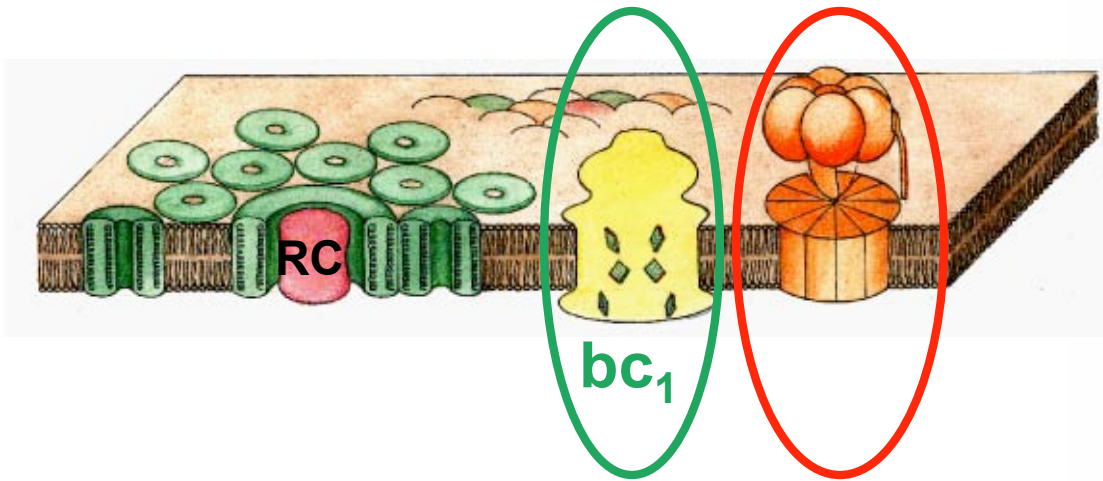
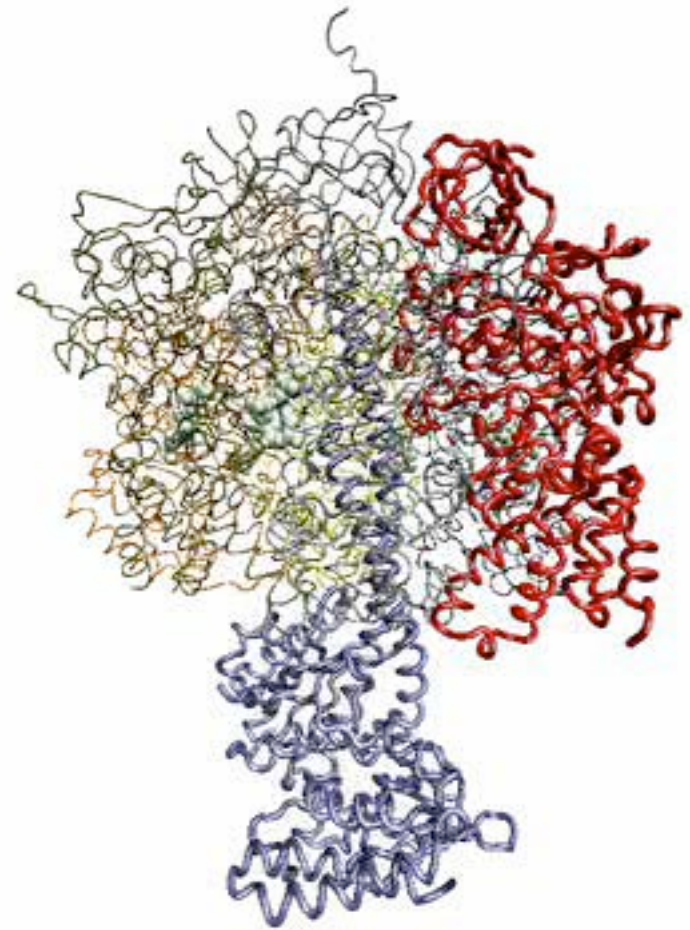


ATPase Synthase - A Molecular Double Motor



bc_1

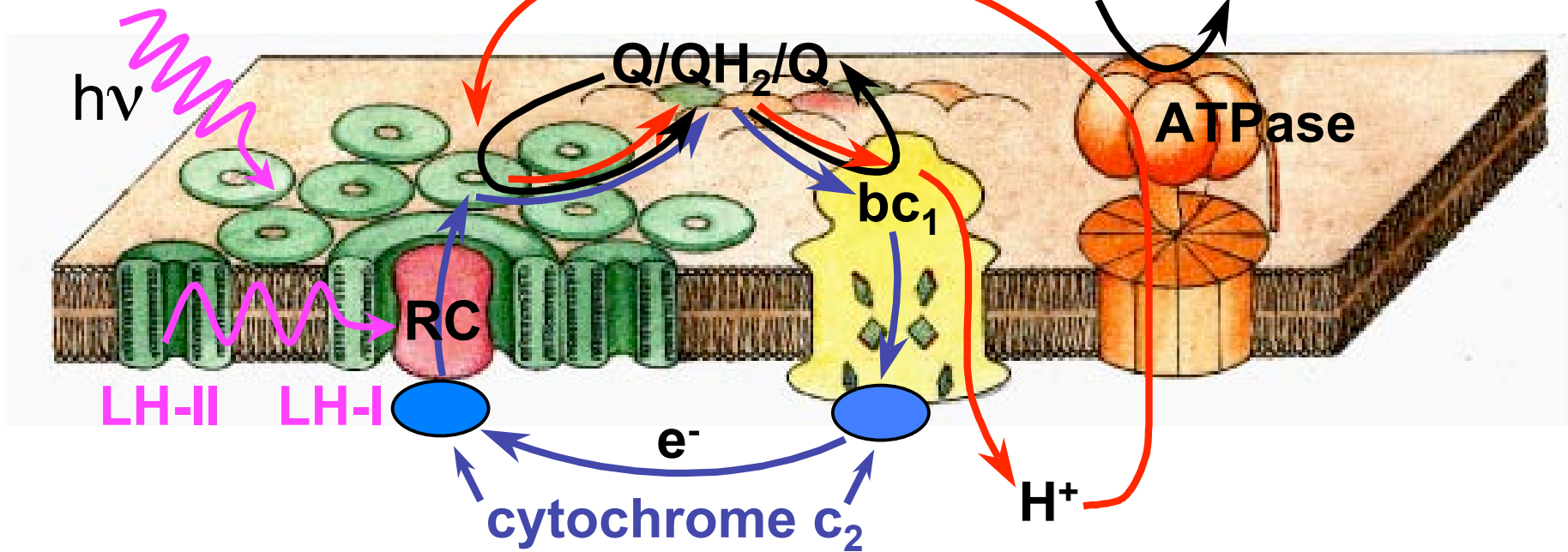


ATPase

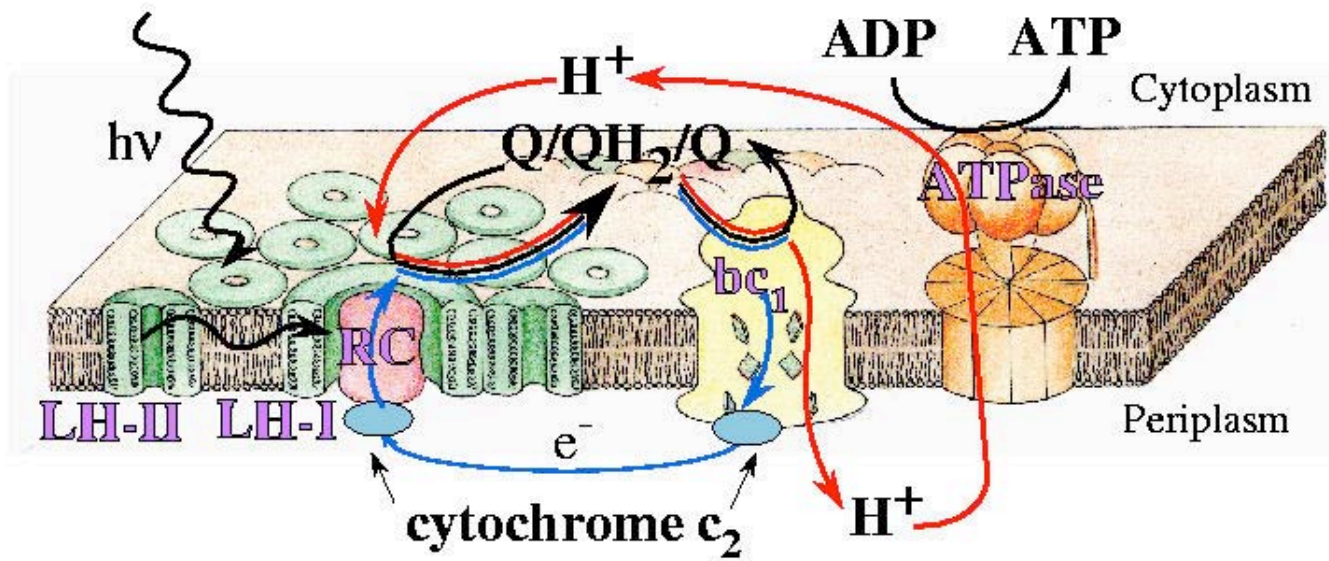
Photosynthetic Unit of Purple Bacteria

Module that converts sun light into chemical energy (ATP)

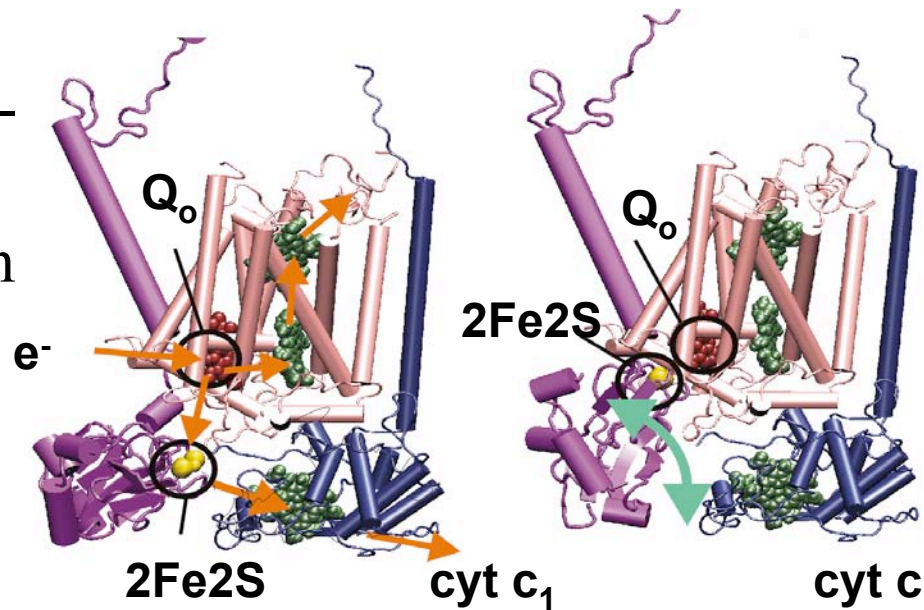
Light in



Mechanism of the bc1 Complex in the Photosynthetic Unit



two pathways for oxidation of Q_o site

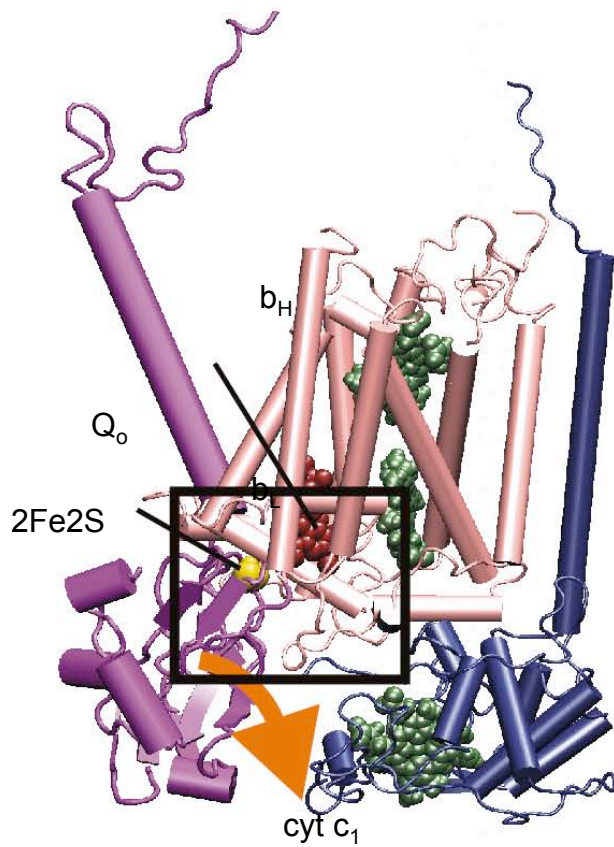


Iron Sulfur Protein (ISP) head rotation can redirect 2nd electron

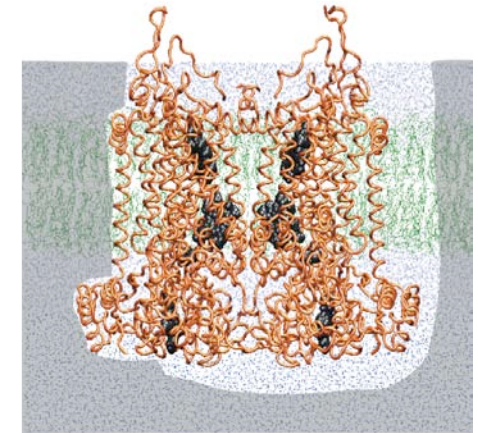
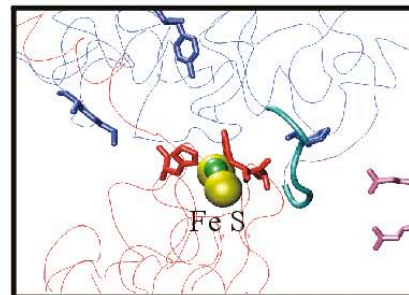
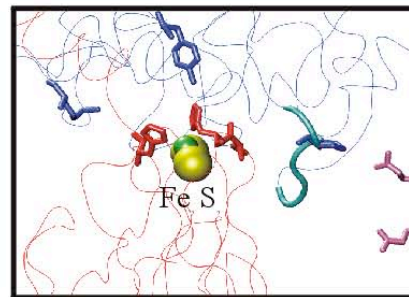
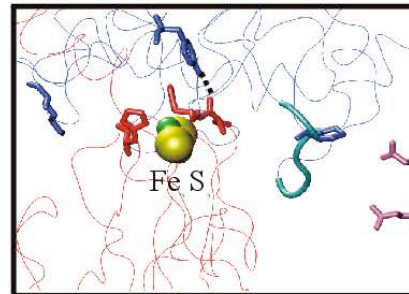
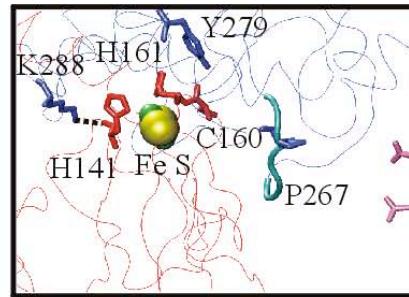
Enforcing domain rotation in the bc₁ complex

Events during torque application to ISP head

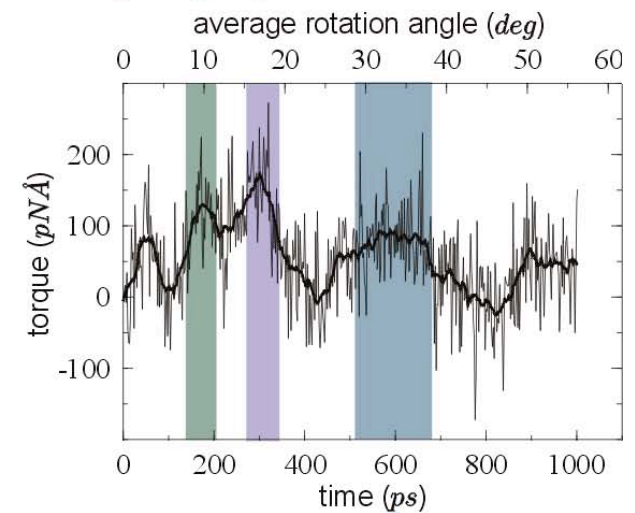
Izrailev et al., Biophys J.,
77:1753-1768 (1999)



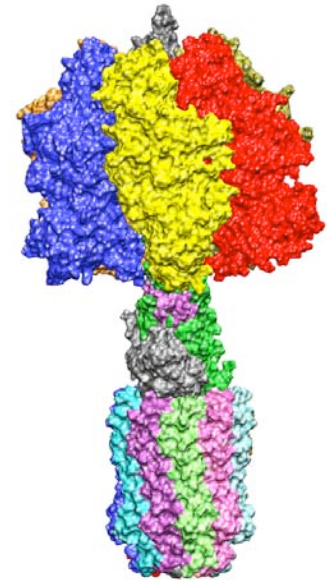
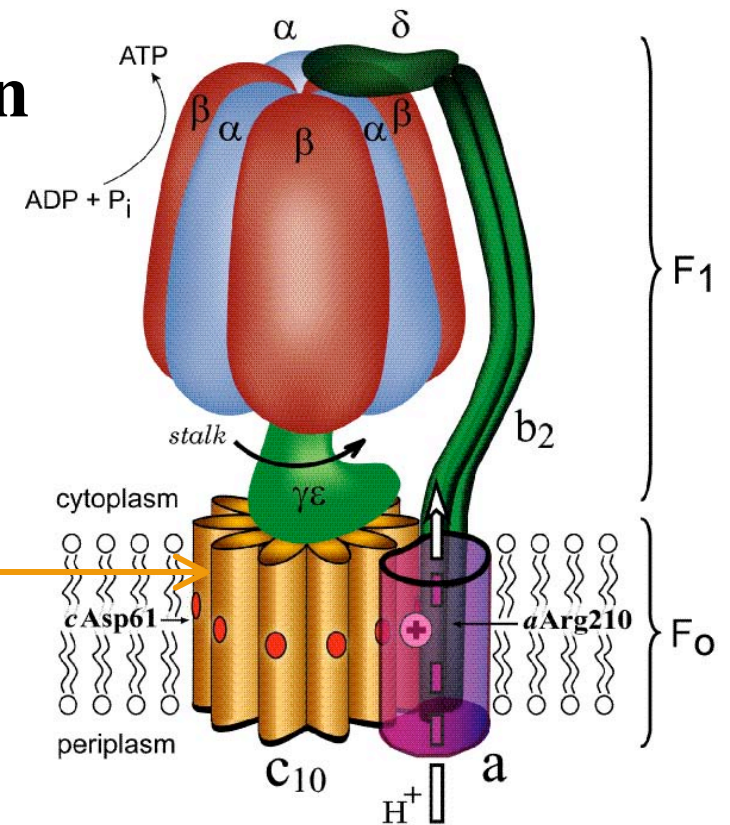
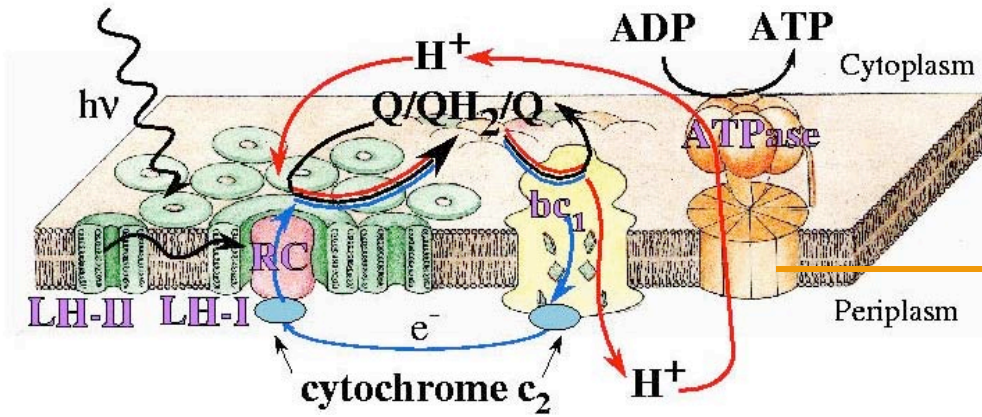
Torque applied
to 126 C_α atoms
K = 70 pN/Å
ω = 0.0561 rad/s



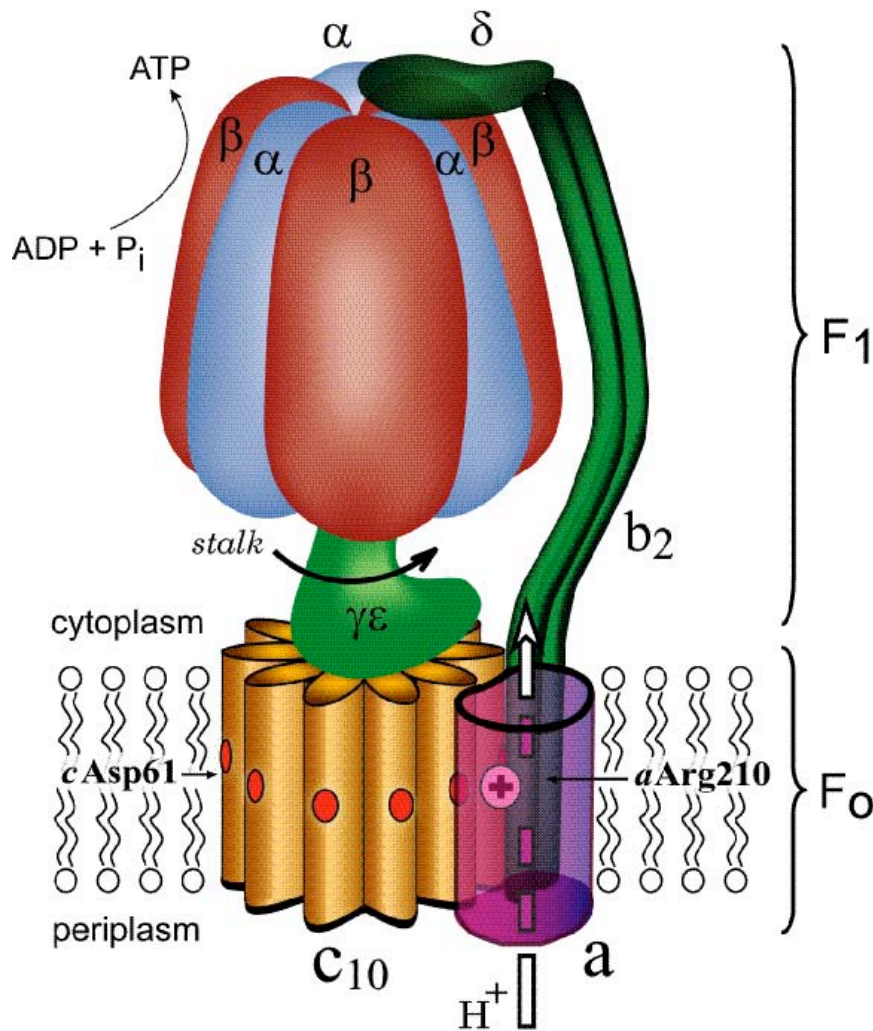
206,720 atoms



Mechanisms of Rotatory Molecular Motor that Converts Voltage (proton gradient) into ATP Synthesis



Adenosine Triphosphate (ATP) Synthase

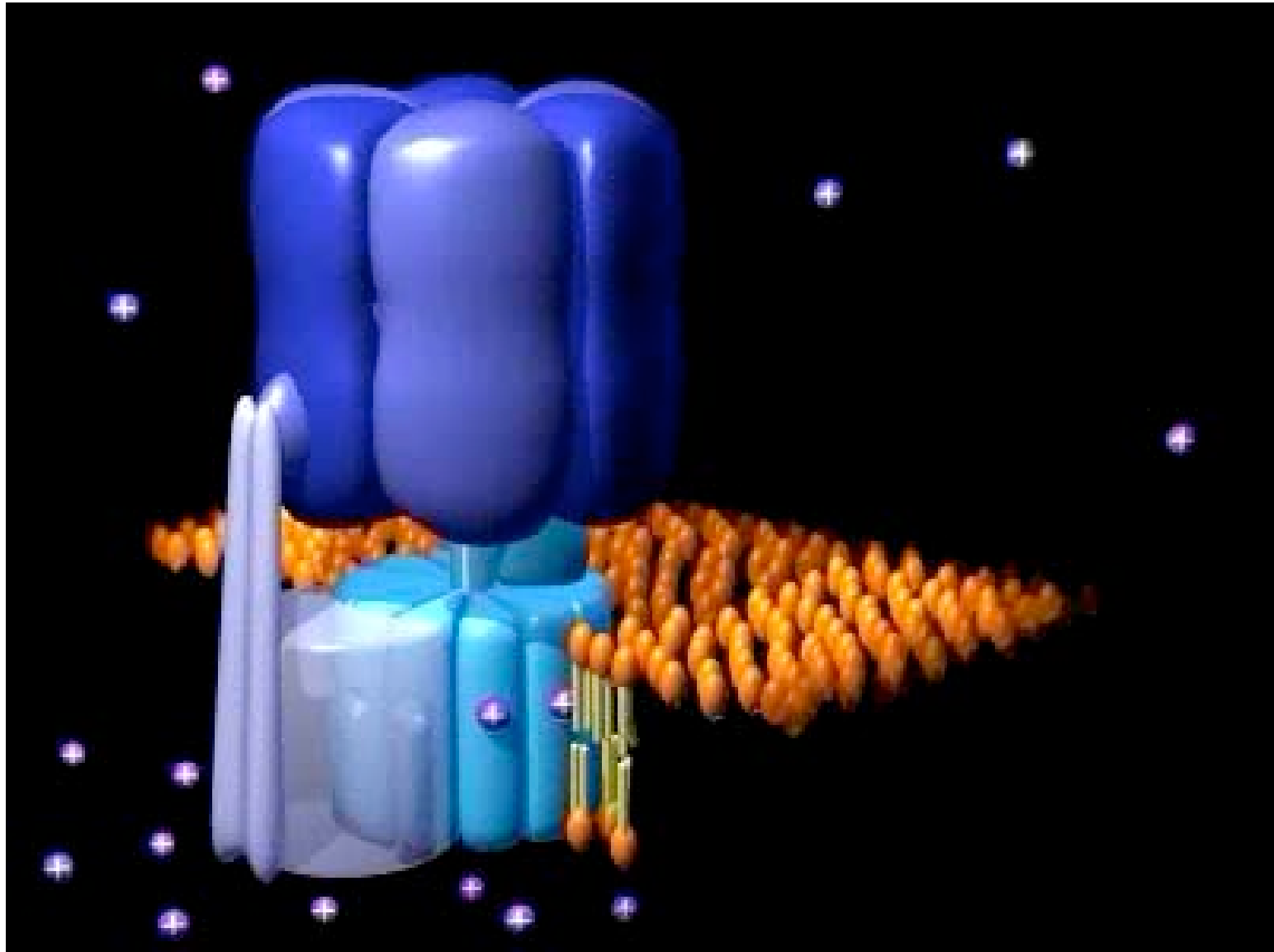


Rotary catalysis: Two protein motors coupled via common central stalk $\gamma\delta$

Solvent exposed F_1 unit ($\alpha_3\beta_3\gamma\delta\epsilon$):
central stalk rotation causes
conformational changes in catalytic
sites, driving ATP synthesis

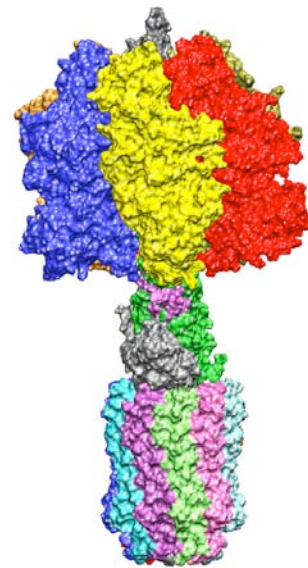
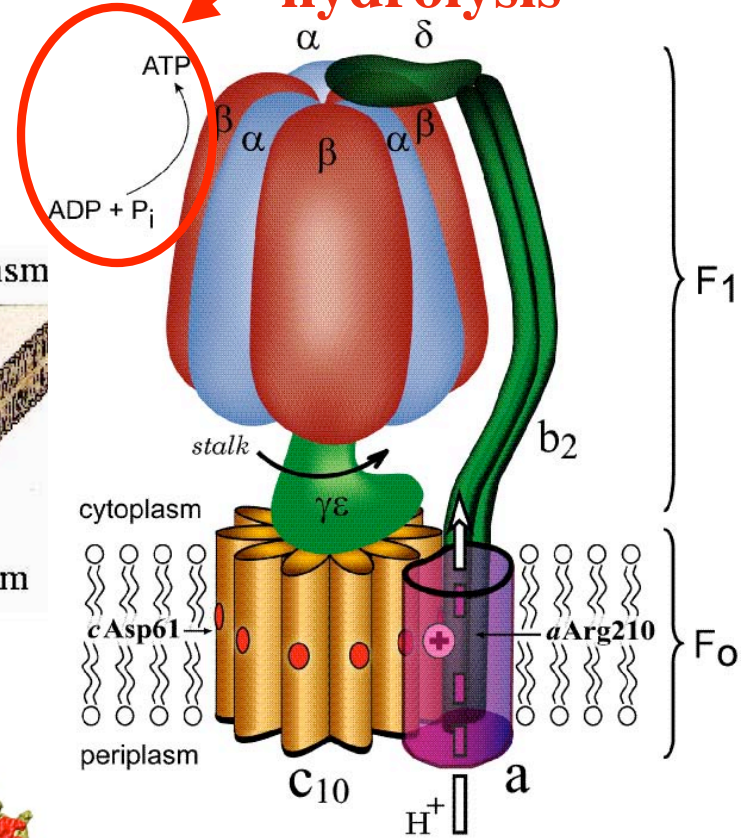
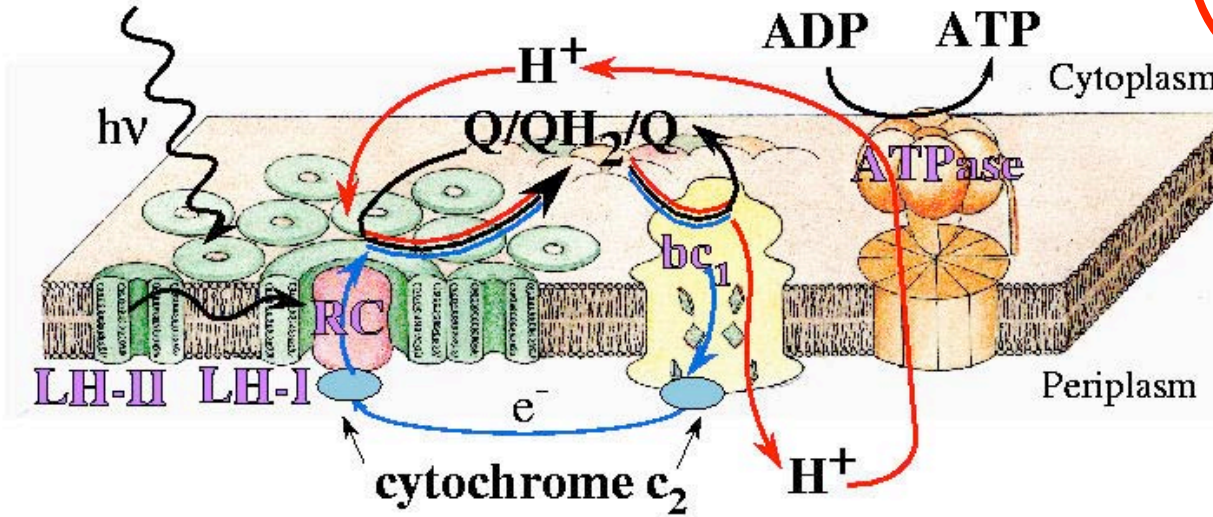
Transmembrane F_0 unit (ab_2c_{10}):
converts proton motive force into
mechanical rotation of central stalk

Animation of the ATP Synthase

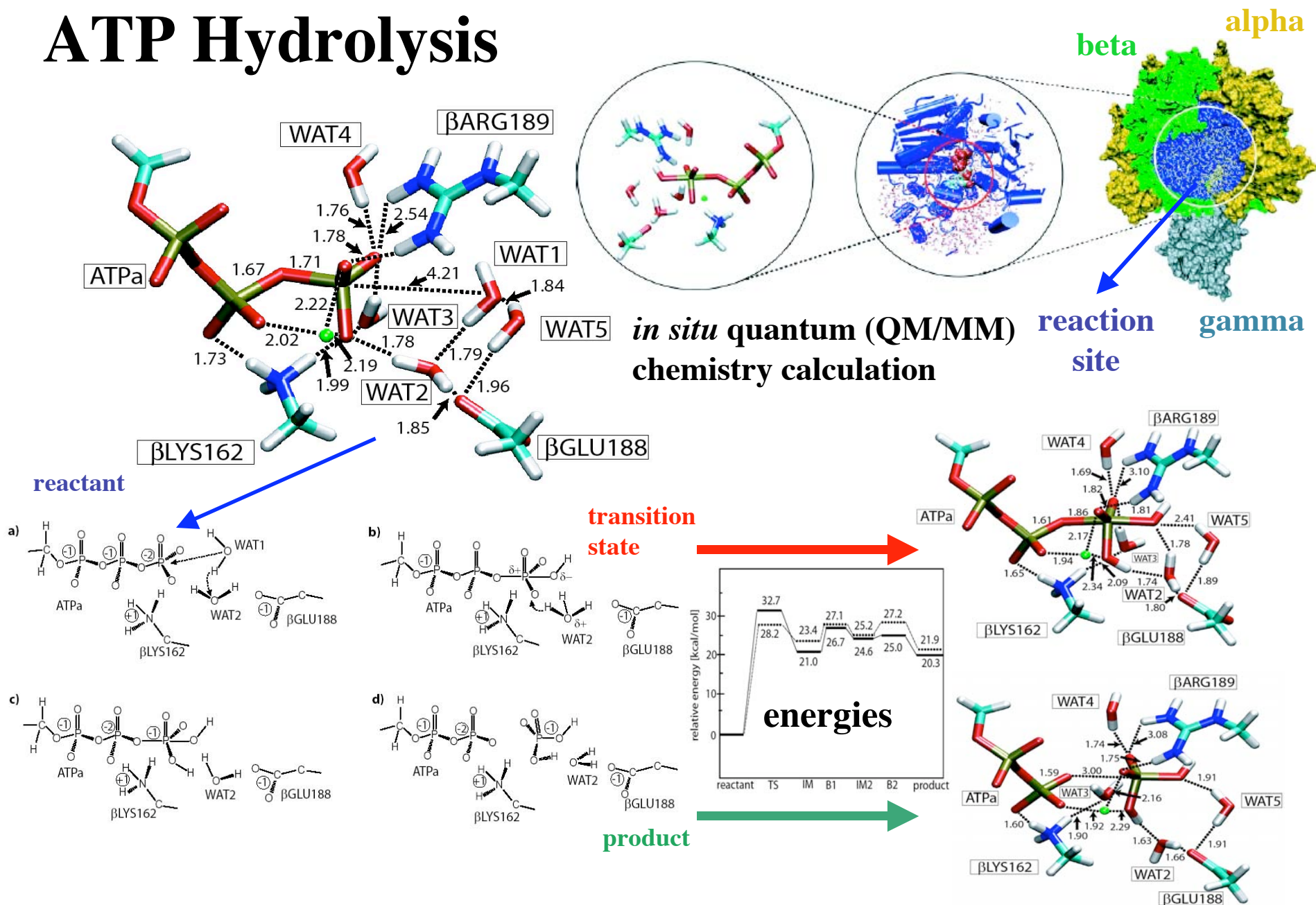


Mechanism of ATP synthase

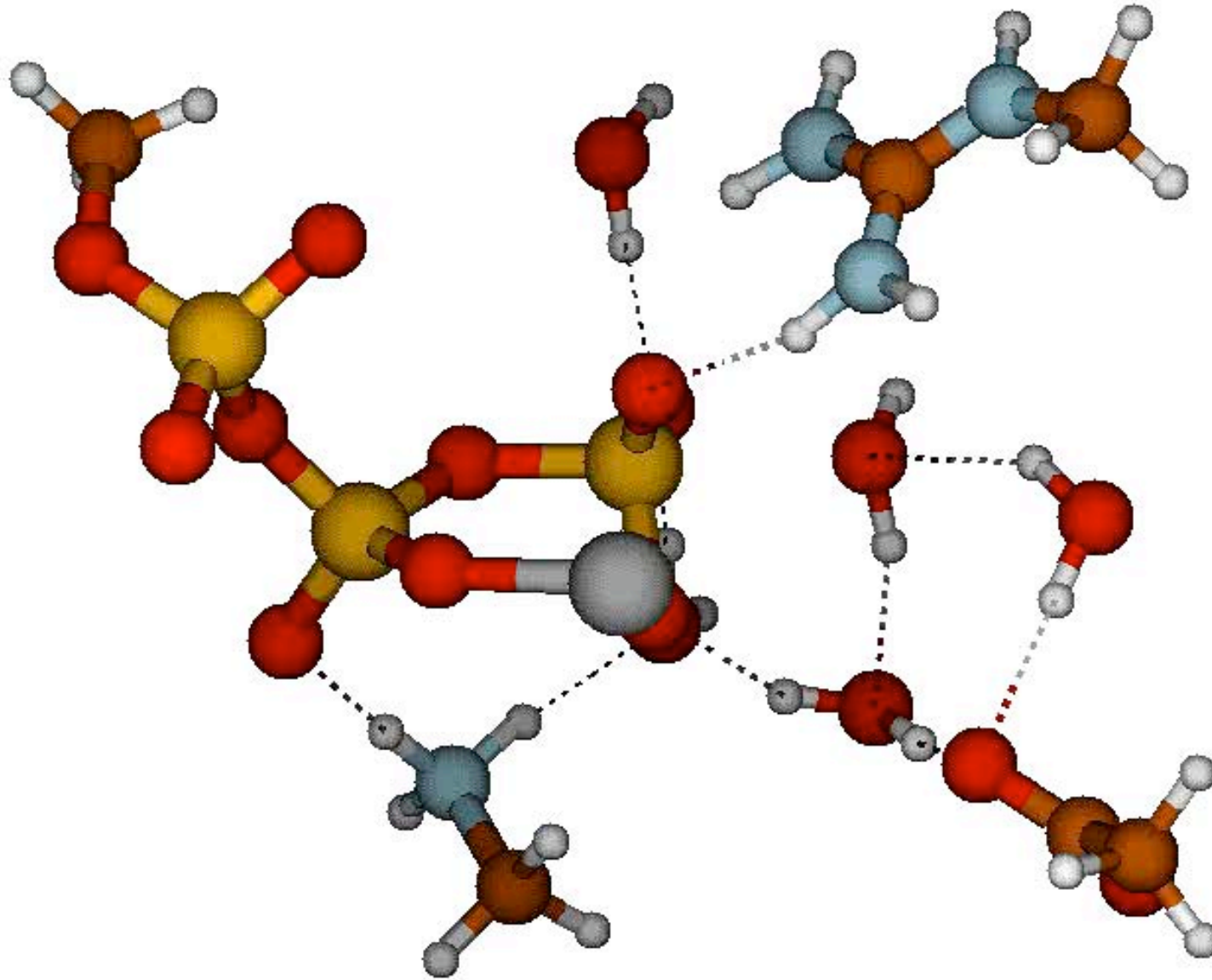
ATP synthesis / hydrolysis



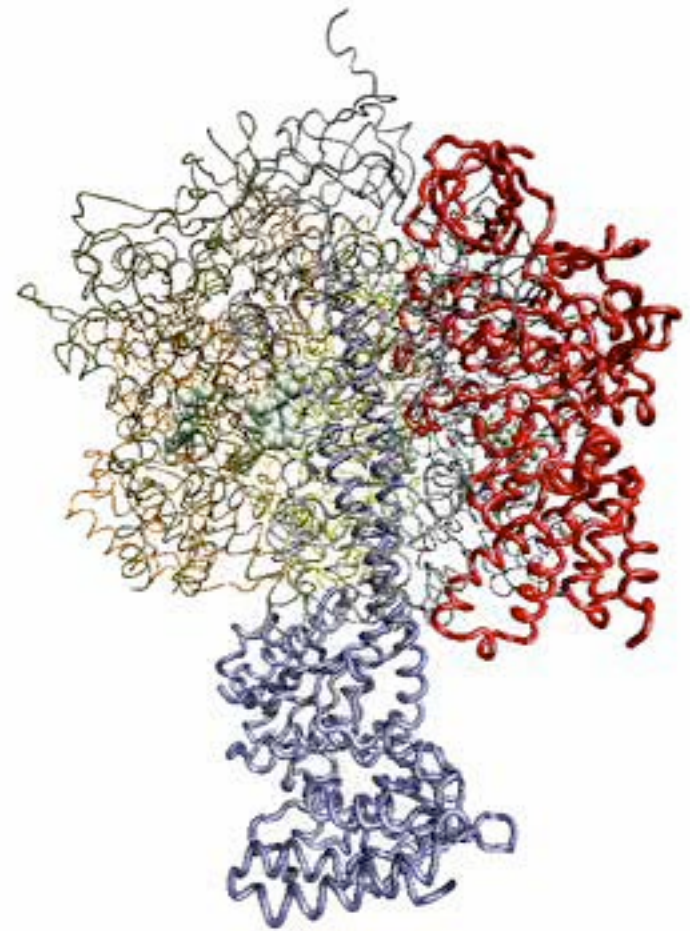
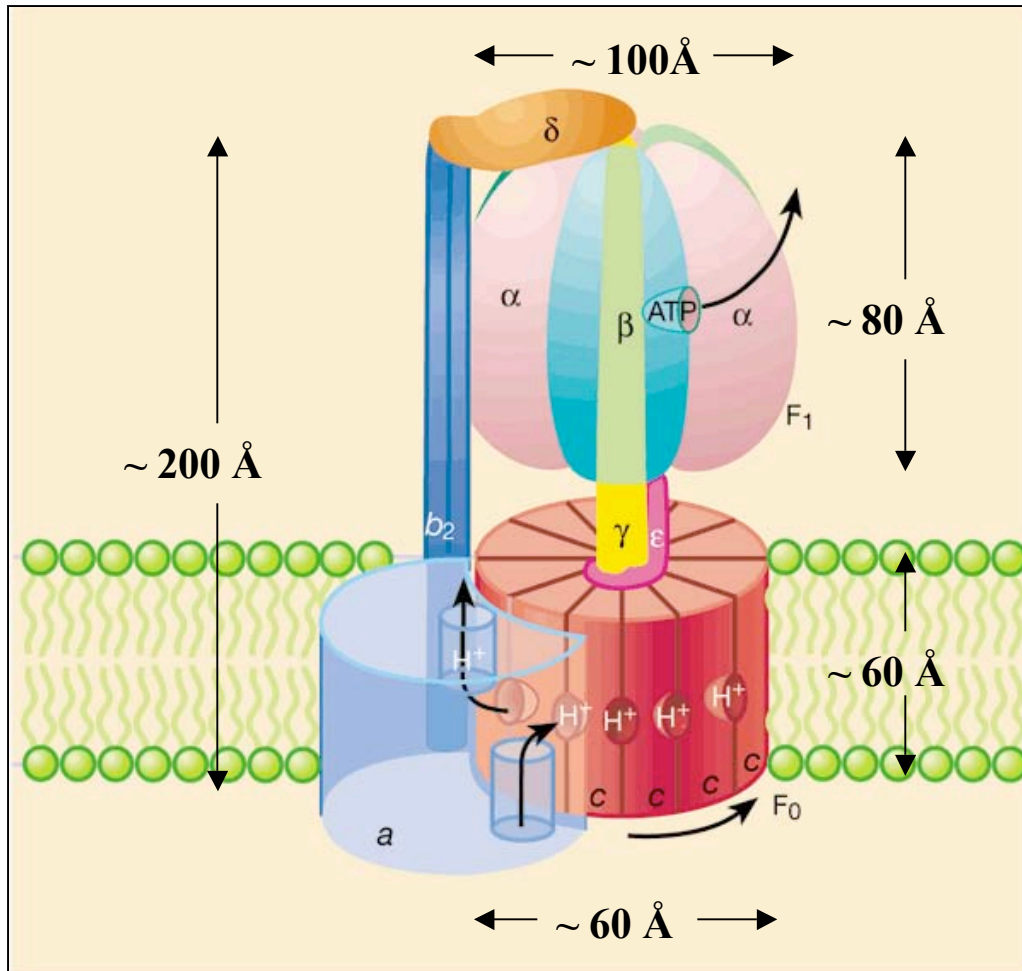
Reaction Mechanism of ATP Hydrolysis



Mechanism of ATP Hydrolysis in F1 ATPase

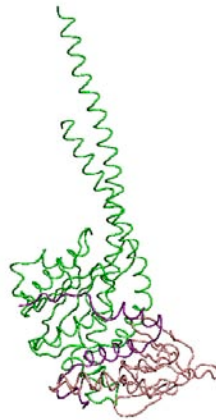


Let's look at F1



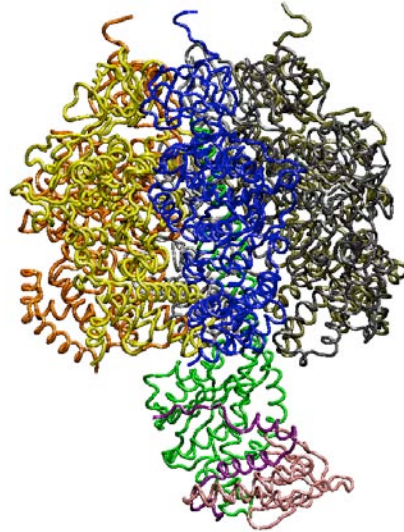
Torque is transmitted between the motors via the central stalk.

Assembling ATP Synthase F₁



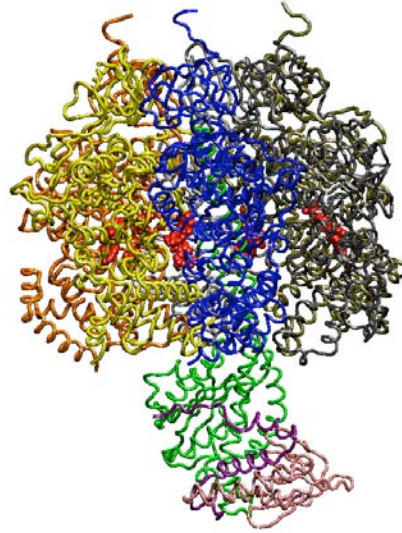
- Start with DCCD-inhibited structure, has near-complete stalk. (Gibbons 2000, PDB code 1E79)
- Total **327,000 atoms** (3325 residues, 92,000 water molecules, nucleotides, and ions).
- The 1.2 ns equilibration + 10.5 ns torque application were performed on NCSA Platinum and PSC Lemieux as parallel NAMD jobs using up to 512 processors.

Assembling ATP Synthase F₁



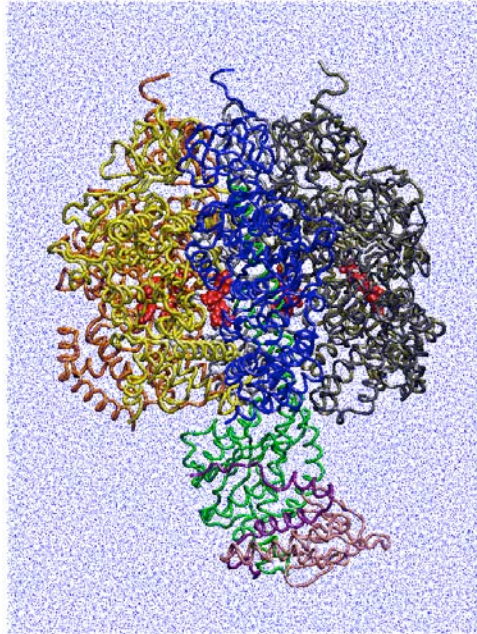
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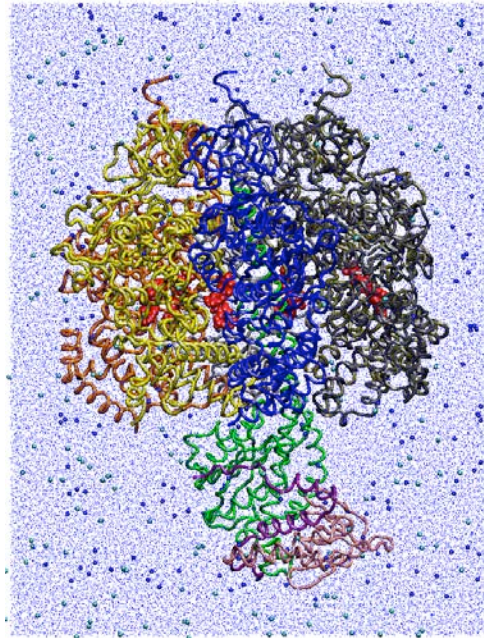
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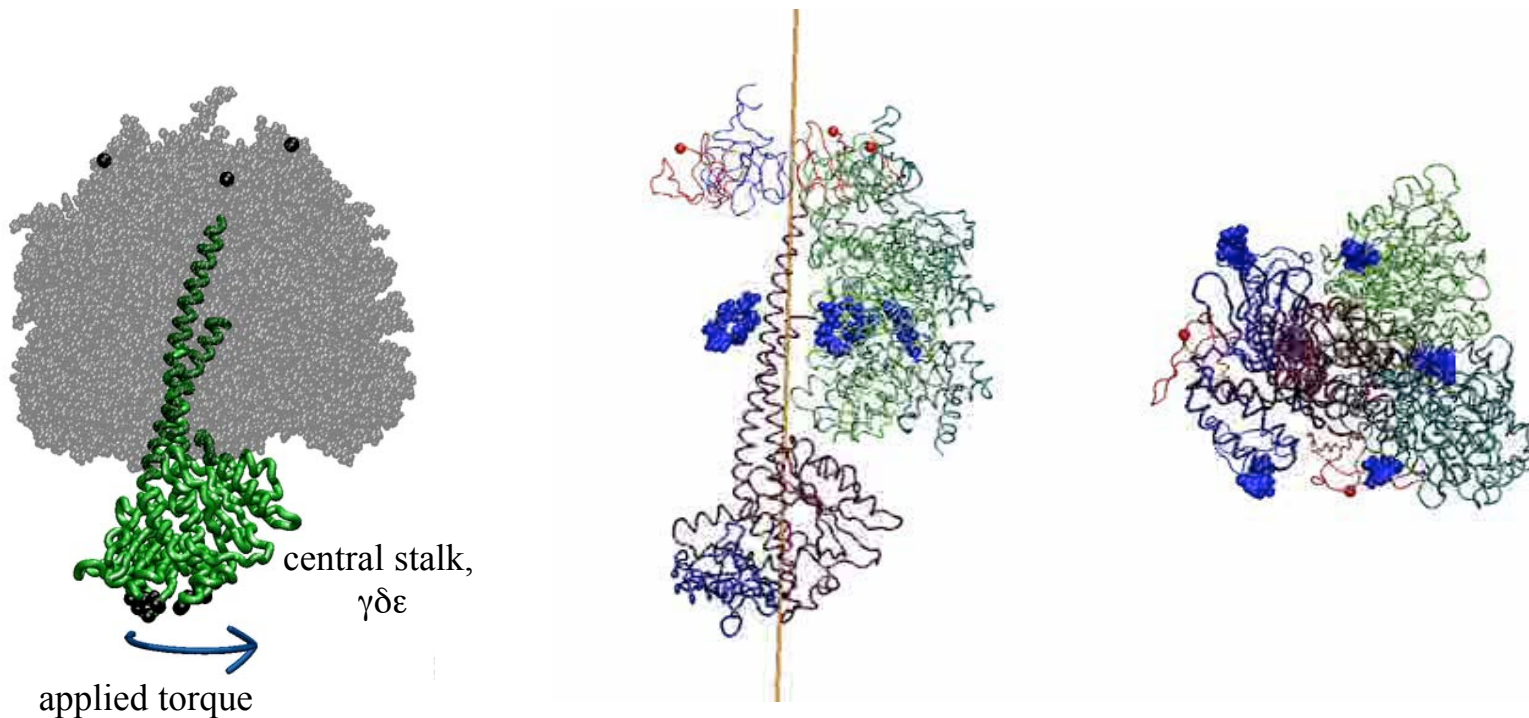
Assembling ATP Synthase F₁



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- Total **327,000 atoms** (3325 residues, 92,000 water molecules, nucleotides, and ions).
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Torque application to F_1

Torque is applied to the central stalk atoms at the F_1 - F_0 interface to constrain their rotation to constant angular velocity $\omega = 24$ deg/ns.

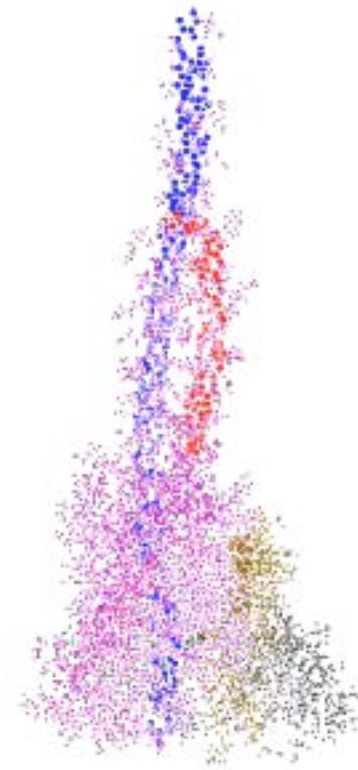
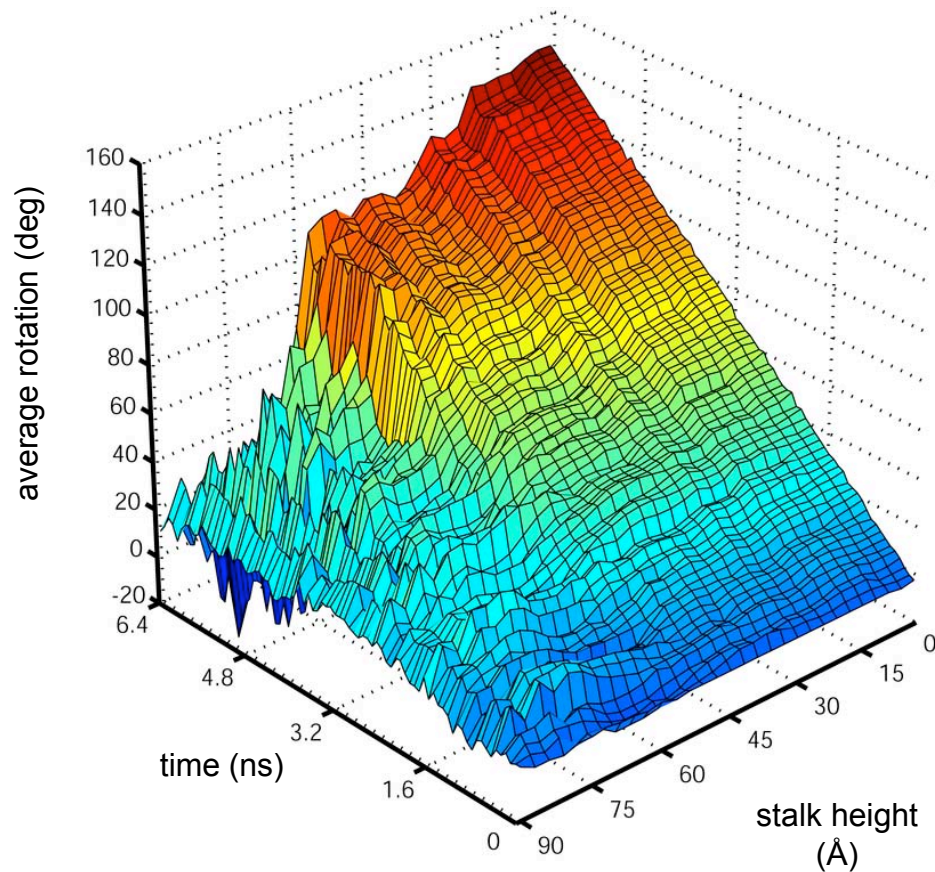
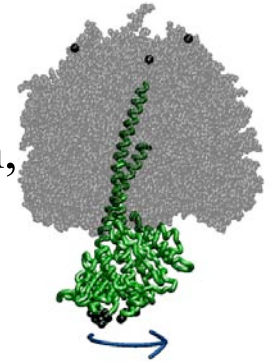


0.0 to 5.0 ns (0 to 120 deg) of torqued F_1 rotation, $\omega = 24$ deg/ns.

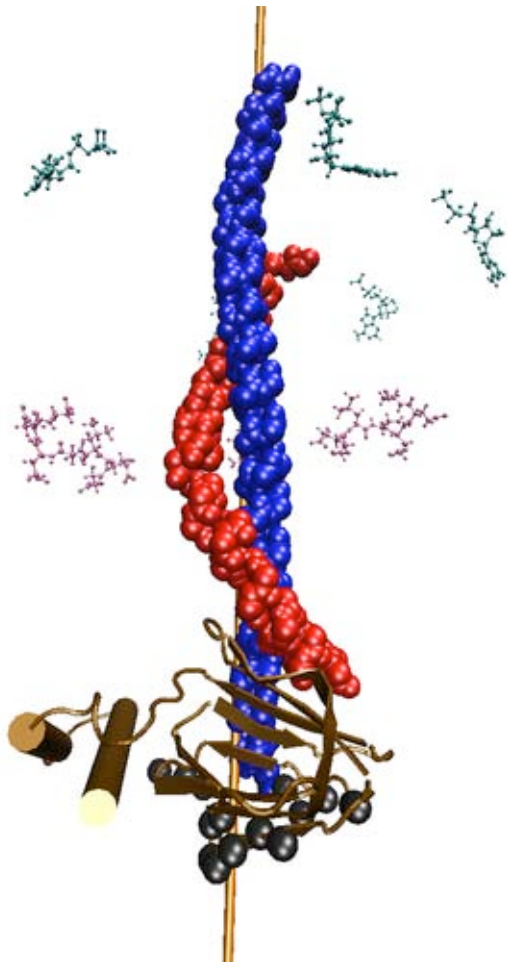
Stalk analysis

Using best RMSD rotation fit for stalk sections binned along axis direction, at 3.0 ns (72 deg) of rotation, we observe:

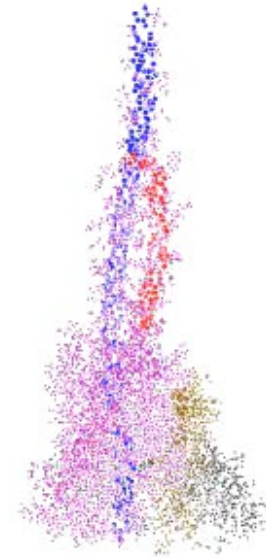
- slowed torque transmission along central stalk



Winding of γ coiled-coil



$t = 3.0$ ns
 $\theta = 72^\circ$

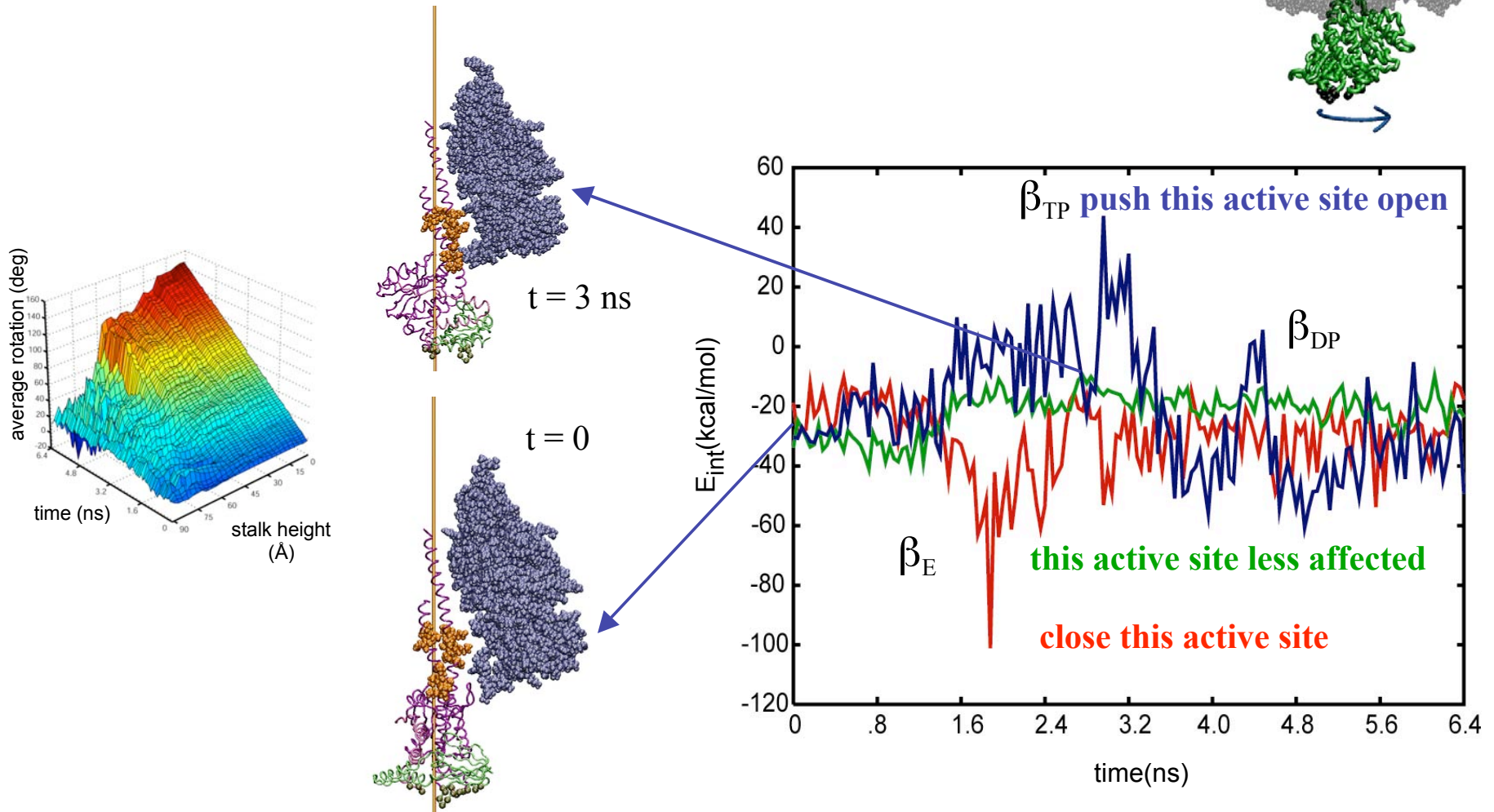
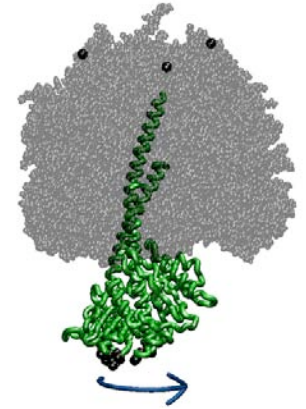


Different coupling for the two γ helices:
1—50, partially via δ subunit
197—272, directly to F_o

Rotation Produces Synthesis-like Events (1)

Around 3 ns (72 deg) of rotation, we observe:

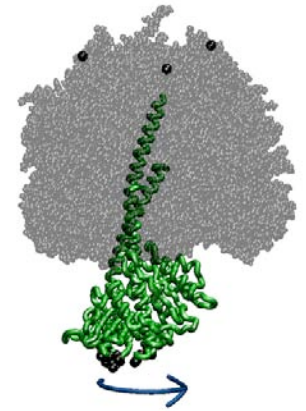
- slowed torque transmission along central stalk
- cooperative interactions at stalk - β subunit interfaces



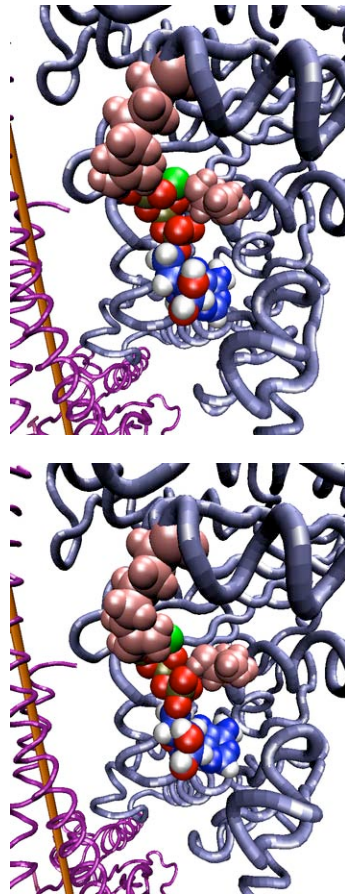
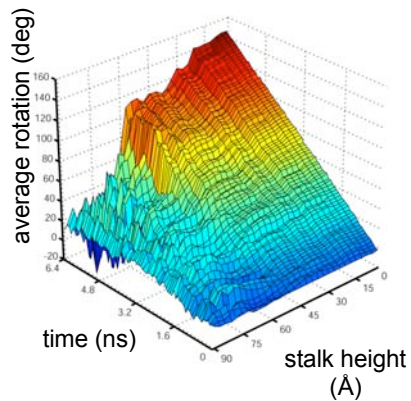
Rotation Produces Synthesis-like Events (3)

At 3.0 ns (72 deg) of rotation, we observe:

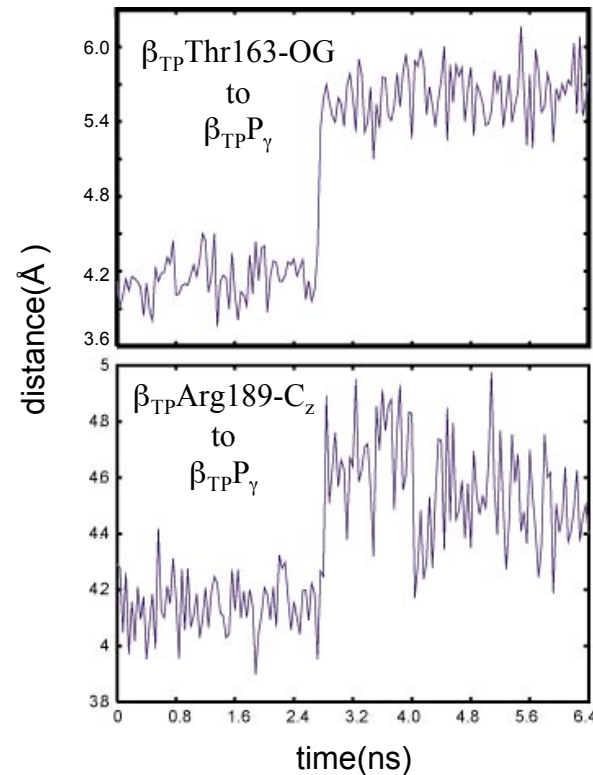
- slowed torque transmission along central stalk
- unbinding from ATP at the β_{TP} catalytic site



0 ns: active site closed

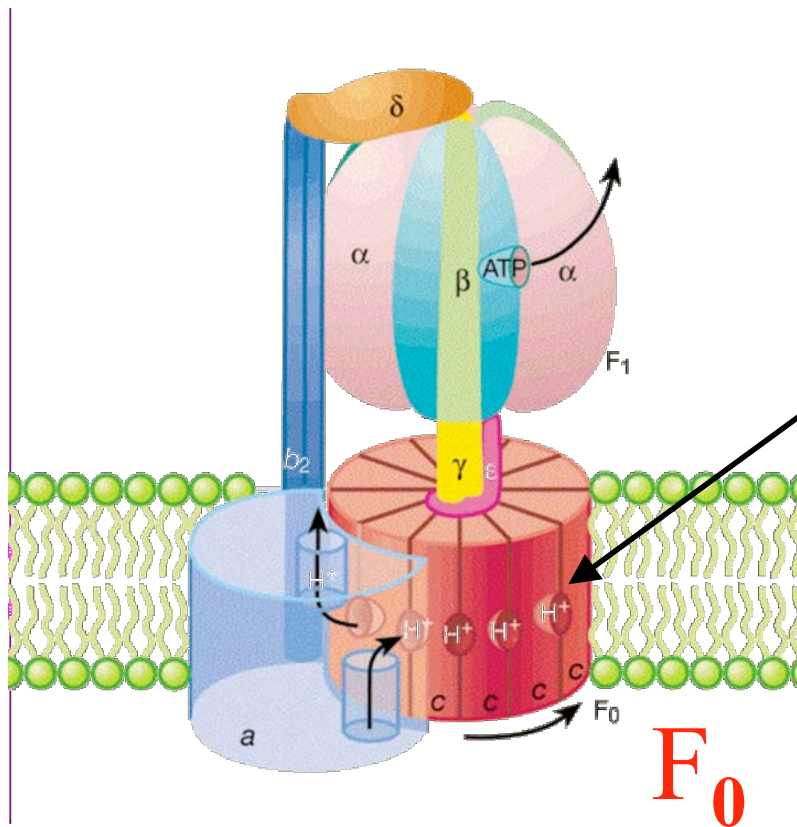


ATP separates from active site residues



3 ns: active site open

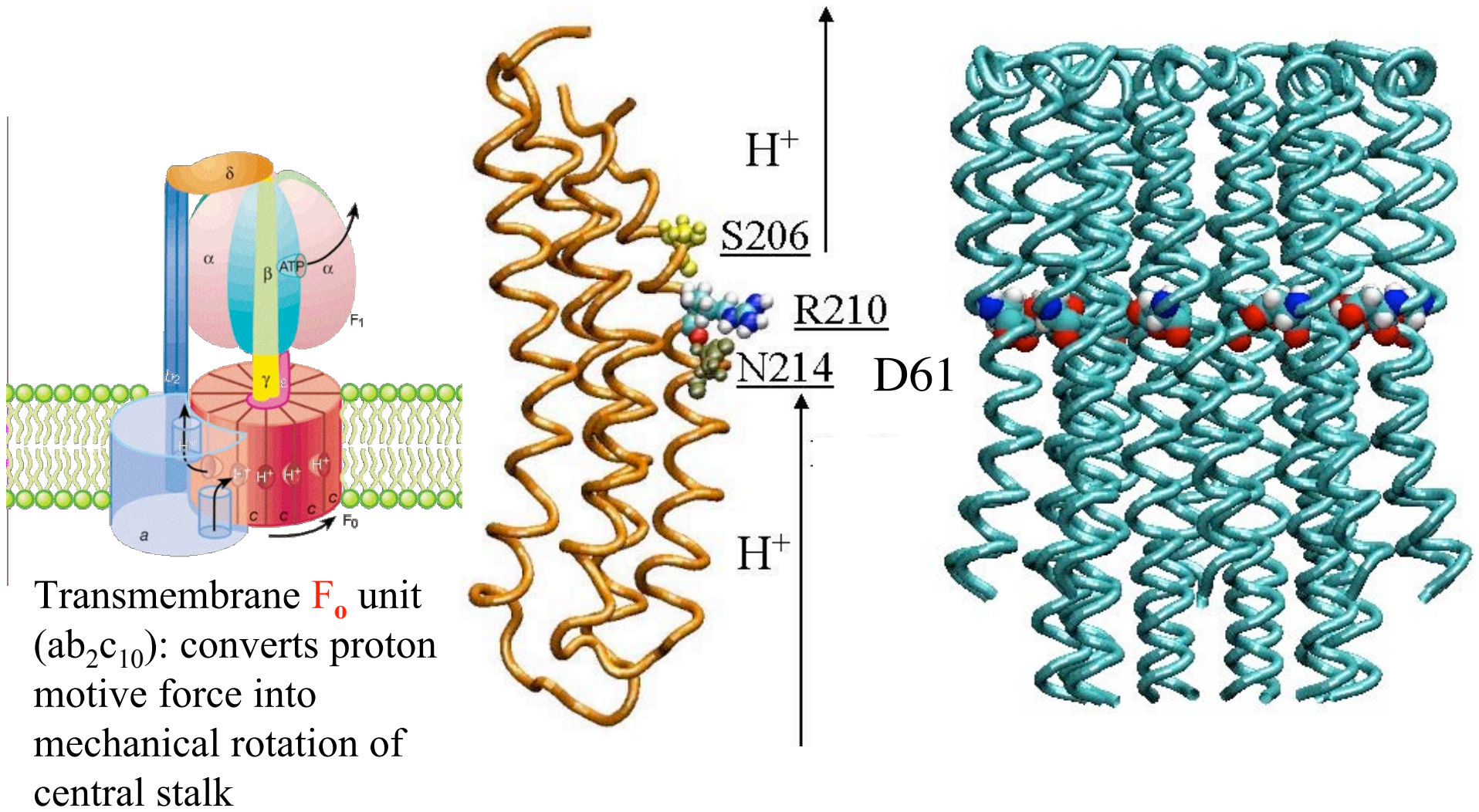
Let's Look at Fo ATP Motor



Asp 61 (D61) side groups take protons

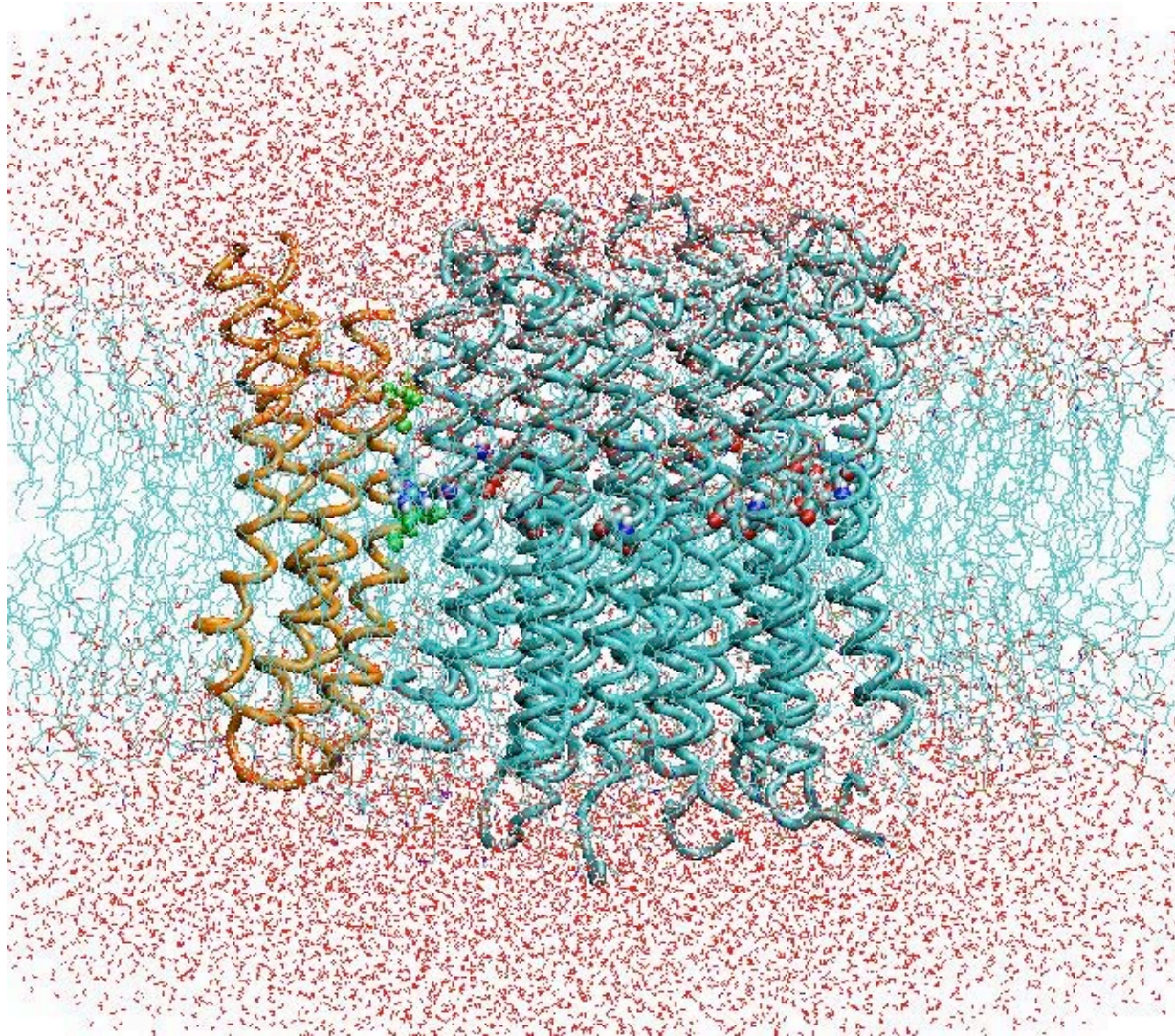
Transmembrane **F₀** unit (ab_2c_{10}):
converts proton motive force into
mechanical rotation of central stalk

Key Amino Acids Participating in Electro-Mechanical Motor



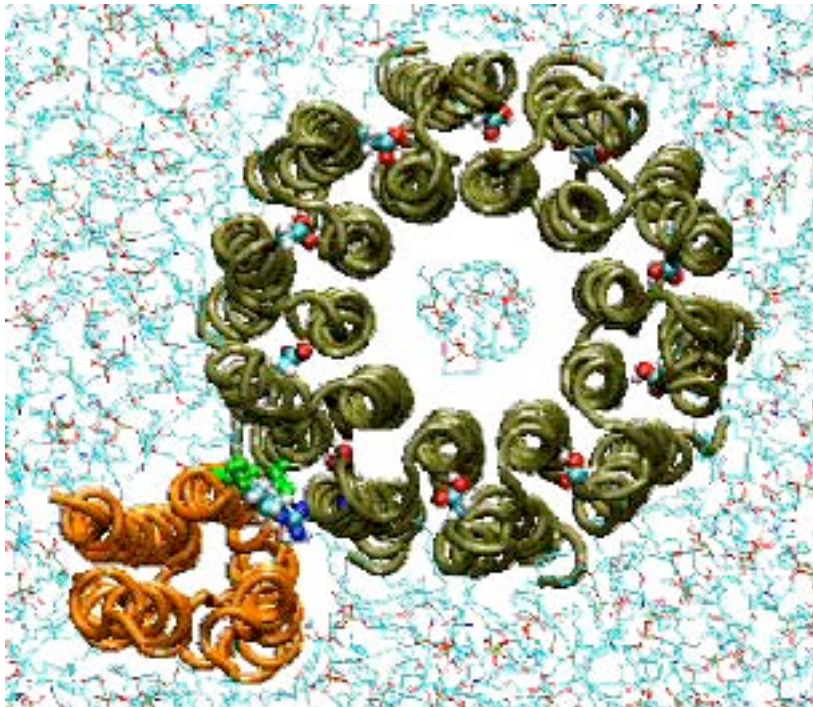
A. Aksimentiev, I. Balabin, R. Fillingame, K. Schulten, *Biophys. J.* 86: 1332-1344 (2004)

System Simulated

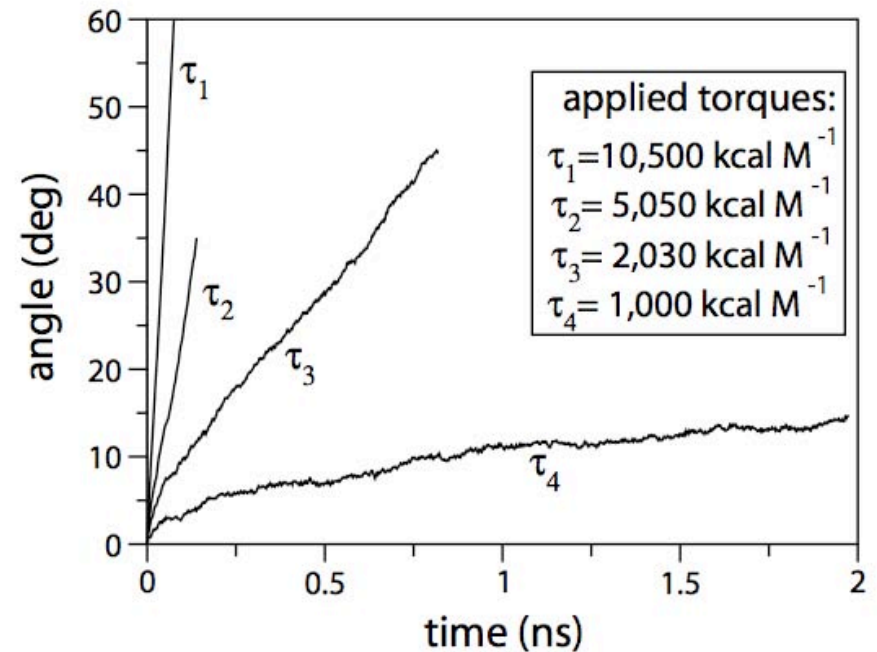


A. Aksimentiev, I. Balabin, R. Fillingame, K. Schulten, *Biophys. J.* (in press)

Forced Rotation of the c10 Subunit



Forces were applied to all backbone atoms of c₁₀



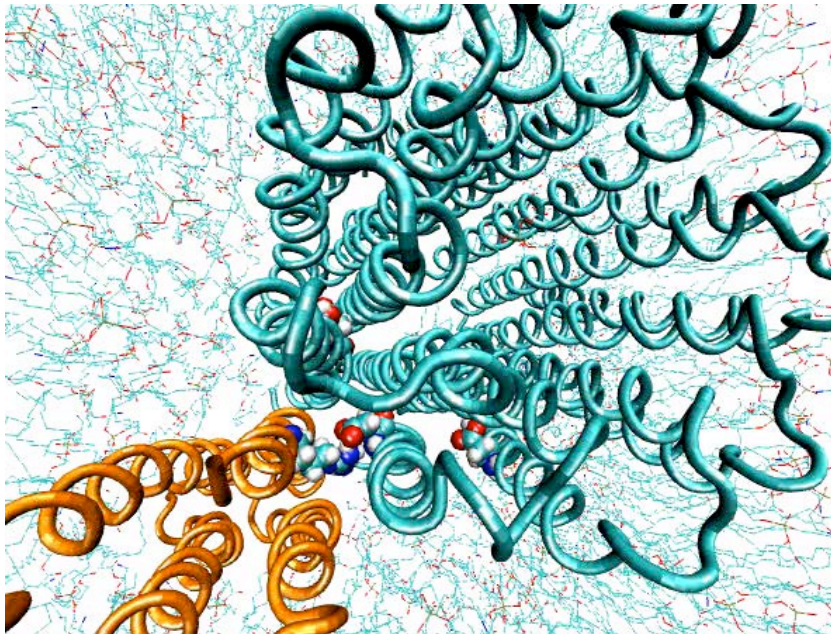
Estimated friction coefficient
 $\zeta \sim 10^5$ kcal/(M sec)

A. Aksimentiev, I. Balabin, R. Fillingame, K. Schulten, *Biophys. J.* (in press)

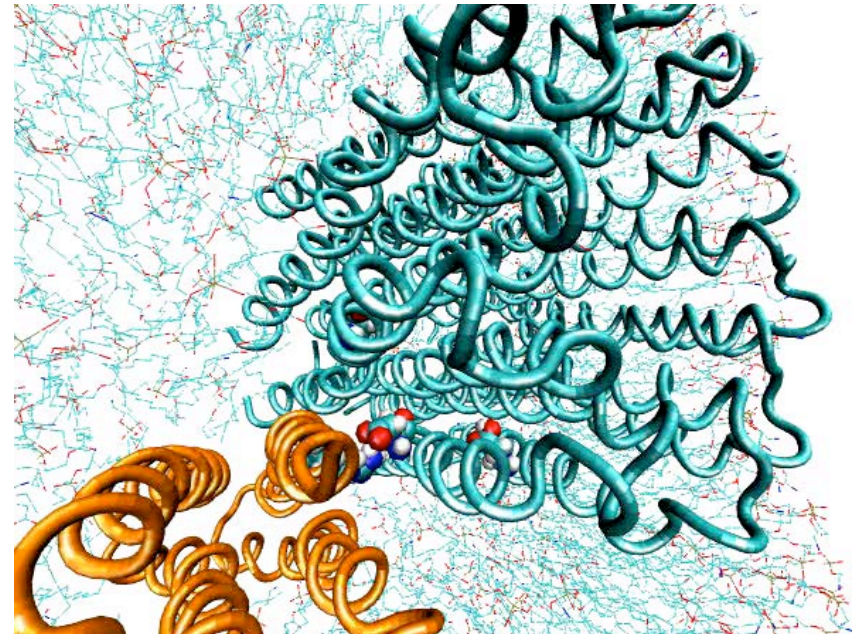
Salt Bridge Arg₂₁₀-Asp₆₁ is Formed

With only one Asp₆₁ residue deprotonated, SMD rotation of c_{10} breaks the structure apart.

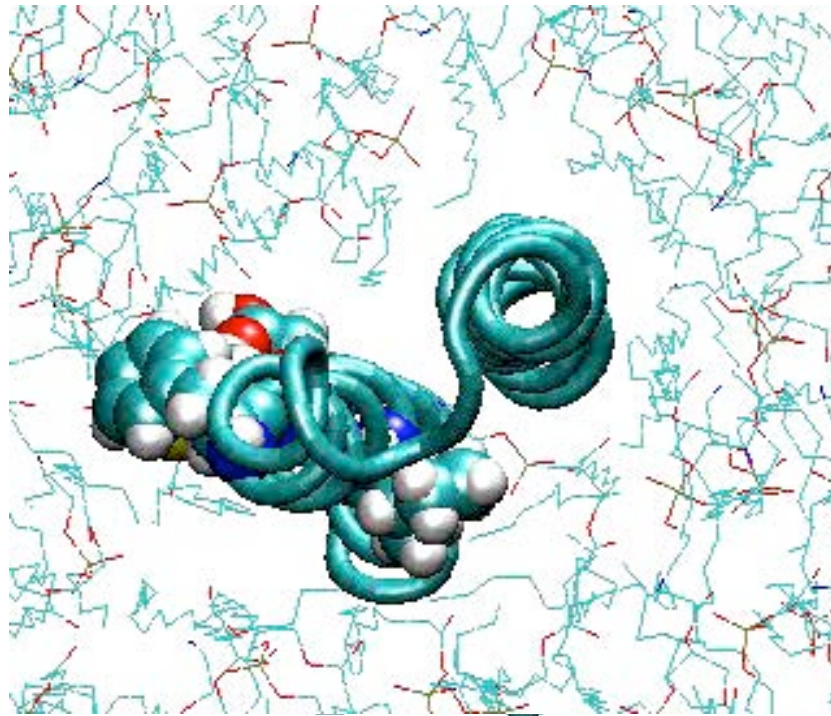
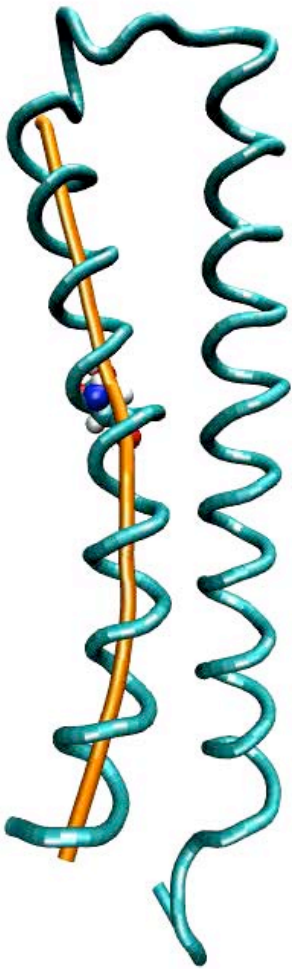
No restraints



Subunit α is restrained

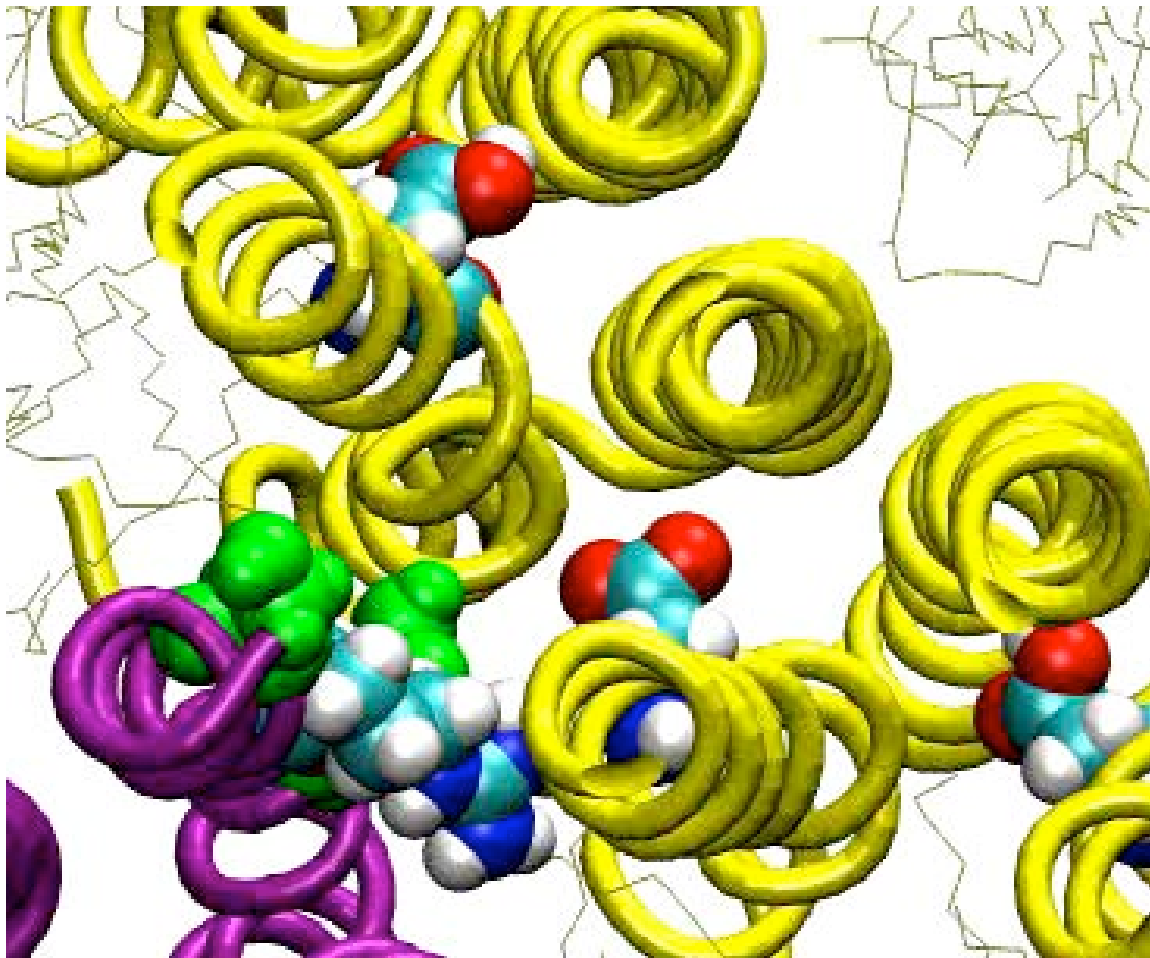


Single Helix Rotation is Feasible



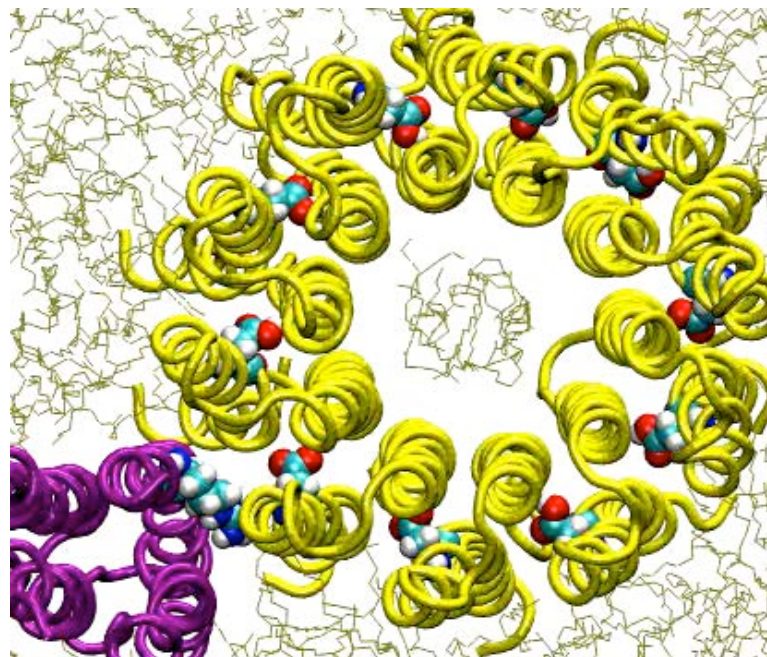
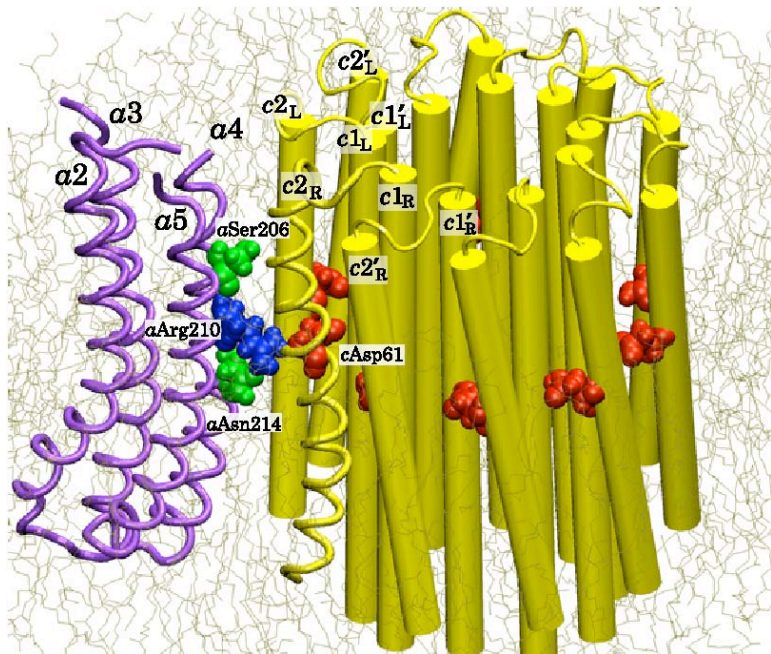
To minimize steric hindrance (critical on nanosecond time scale), helix was forced to rotate in a reptation tube (local pivot points and directors).

Salt Bridge Cannot be Broken, but Transferred



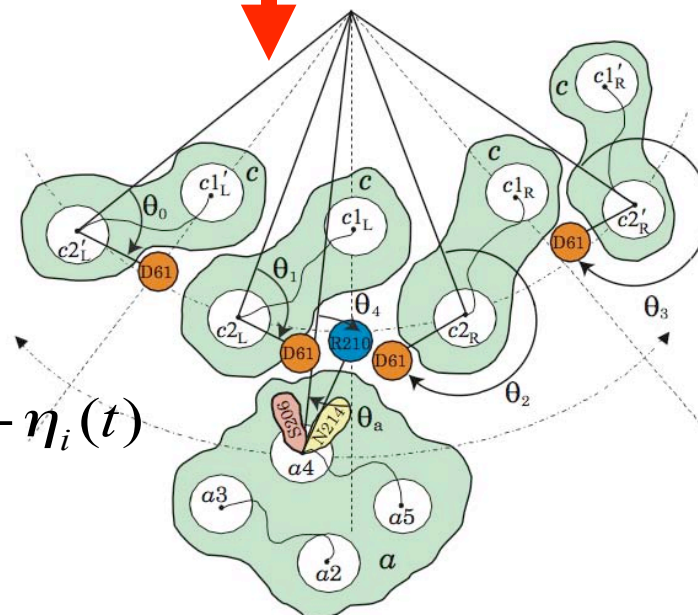
The salt bridge can be transferred by the concerted rotation of the c_{10} complex and the outer TMH of subunit c

Overall Mechanism: Theoretical Challenge

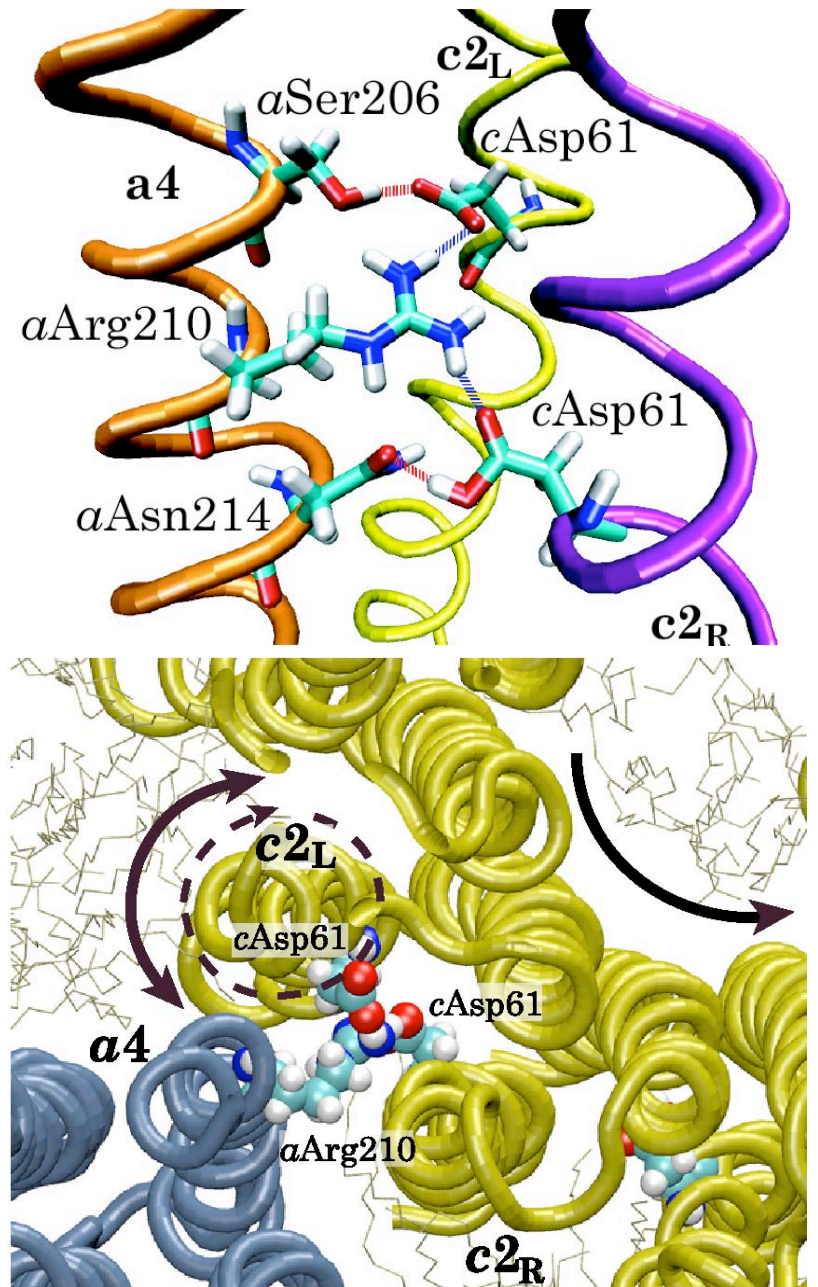
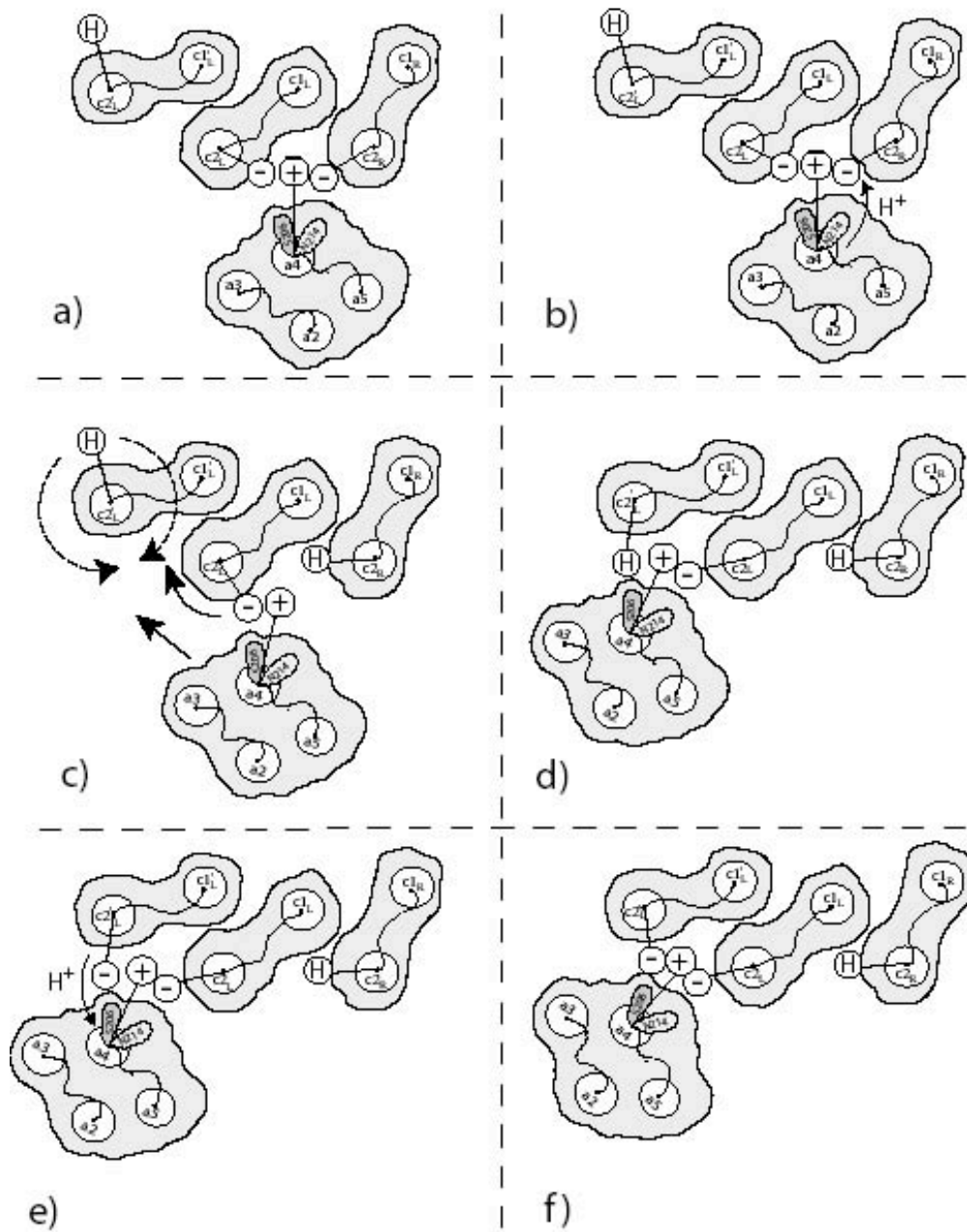


From ns simulation to ms model!

$$\xi_i \frac{d\theta_i}{dt} = - \frac{d}{d\theta_i} [U_{\text{group}} + U_{\text{hydroph.}} + U_{\text{internal}}] + \eta_i(t)$$

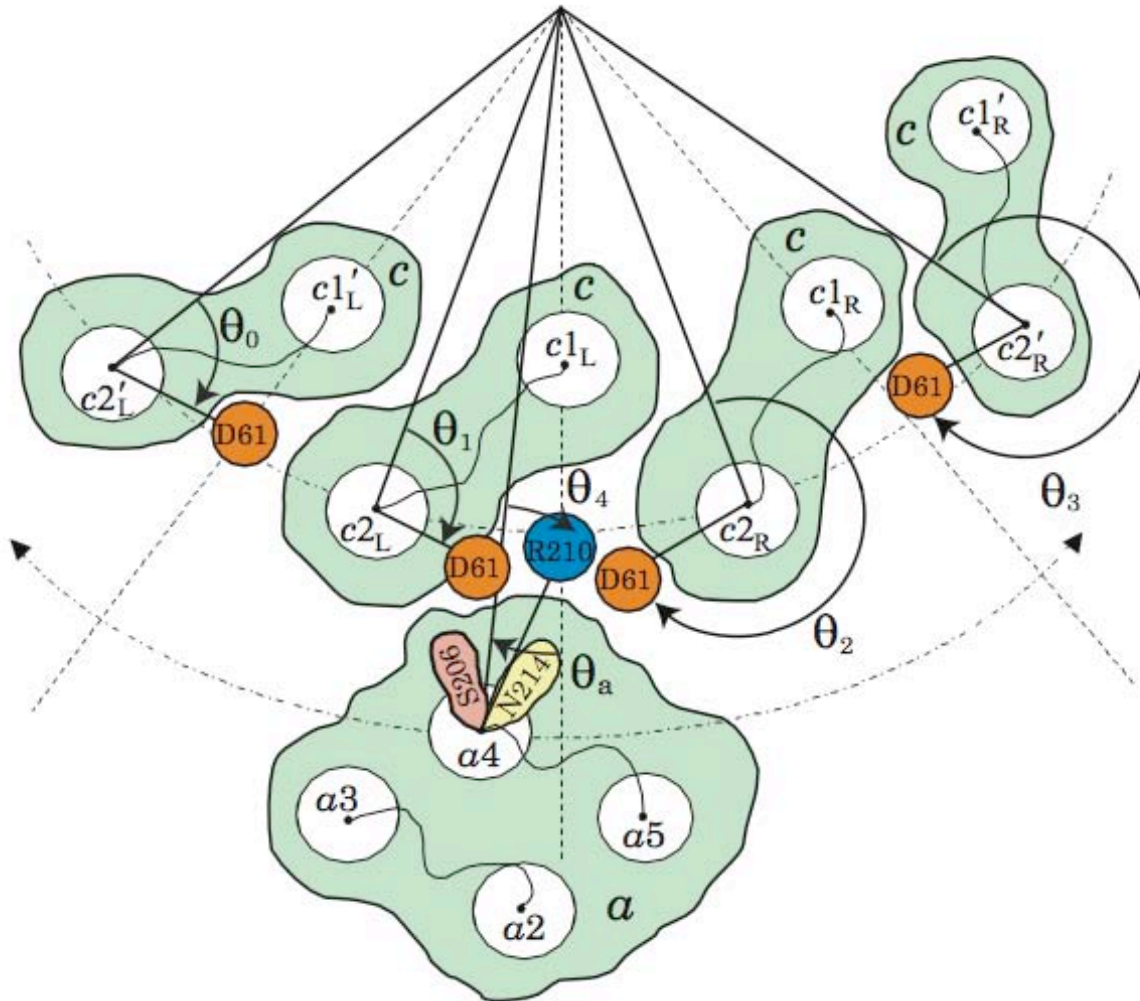


Key Steps in the Mechanism of the Fo Motor



Stochastic Model

Extends Simulation to ms Time Scale



6 degrees of freedom:

$\theta_0, \theta_1, \theta_2, \theta_3, \theta_4$ are

TMH rotation

angles; θ_A - position

of the a subunit.

Each Asp61 can be in

either of two chemical

states (protonated or

deprotonated).

$$\xi_i \frac{d\theta_i}{dt} = - \frac{d}{d\theta_i} [U_{\text{group}} + U_{\text{hydroph.}} + U_{\text{internal}}] + \eta_i(t)$$

Stochastic Simulations of F_0 Operation

Time evolution of rotation angles θ_1 (black), θ_2 (red), θ_4 (green), and θ_A (blue). Motor rotation speed is close to physiological.

