

NSF Summer School UIUC 2003Molecular Machines of theLiving Cell : PhotosyntheticUnit from Light Absorption toATP SynthesisMelihThorstenSenerIoanKosztin

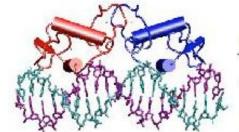


Ana Damjanovic Theoretical Biophysics Group (also Sanghyun Park, Deyu Lu, and Ulrich Kleinekathoefer)

Research Opportunities in the Teraflop Era

Towards Larger Molecules

BPTI 3K atoms



Estrogen Receptor 36K atoms (1996)

> ATP Synthase 327K atoms (2001)

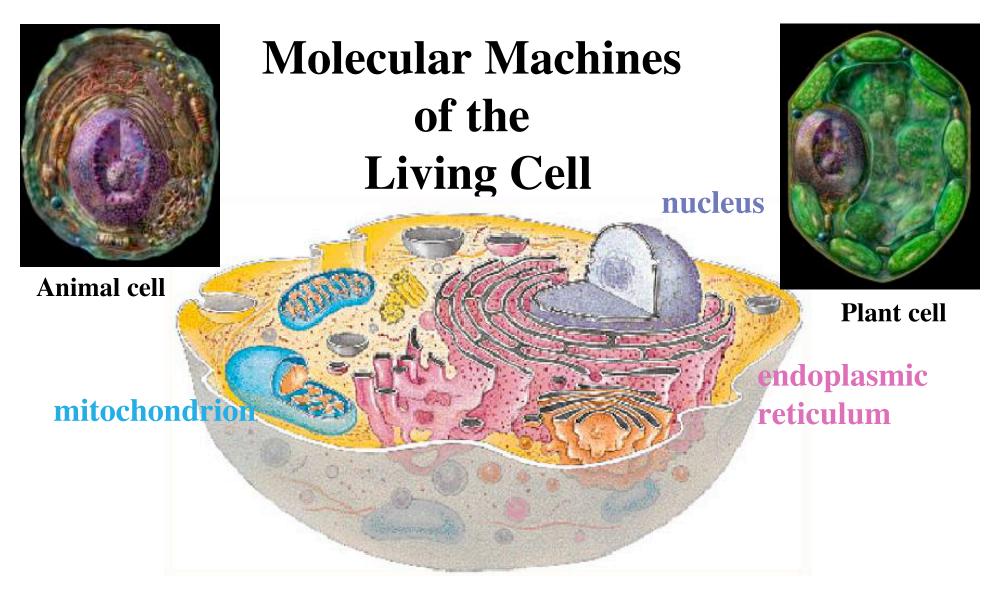
• Studying protein-protein and protein-nucleic acid recognition and assembly.

• Investigating integral functional units (membrane proteins, signal transduction, motors, bioenergetic apparatus).

• Bridging the gap between computationally feasible and functionally relevant time scales.

• Combining classical molecular dynamics simulations with quantum chemical forces.

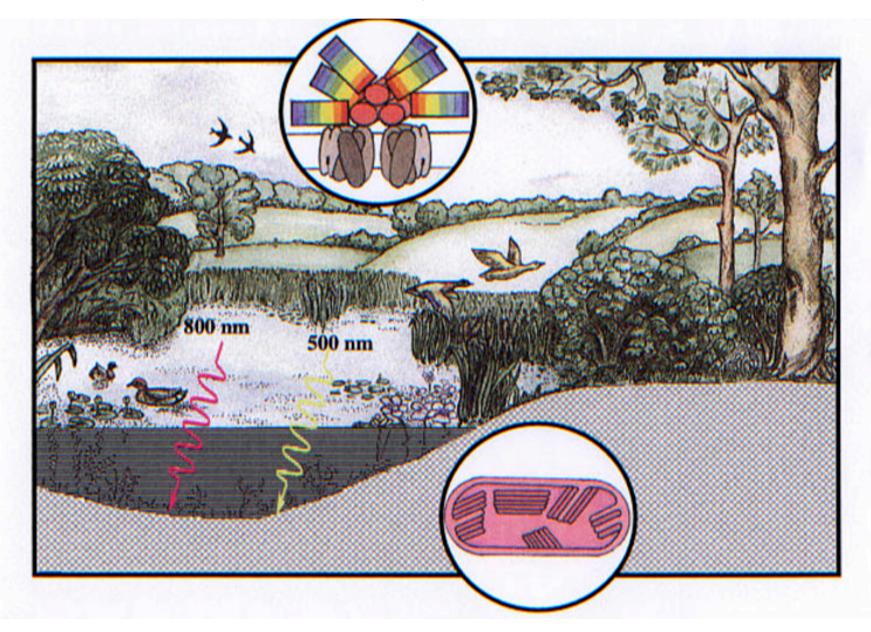
• Describing integral cell functions.



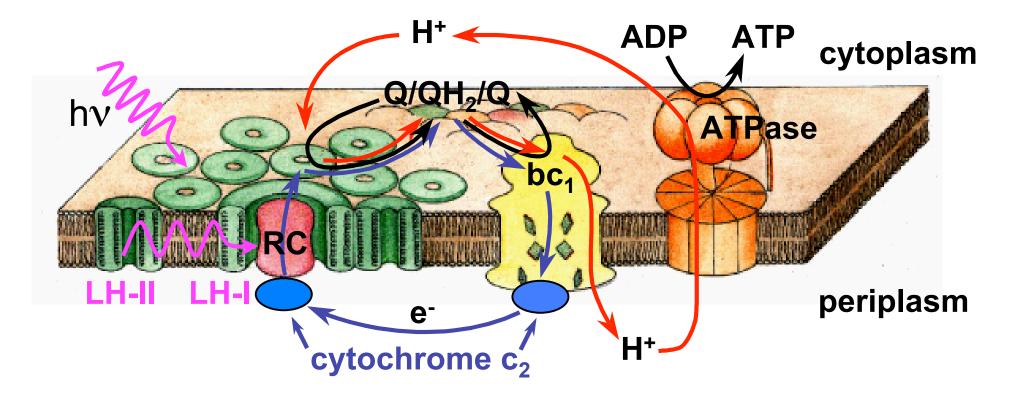
Study of integral cell functions:

gene storage, regulation, and expression; protein synthesis and degradation; energy conversion and storage; cell motion; cell signaling; metabolic pathways; ...

Habitats of Photosynthetic Life Forms



Photosynthetic Apparatus of Purple Bacteria



RC - Photosynthetic Reaction Center

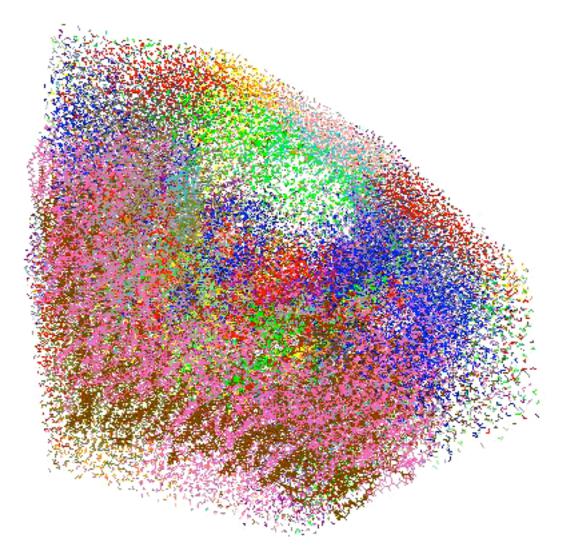
LH – Light Harvesting Complex

Structure of RC+LH-I+Cyt System

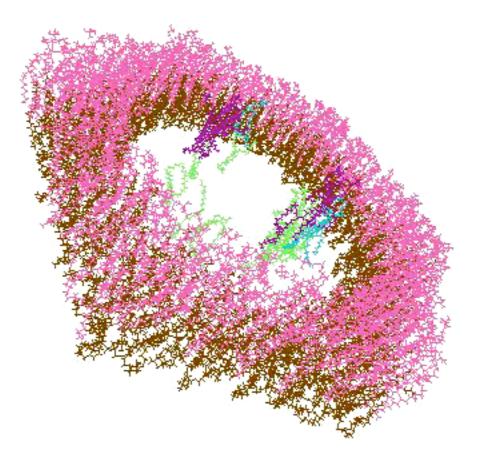
bc

cytochrome c₂

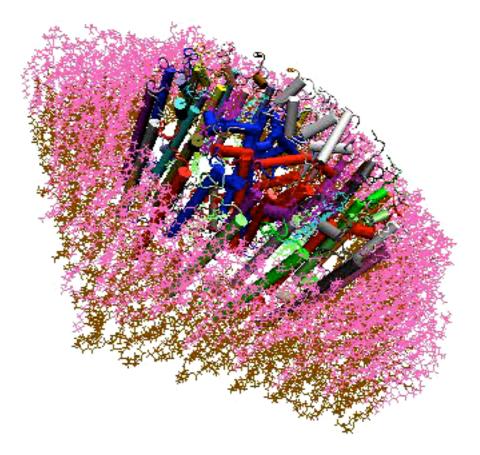
Focussing on the Structure of RC + LH-I



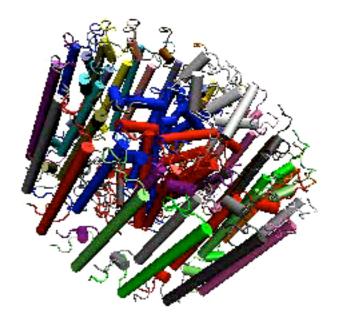
System of Water - Lipids - Protein



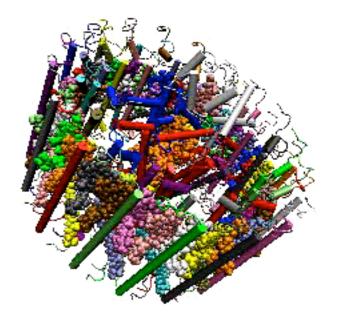
Lipids only



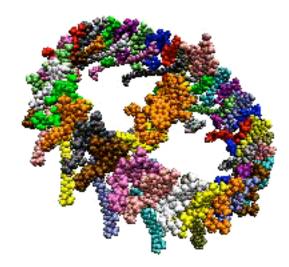
Lipids and Proteins



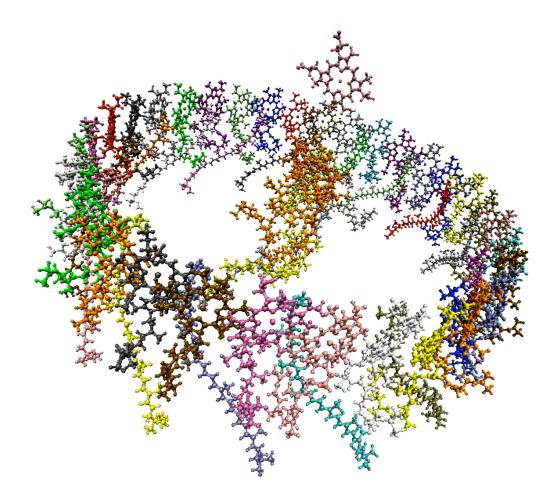
Proteins only



Proteins and Chromophores

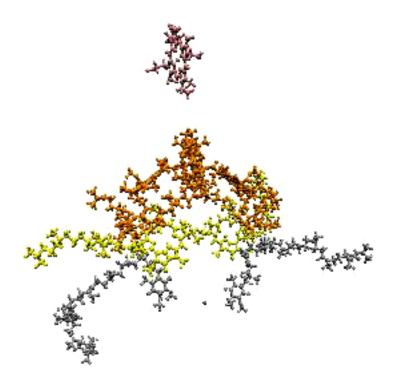


Chromophores only

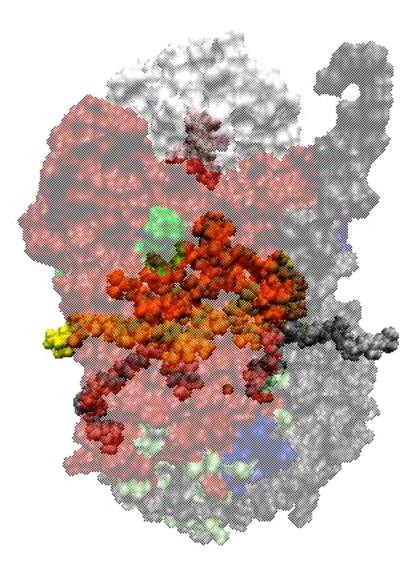


Chromophores only

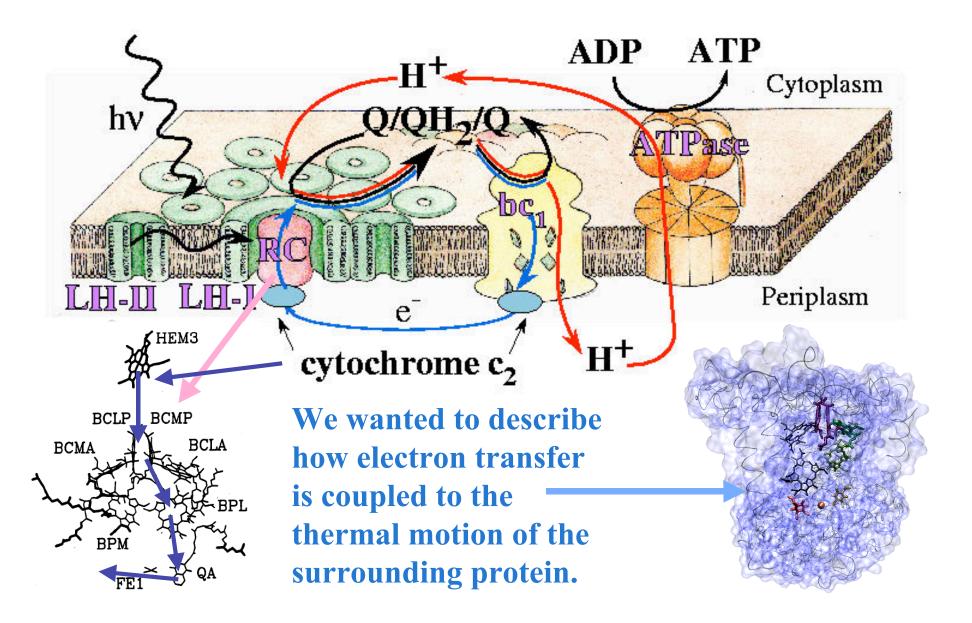
Electron Transfer Chain in RC + Cyt c Complex



Role of the Protein Matrix on Electron Transfer



Role of Thermal Disorder on Electron Transfer in the Photosynthetic Reaction Center



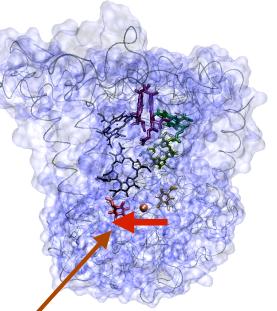
Electron Transfer Process Coupled to the Protein Matrix

We assumed that the electron transfer $Q_A^- Q_B^- > Q_A Q_B^-$ is coupled to an ensemble of oscillators representing the protein matrix

Hamiltonian
$$\hat{H}_{qo}^{(s)} = \begin{pmatrix} \hat{H}_r^{(s)} & v \\ v & \hat{H}_p^{(s)} + E \end{pmatrix}$$

Protein matrix is a bath of oscillators linearly coupled to the electron transfer according to

$$\hat{H}_r = \sum_j \left(\frac{\hat{p}_j^2}{2M_j} + \frac{1}{2} M_j \omega_j^2 q_j^2 \right)$$
$$\hat{H}_p = \sum_j \left(\frac{\hat{p}_j^2}{2M_j} + \frac{1}{2} M_j \omega_j^2 \left(q_j - \frac{c_j}{M_j \omega_j^2} \right)^2 \right)$$



Dong Xu and Klaus Schulten. Chemical Physics, 182: 91--117, 1994.

Klaus Schulten. In D. Bicout and M. J. Field, editors, Proc. Ecole de Physique des Les Houches, pp 85--118, Les Editions de Physique, Springer, Paris, 1995.

Klaus Schulten. Science, 290:61--62, 2000.

Electron Transfer Process Coupled to the Protein Matrix

Rate for an ensemble of oscillators (spin boson model, Legett et al)

$$k_{qb}(R \to P) = \frac{v^2}{\hbar^2} \int_{-\infty}^{+\infty} dt \; e^{itE/\hbar} \; e^{iQ_1(t)/\pi\hbar} \; e^{-Q_2(t)/\pi\hbar}$$

Relaxation rate

$$k_{\rm rel} = \frac{2v^2}{\hbar^2} \int_0^{+\infty} dt \cos(-tE/\hbar) \cos(Q_1(t)/\pi\hbar) e^{-Q_2(t)/\hbar}$$
$$Q_1(t) = \frac{\pi}{2} \sum_j \frac{c_j^2}{\hbar\omega_j^3} \sin\omega_j t$$
$$Q_2(t) = \frac{\pi}{2} \sum_j \frac{c_j^2}{\hbar\omega_j^3} \coth\frac{\hbar\omega_j}{2kT} [1 - \cos(\omega_j t)]$$

 $(t)/\pi\hbar$

But we didn't know all the coupling constants c_i? All we needed to know was J

$$J(\omega) = \frac{\pi}{2} \sum_{j} \frac{c_j^2}{\omega_j} \,\delta(\omega - \omega_j) \frac{Q_1(t)}{Q_2(t)} = \frac{\int_0^\infty d\omega \,\omega^{-2} J(\omega) \sin\omega t}{Q_2(t)} = \frac{\pi}{2} \int_0^\infty d\omega \,\omega^{-2} J(\omega) \coth\frac{\hbar\omega}{2kT} (1 - \cos\omega t)$$

Electron Transfer Process Coupled to the Protein Matrix

Relaxation rate

$$k_{rel} = \frac{2v^2}{\hbar^2} \int_0^{+\infty} dt \cos(tE/\hbar) \cos(Q_1(t)/\pi\hbar) e^{-Q_2(t)/\pi\hbar}$$

$$Q_1(t) = \int_0^{\infty} d\omega \, \omega^{-2} J(\omega) \sin\omega t$$

$$Q_2(t) = \frac{\pi}{2} \int_0^{\infty} d\omega \, \omega^{-2} J(\omega) \cosh\frac{\hbar\omega}{2kT} (1 \cos\omega t)$$

$$\frac{J(\omega)}{\omega} = \frac{\sigma^2}{k_B T} \int_0^{\infty} dt \, C(t) \cos\omega t$$

$$C_{\epsilon\epsilon}(t) = \frac{\langle (\epsilon(t) - \langle \epsilon \rangle) (\langle \epsilon(0) - \langle \epsilon \rangle) \rangle}{\langle \epsilon(0) - \langle \epsilon \rangle \rangle^2}$$
energy gap correlation function

$$\sigma \text{ rms deviation of energy gap}$$

$$k_{rel} = \frac{\sqrt{(\omega)}{4\pi} \int_0^{\infty} dt \, C(t) \cos\omega t$$

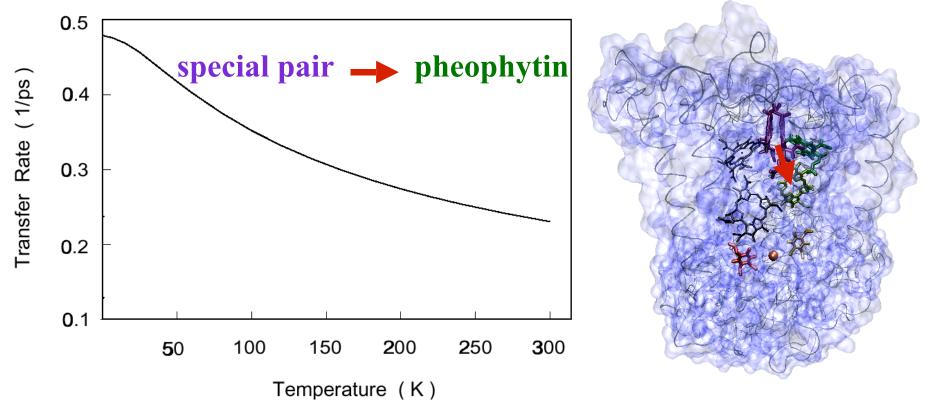
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Temperature Dependence of Electron Transfer Rate

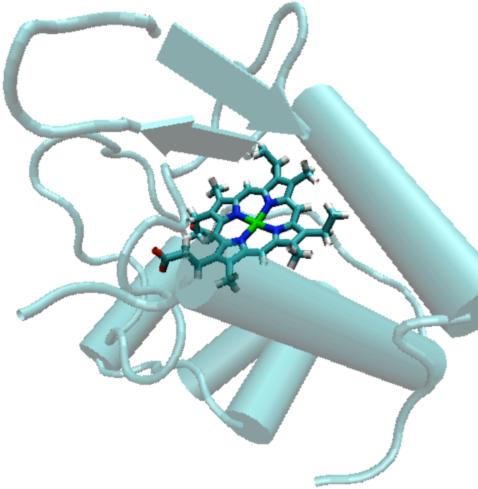


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Coupling protein motion to electron transfer via MD

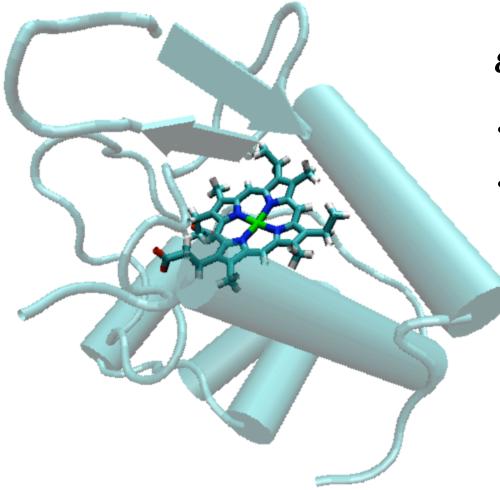


• Cytochrome c_2 from purple bacterium *Rhodobacter sphaeroides*.

• Serves as electron carrier between bc1-complex and reaction center

When the gene encoding cytochrome c_2 is deleted from *Rb. sphaeroides*, the bacterium is unable to grow photosynthetically.

The energy gap function



$$\varepsilon(t) = E_P(t) - E_R(t)$$

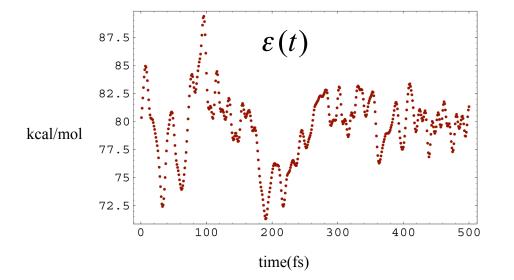
- *R*: reactant state (reduced)
- *P*: product state (oxidized)

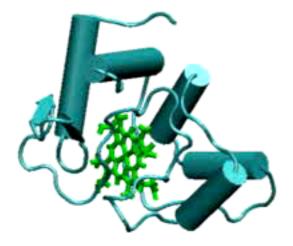
Tutorial:

You will do two consecutive NAMD runs.

- obtain an MD trajectory
- evaluate $\mathcal{E}(t)$ at each frame of the first trajectory through a second NAMD run

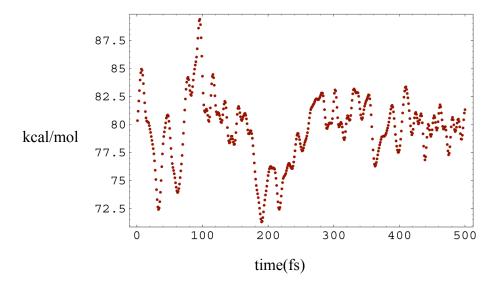
MD simulation of the electron transfer process





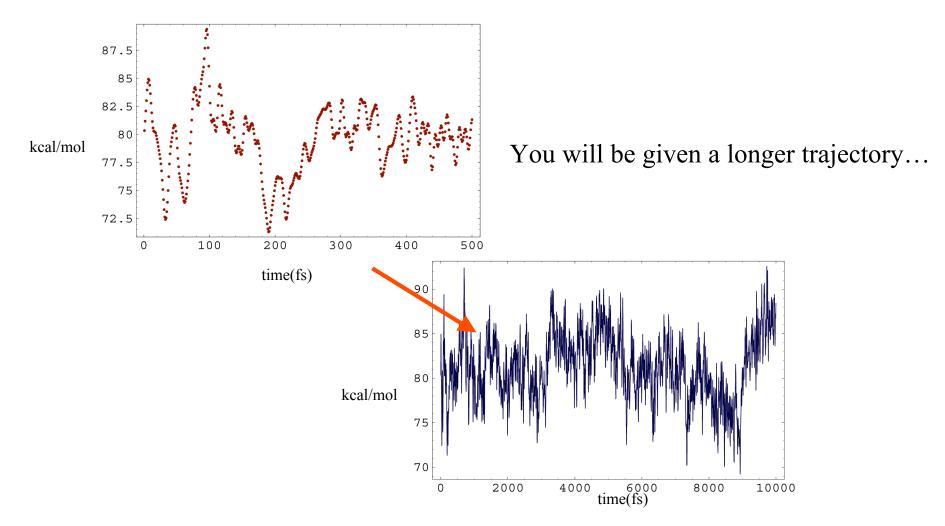
- ~12000 atoms solvated system
- Already minimized and equilibrated
- You will continue from a restart file (so, you do not need to worry about velocity relaxation)

The energy gap function

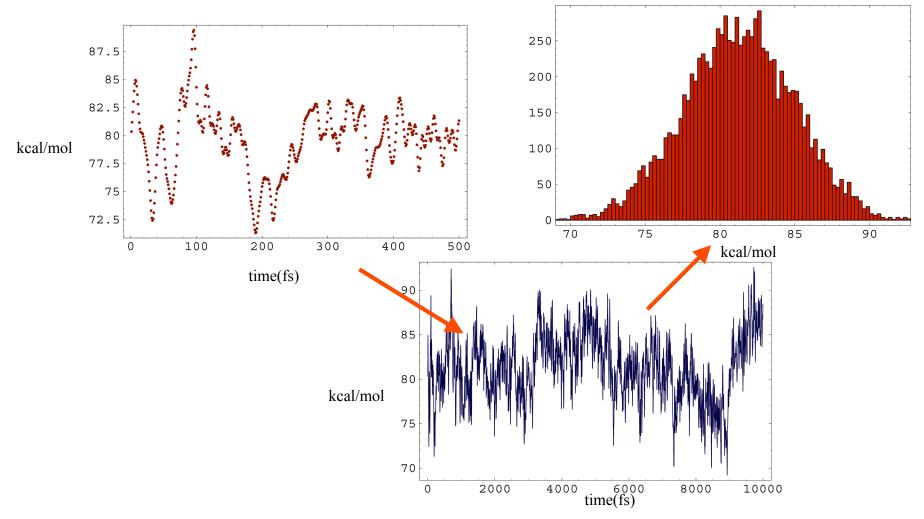


Result from the first 500fs

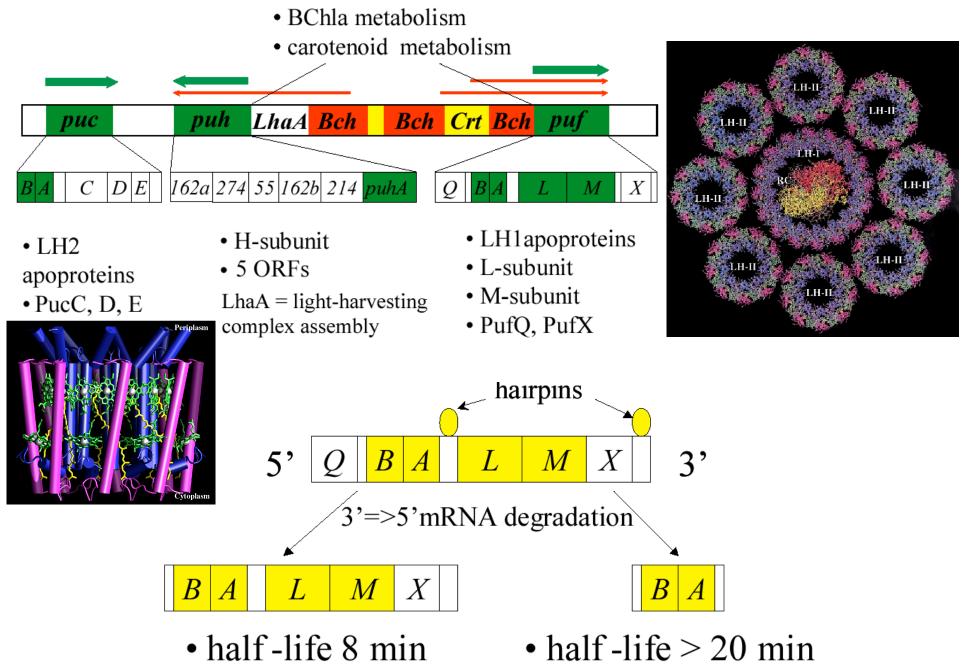
The energy gap function



The energy gap function



Genomic Organization of the Light Harvesting Complexes



Photosynthetic apparatus of purple bacteria

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