub> =C <sub>8</sub> =C <sub>9</sub> =C <sub>10</sub>	17.03	2.0	180.00
C <sub>8</sub> =C <sub>9</sub> =C <sub>10</sub> =C <sub>11</sub>	37.28	2.0	180.00
C <sub>9</sub> =C <sub>10</sub> =C <sub>11</sub> =C <sub>12</sub>	22.50	2.0	180.00
C <sub>10</sub> =C <sub>11</sub> =C <sub>12</sub> =C <sub>13</sub>	35.08	2.0	180.00
C <sub>11</sub> =C <sub>12</sub> =C <sub>13</sub> =C <sub>14</sub>	28.30	2.0	180.00
C <sub>12</sub> =C <sub>13</sub> =C <sub>14</sub> =C <sub>15</sub>	29.46	2.0	180.00
C <sub>13</sub> =C <sub>14</sub> =C <sub>15</sub> =N <sub>16</sub>	30.43	2.0	180.00
C <sub>14</sub> =C <sub>15</sub> =N <sub>16</sub> -C <sub>s</sub>	28.76	2.0	180.00

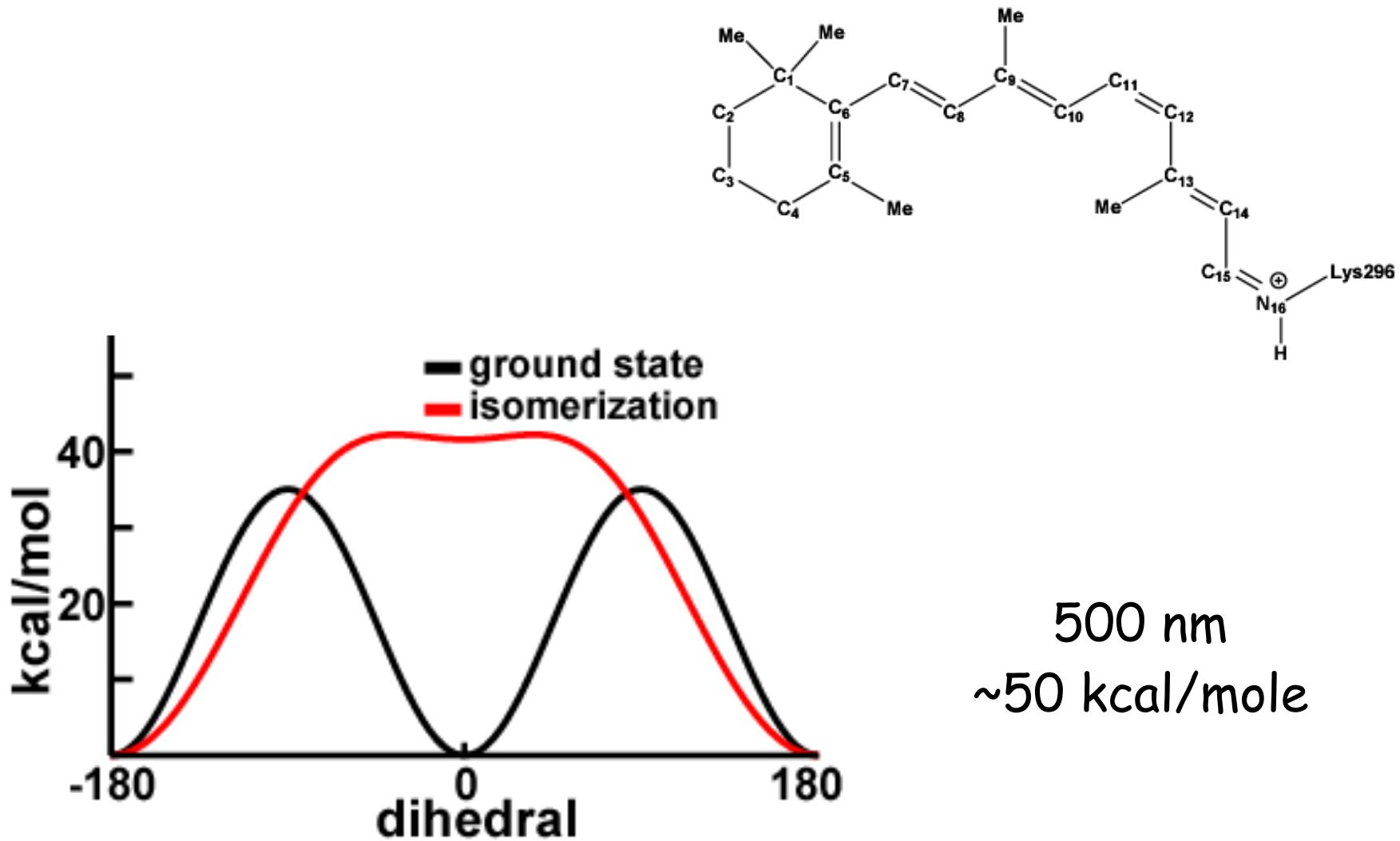
Tajkhorshid et al., 1999.

\* $E_i^{\text{dihedral}} = (1/2)k_i[1 + \cos(n_i\varphi_i - \delta_i)]$ .

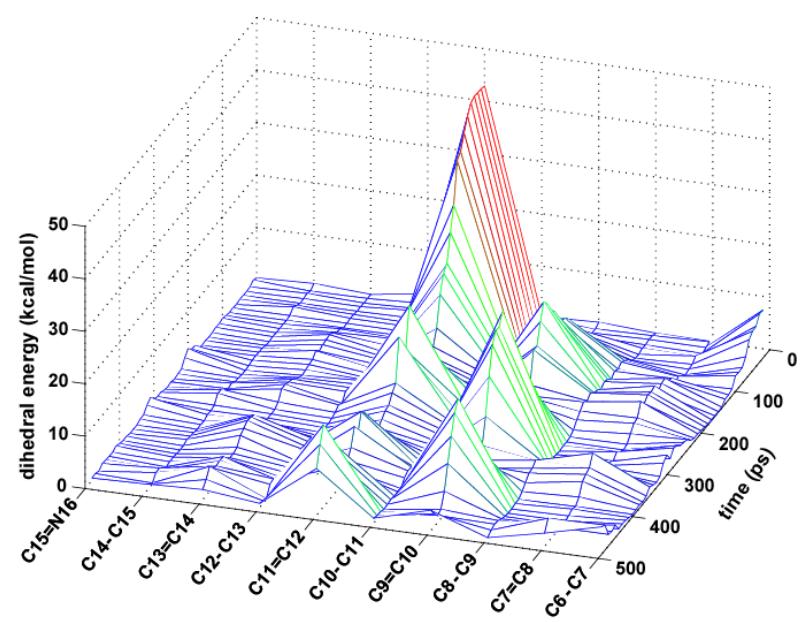
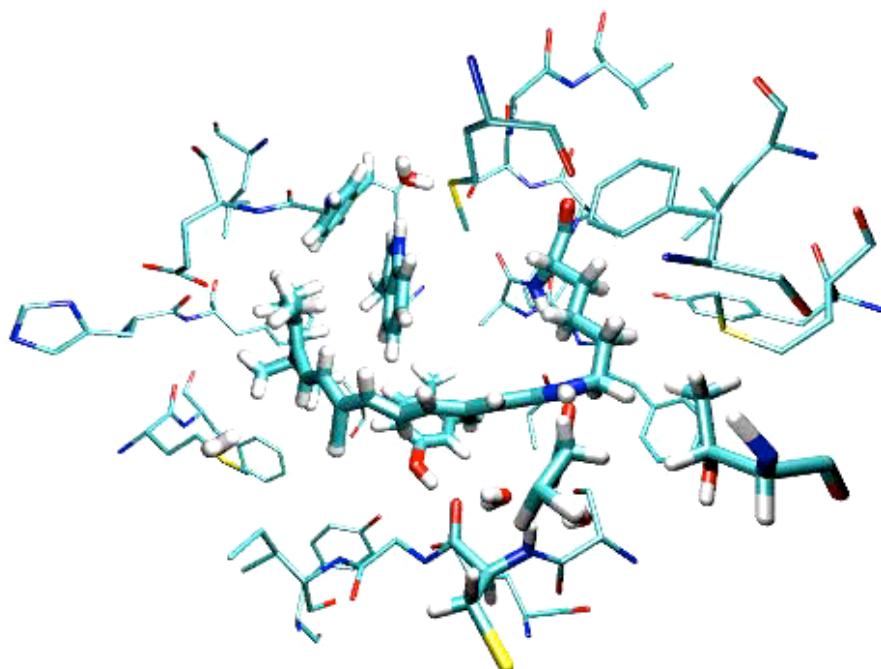
# Coupling of electronic excitation and conformational change in bR



# Inducing isomerization

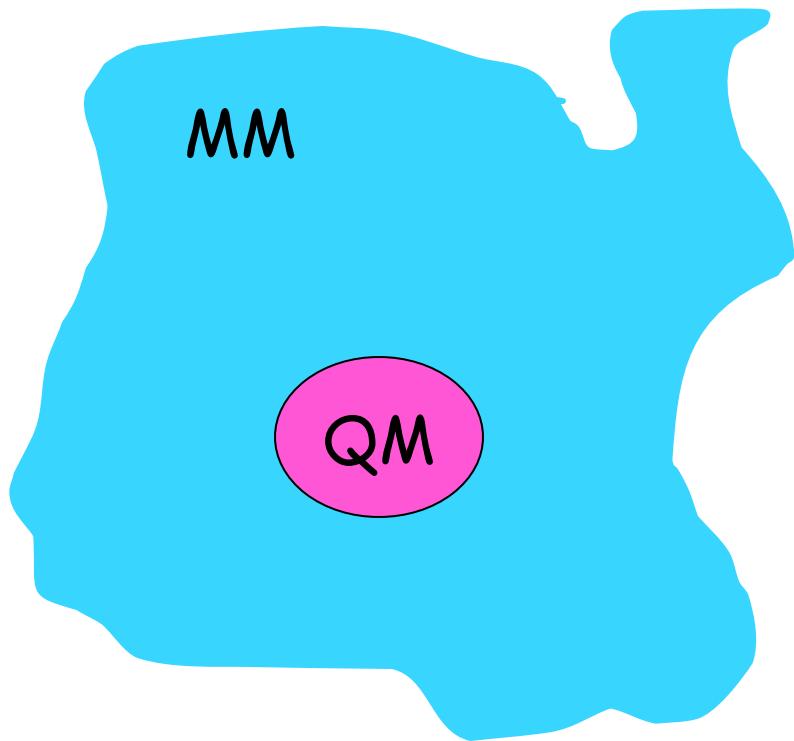


# Classical Retinal Isomerization



Twist Propagation

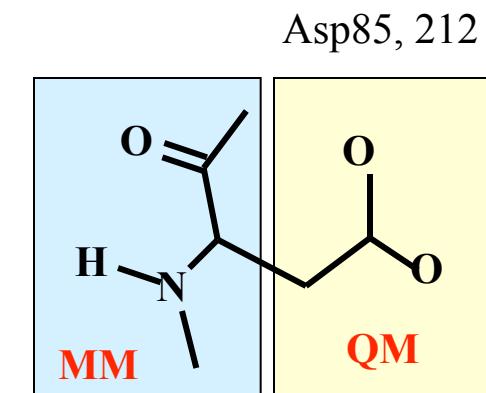
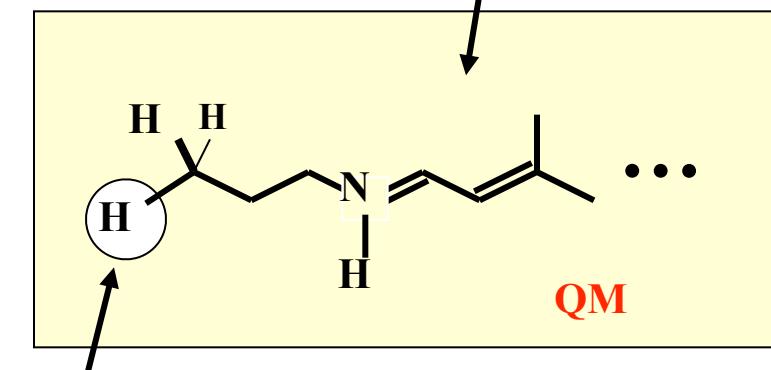
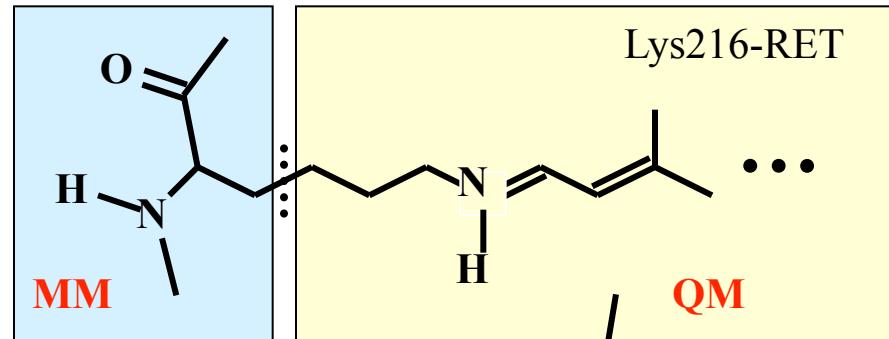
# QM/MM calculations



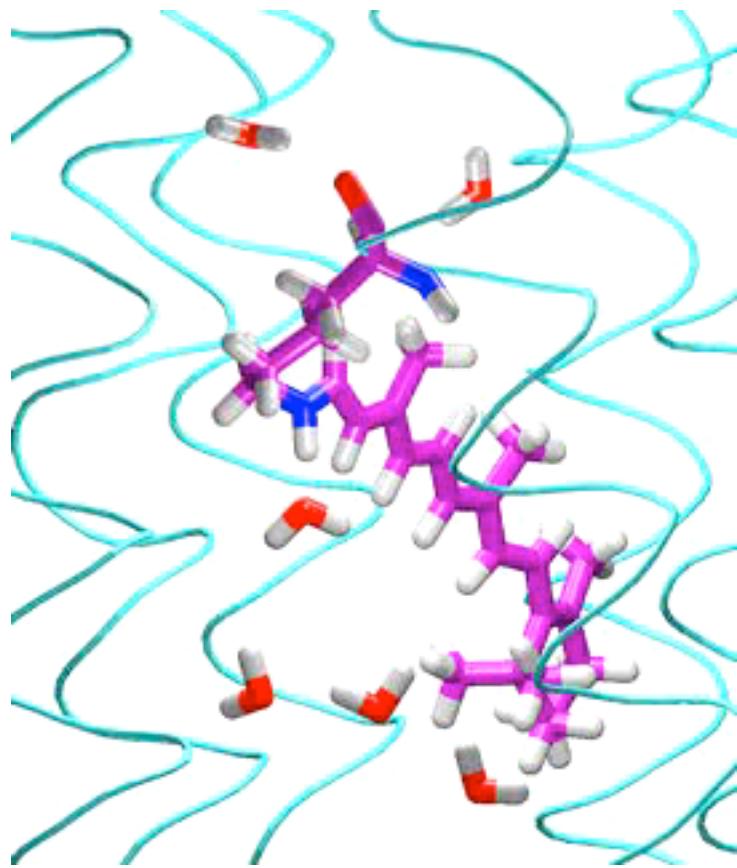
$$\hat{H} = \sum_i \frac{1}{2} p_i^2 + \sum_i \sum_A \frac{Z_A}{r_{iA}} + \sum_{i>j} \frac{1}{r_{ij}} + \sum_{A>B} \frac{Z_A Z_B}{r_{AB}}$$

$$+ \sum_i \sum_p \frac{q_p}{r_{ip}} + \sum_A \sum_p \frac{Z_A q_p}{r_{Ap}}$$

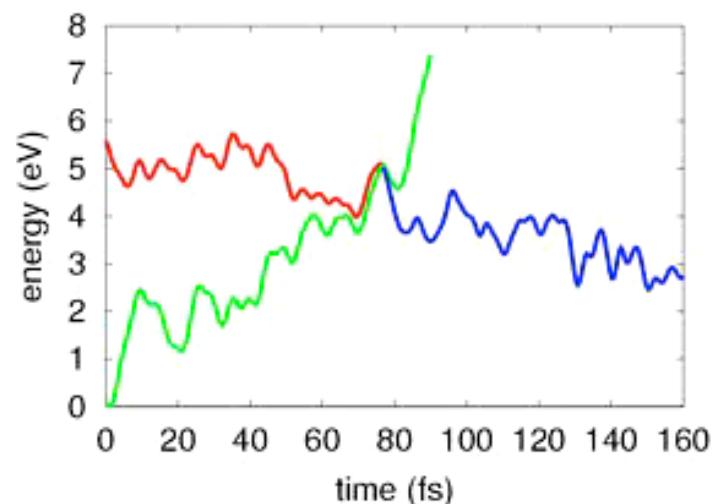
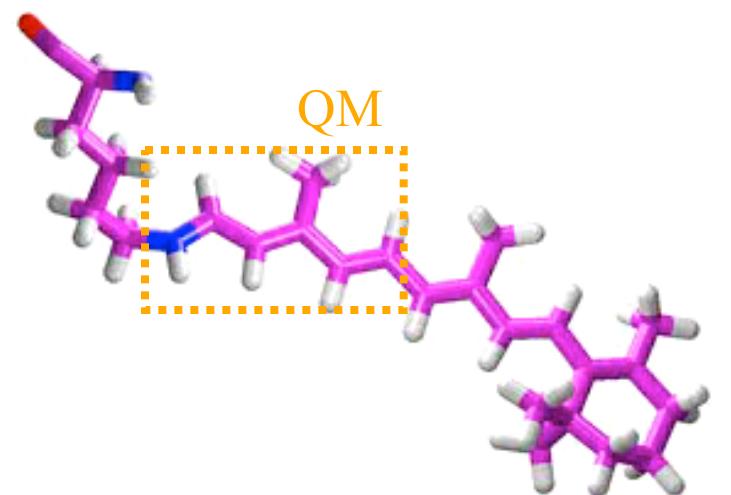
$$+ V_{QM-MM}^{MM} + V_{MM}^{MM}$$



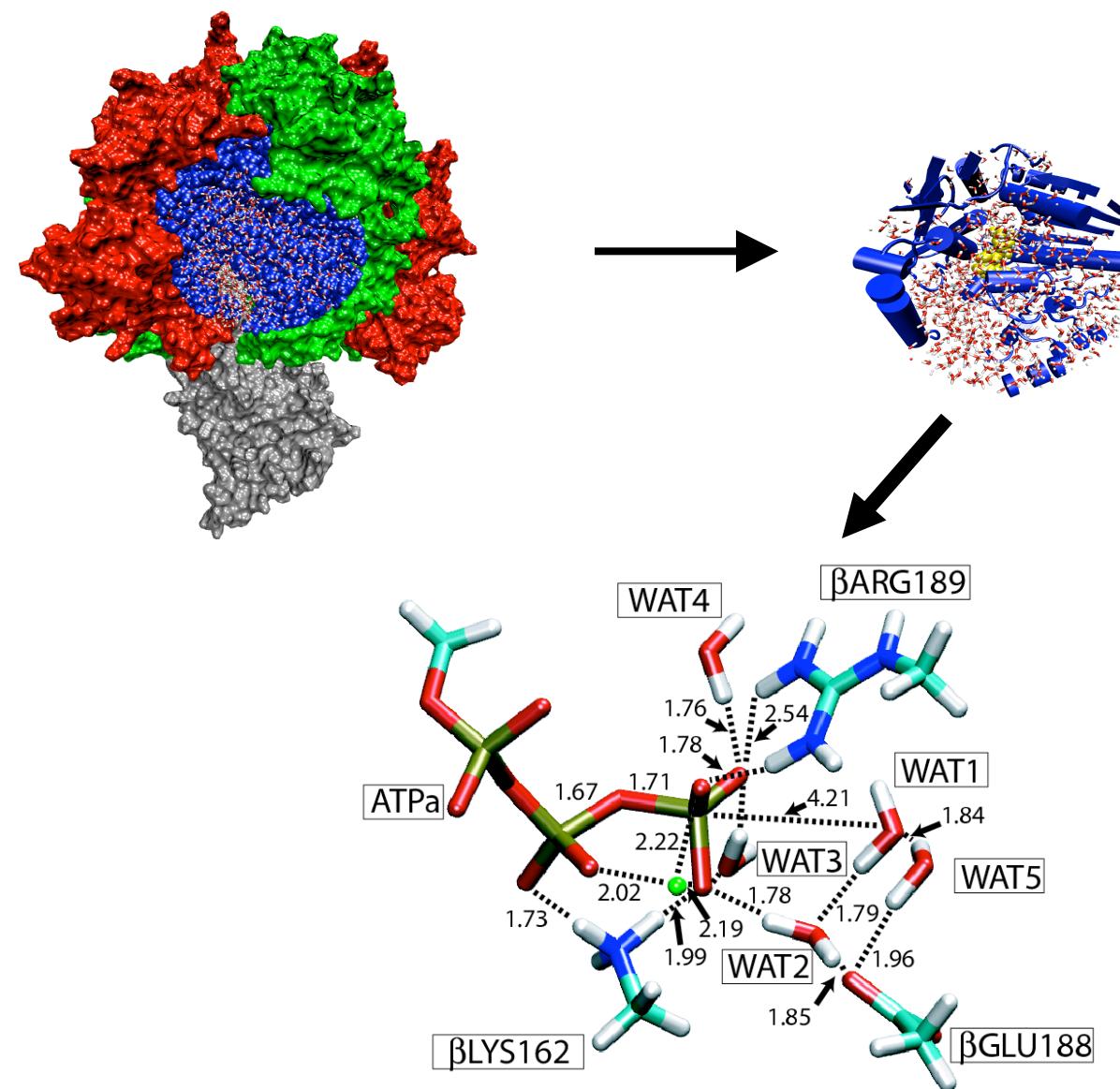
# Ab Initio QM/MM Excited State MD Simulation



Quantum mechanical (QM)  
treatment of the chromophore,  
and force field (MM) treatment of  
the embedding protein



# QM/MM calculation of ATP hydrolysis



# Coarse grain modeling of lipids

150 particles



9 particles!

(A)

